

Programming & Data Analysis with 'R'

Diarmuid O'Briain



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Document Outline

This document brings together a collection of different R experiences. It documents work carried out during a week long training on "*Statistical Data Analysis using R-software*" that took place from 24 - 28 September 2018 at the Directorate of Research and Graduate Training (DRGT), Makerere University, Kampala, Uganda that was delivered by Associate Professor Matthew Low and Dr. Matthew Hiron of the Swedish University of Agricultural Sciences (SLU) to develop the programming aspect of R with a focus on the analysis of quantitative data sets. I have included a section on qualitative data analysis as R has some tools in that area too.

The document explores the fundamentals of how to use *R*. Installing *R*, the structure of *R*, vectors, matrices, arrays, lists, tables, data-frames as well as flow control and user-defined functions. Working with quantitative data, linear models, predictions, probability distributions, distribution models and generalised linear mixed models. Plotting data for visual output. Finally both inductive and deductive approaches to the analysis of qualitative data is explored.

Diarmuid Ó Briain

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Table of Abbreviations

AIC	Akaike Information Criterion	
ANCOVA	Analysis of covariance	
ANOVA	Analysis of Variance	
CO ₂	Oxygen	
CRAN	Comprehensive R Archive Network	
CSS	Cascading Style Sheets	
CSV	Comma Separated file	
DRGT	Directorate of Research and Graduate Training	
DV	Dependent Variable	
FUN	Function	
GLM	Generalised Linear Model	
GLMM	Generalised Linear Mixed Model	
GUI	Graphical User Interface	
HTML	Hypertext Markup Language	
IV	Independent Variable	
LMM	Linear Mixed-effects Model	
MANOVA	Multivarite ANOVA	
NLME	Non-linear Mixed-Effects Model	
p-test	Statistical method used to assess one or more hypotheses within a population or a proportion within a population	
R	Open-source free statistical programming language used by scientists	
REML	Restricted (or Residual, or Reduced) Maximum Likelihood	
RQDA	R Qualitative Data Analysis	
SD	Standard Deviation	
SE	Standard Errors	
SLU	Swedish University of Agricultural Sciences	
SQL	Structured Query Language	
Student t-test	t-test introduced by William Sealy Gosset	
SVG	Scalable Vector Graphics	
t-test	Analysis of two populations means through the use of statistical examination	
Tukey	Tukey's Honest Significant Difference	
Welch t-test	Adaptation of Student's t-test	
YAML	YAML Ain't Markup Language	

1. Installing R

This document is based on implementation on the GNU/Linux operating system. It assumes a Debian GNU/Linux based distribution like Debian, Ubuntu, Linux Mint, Elementary OS or a host of others. If the GNU/Linux operating system is RedHat or Fedora based like CentOS then the *yum* package manager is used. Replace the *sudo apt-get install <package name>* with *sudo yum install <package name>*. For other operating systems like <u>Apple macOS</u> or <u>https://cloud.r-project.org/bin/windows/</u><u>Microsoft</u> please consult the relevant r-project links.

R can be used directly from the terminal shell and text editor tools like *xed* and *kate* have built in highlight modes, typically under the *Scientific* sections of these programs. Alternatively install an Integrated Development Environment (IDE) like *Rstudio*.

To operate with the terminal shell and text editors install the following base package.

```
$ sudo apt-get install r-base
```

If *Rstudio* is required the also install the following packages.

```
$ sudo apt-get install rstudio
$ sudo apt-get install r-cran-rstudioapi
```

This document will proceed with the assumption of a terminal shell and a text editor.

2. Introduction

2.1 Simple R script

Starting out with a simple R script and redirect it into the R interpreter. Copy from *cat* to *EOM* on its own and paste to a terminal shell, best to do this in a directory specifically for the purpose. This creates a file called *HelloWorld*.R in the working directory. Redirect the file to the R interpreter.

```
$ cat << EOM >> HelloWorld.R
## run this code in R
## Hello World script
plot(x=1, y=1, typ="n", xlab="", ylab="", bty="n", xaxt="n", yaxt="n", ylim=c(-1,2))
text(x=1, y=1,"'R' is a language for statistical analysis", cex=2, col="red")
text(x=1, y=0.5, "R: Hello World", cex=3, col="blue")
EOM
```

\$ R < test.R</pre>

A file called *Rplots.pdf* is generated in the working directory.

'R' is a language for statistical analysis



Illustration 1: R: Hello World

2.2 Another simple script

Some basic maths in another script.

```
$ cat << EOM >> Introduction.R
# R introduction
1+3
4*7
a=3
a+7
EOM
$ R < Introduction.R --vanilla</pre>
R version 3.4.4 (2018-03-15) -- "Someone to Lean On"
Copyright (C) 2018 The R Foundation for Statistical Computing
Platform: x86_64-pc-linux-gnu (64-bit)
R is free software and comes with ABSOLUTELY NO WARRANTY.
You are welcome to redistribute it under certain conditions.
Type 'license()' or 'licence()' for distribution details.
  Natural language support but running in an English locale
R is a collaborative project with many contributors.
Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.
Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.
> # R introduction
> 1+3
[1] 4
> 4*7
[1] 28
> a=3
> a+7
[1] 10
```

2.3 Start a new script

In both of these cases the script was redirected to the R interpreter. It is also possible to make the script executable and allow it find the interpreter by the inclusion of a shebang (#!) line on the first line referring the operating system to the to the path of the R interpreter. Make the file executable and run it.

```
$ cat << EOM >> vi my_first_script.R
#!/usr/bin/Rscript
# R introduction
1 + 3
4 * 7
a = 3
a + 7
EOM
$ chmod +x my_first_script.R
$ ./my_first_script.R
[1] 4
[1] 28
[1] 10
```

2.4 Running an R shell

An R shell can be ran to execute a small number of commands. Many of the examples in this document are executed in this way.

```
$ R
R version 3.4.4 (2018-03-15) -- "Someone to Lean On"
Copyright (C) 2018 The R Foundation for Statistical Computing
Platform: x86_64-pc-linux-gnu (64-bit)
R is free software and comes with ABSOLUTELY NO WARRANTY.
You are welcome to redistribute it under certain conditions.
Type 'license()' or 'licence()' for distribution details.
Natural language support but running in an English locale
R is a collaborative project with many contributors.
Type 'contributors()' for more information and
'citation()' on how to cite R or R packages in publications.
Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.
>
```

To run *R* without the message each time add the *--quiet* option.

```
$ R --quiet
>
```

2.5 R errors

Consider the example below. As *b* has not been defined *R* has no way of carrying out the instruction and will therefore throw back an error as demonstrated.

```
$ R --quiet
> b + 7
Error: object 'b' not found
```

2.6 Commenting

Annotate code as you write, it makes it easier to read later. The annotations are preceded with a **#** symbol. The *Rscript* interpreter will ignore the annotations.

```
$ cat << EOM >> vi my_first_annotate.R
#!/usr/bin/Rscript
a = list(c(1, 2, 3, 4)),
                         # A vector
         "Some",
                        # Add characters
         "Words",
                         # Add a second set of characters
         c(5,6,7,8)
                         # A second vector
        )
## Exercise
str (a)
                         # Output the structure of 'a'
m = lapply(a[1], mean)  # Apply the mean function to 1st list in 'a'
n = lapply(a[4], max)  # Apply the max function to 4st list in 'a'
a$means = c(m, n)
                         # Add a new item to the list
print(a)
EOM
$ chmod +x my_first_annotate.R
```

```
$ ./my first annotate.R
List of 4
$ : num [1:4] 1 2 3 4
$ : chr "Some"
$ : chr "Words"
$ : num [1:4] 5 6 7 8
[[1]]
[1] 1 2 3 4
[[2]]
[1] "Some"
[[3]]
[1] "Words"
[[4]]
[1] 5 6 7 8
$means
$means[[1]]
[1] 2.5
$means[[2]]
[1] 8
```

2.6.1 Multiline commenting

Many programming languages offer the ability to do multiline commenting. This is useful if there are some lines in the script that you want the script to skip without the need to put *#* before each line in the block. In this case use the structure below around the block. In this case the test is always FALSE and therefore the lines are skipped.

```
if (FALSE){
Lines to be ignored.
}
```

To demonstrate. Watch what happens if this structure is added to the script. The lines between 'if(FALSE){' and '}' are bypassed by the R interpreter.

```
$ cat << EOM >> multiline_comment.R
#!/usr/bin/Rscript
a = list(c(1,2,3,4),
                        # A vector
         "Some",
                        # Add characters
        "Words",
                        # Add a second set of characters
        c(5,6,7,8)
                       # A second vector
       )
## Exercise
str (a)
                         # Output the structure of 'a'
if(FALSE){
m = lapply(a[1], mean)  # Apply the mean function to 1st list in 'a'
n = lapply(a[4], max)
                       # Apply the max function to 4st list in 'a'
asmeans = c(m, n)
                        # Add a new item to the list
}
print(a)
EOM
```

```
$ R < my_first_annotate.R
List of 4
$ : num [1:4] 1 2 3 4
$ : chr "Some"
$ : chr "Words"
$ : num [1:4] 5 6 7 8
[[1]]
[1] 1 2 3 4
[[2]]
[1] "Some"
[[3]]
[1] "Words"
[[4]]
[1] 5 6 7 8
```

So what happened?

2.7 Listing the existing objects in R

Is(): Returns a vector of character strings giving the names of the objects in the specified environment.

objects(): Same as *ls()*.

```
> ls()
[1] "df" "model1" "model2" "model3" "model4" "realmodel"
[7] "v" "v1" "v2" "v3" "v4" "x"
```

2.7.1 Clearing the existing objects in R

rm(): Removes objects specified successively as character strings, or in a character vector *list*, or through a combination of both.

remove(): Same as rm().

To clear the environment of objects, supply a *list* of objects to the *rm()* or *remove()* functions.

> ls() [1] "df" [7] "v"	"model1" "v1"	"model2" "v2"	"model3" "v3"	"model4" "v4"	"realmodel" "x"
<pre>> rm(list=ls())</pre>					
> ls() character(0)					
or alternatively:					
> objects() [1] "df" [7] "v"	"model1" "v1"	"model2" "v2"	"model3" "v3"	"model4" "v4"	"realmodel" "x"
<pre>> remove(list=ob</pre>	jects())				
<pre>> objects() character(0)</pre>					

3. R structure

rnorm(): random generation for the normal distribution.

rnorm(5,mean=0,sd=1)

Note: rnorm(n, mean = , sd =) is used to generate n normal random numbers with arguments mean and standard deviation.

```
> rnorm(5,mean=0,sd=1)
[1] 1.3136807 -1.2612950 0.9052489 0.8711972 -1.7449344
```

```
> plot(rnorm(5,mean=0,sd=1))
```



function(function(object))

Note: The *R abs()* method is one of the R mathematics functions, which is used to return the absolute positive value of an individual number, or an expression.

```
> abs(rnorm(5,mean=0,sd=1))
[1] 0.07609716 0.57743538 1.10299258 0.42479172 1.41277626
```

function(function(function(object)))

```
> mean(abs(rnorm(100,mean=0,sd=1)))
[1] 0.8621388
```

4. Getting help with functions

There are a number of helpful tools in *R* to get more information.

- **args():** Lists the arguments of a function give as an argument.
- help(), ?: Get the man page for the function given as an argument.
- **example():** Get examples of how the function given as an argument can be used.

4.1 Arguments of a function

To understand a new function and one needs the available arguments within the function.

function(arguments)

```
> args(rnorm)
function (n, mean = 0, sd = 1)
NULL
```

- n User specified, i.e. number of numbers
- **mean** By default the mean = 0
- sd Default standard deviation = 1

4.2 help()

> help() or
persp

package:graphics

R Documentation

Perspective Plots

Description:

This function draws perspective plots of a surface over the x-y plane. 'persp' is a generic function.

Usage:

persp(x, ...)

4.3 example()

Example function gives examples of the function given to it as an argument.

```
> example(mean)
mean> x <- c(0:10, 50)
mean> xm <- mean(x)
mean> c(xm, mean(x, trim = 0.10))
[1] 8.75 5.50
```

More information can be found online at:

quick-r

inside-r

r-graph gallery

5. R packages

5.1 The Comprehensive R Archive Network (CRAN)

The CRAN is a resource of packages that can be added to *R*.

The Comprehensive R Archive Network website

5.2 Installing Packages in R

To install packages an internet connection is necessary.

> install.packages("lme4")

Once the package is downloaded it is there but the package must be *loaded* to use it.

> library(lme4)

In a script that has a dependency on a package it is helpful to have a line at the top of the file that causes the package to be installed if the package is not already installed. In the example, if the package *lme4* is not installed then *R* will install it first before continuing with the script. If it is installed then the line is ignored. The *library('lme4')* line loads the library such that the functions within the package *lme4* become available within the program.

```
> if(!require(lme4)){install.packages("lme4")}
```

> library(lme4)

5.3 Check installed packages

Check packages already installed with the *ip* command.

> ip

	Package	LibPath	Version	Priority
base	"base"	"/usr/lib/R/library"	"3.4.4"	"base"
boot	"boot"	"/usr/lib/R/library"	"1.3-20"	"recommended"
class	"class"	"/usr/lib/R/library"	"7.3-14"	"recommended"
cluster	"cluster"	"/usr/lib/R/library"	"2.0.6"	"recommended"
codetools	"codetools"	"/usr/lib/R/library"	"0.2-15"	"recommended"
compiler	"compiler"	"/usr/lib/R/library"	"3.4.4"	"base"
datasets	"datasets"	"/usr/lib/R/library"	"3.4.4"	"base"
foreign	"foreign"	"/usr/lib/R/library"	"0.8-70"	"recommended"
graphics	"graphics"	"/usr/lib/R/library"	"3.4.4"	"base"
grDevices	"grDevices"	"/usr/lib/R/library"	"3.4.4"	"base"
grid	"grid"	"/usr/lib/R/library"	"3.4.4"	"base"
KernSmooth	"KernSmooth"	"/usr/lib/R/library"	"2.23-15"	"recommended"
lattice	"lattice"	"/usr/lib/R/library"	"0.20-35"	"recommended"
MASS	"MASS"	"/usr/lib/R/library"	"7.3-49"	"recommended"
Matrix	"Matrix"	"/usr/lib/R/library"	"1.2-12"	"recommended"
methods	"methods"	"/usr/lib/R/library"	"3.4.4"	"base"
mgcv	"mgcv"	"/usr/lib/R/library"	"1.8-23"	"recommended"
nlme	"nlme"	"/usr/lib/R/library"	"3.1-131"	"recommended"
nnet	"nnet"	"/usr/lib/R/library"	"7.3-12"	"recommended"
parallel	"parallel"	"/usr/lib/R/library"	"3.4.4"	"base"
rpart	"rpart"	"/usr/lib/R/library"	"4.1-13"	"recommended"
spatial	"spatial"	"/usr/lib/R/library"	"7.3-11"	"recommended"
splines	"splines"	"/usr/lib/R/library"	"3.4.4"	"base"
stats	"stats"	"/usr/lib/R/library"	"3.4.4"	"base"
stats4	"stats4"	"/usr/lib/R/library"	"3.4.4"	"base"
survival	"survival"	"/usr/lib/R/library"	"2.41-3"	"recommended"
tcltk	"tcltk"	"/usr/lib/R/library"	"3.4.4"	"base"

"tools" "/usr/lib/R/library" "3.4.4" "base" tools "/usr/lib/R/library" "3.4.4" utils "utils" "base" Depends base NA boot "R (>= 3.0.0), graphics, stats" "R (>= 3.0.0), stats, utils" class "R (>= 3.0.1)" cluster "R (>= 2.1)" codetools compiler NA datasets NΑ foreign "R (>= 3.0.0)" graphics NA grDevices NA grid NA KernSmooth "R (>= 2.5.0), stats" "R (>= 3.0.0)" lattice "R (>= 3.1.0), grDevices, graphics, stats, utils" MASS Matrix "R (>= 3.0.1)" methods NA "R (>= 2.14.0), nlme (>= 3.1-64)" mgcv nlme "R (>= 3.0.2)" "R (>= 2.14.0), stats, utils" nnet parallel NΑ "R (>= 2.15.0), graphics, stats, grDevices" rpart spatial "R (>= 3.0.0), graphics, stats, utils" splines NA stats NA stats4 NA survival "R (>= 2.13.0)" tcltk NA tools NA utils NA Imports LinkingTo base NA NA boot NA NA class "MASS" NA "graphics, grDevices, stats, utils" cluster NA codetools NA NA compiler NA NA datasets NA NA "methods, utils, stats" foreian NA "grDevices" graphics NA grDevices NA NA "grDevices, utils" grid NA KernSmooth NA NA lattice "grid, grDevices, graphics, stats, utils" NA "methods" MASS NA "methods, graphics, grid, stats, utils, lattice" Matrix NA methods "utils, stats" NA "methods, stats, graphics, Matrix" mgcv NA "graphics, stats, utils, lattice" nlme NA nnet NA NA parallel "tools, compiler" NA rpart NA NA spatial NA NA splines "graphics, stats" NA stats "utils, grDevices, graphics" NA stats4 "graphics, methods, stats" NA survival "graphics, Matrix, methods, splines, stats, utils" NA "utils" tcltk NA tools NΑ NA utils NA NA Suggests "methods" base boot "MASS, survival" class NA "MASS" cluster

codetools NA compiler NA datasets NΑ foreian NA graphics NA grDevices "KernSmooth" "lattice" grid KernSmooth "MASS" lattice "KernSmooth, MASS, latticeExtra" "lattice, nlme, nnet, survival" MASS Matrix "expm, MASS" methods "codetools" "splines, parallel, survival, MASS" mgcv "Hmisc, MASS" nlme "MASS" nnet "methods" parallel "survival" rpart "MASS" spatial "Matrix, methods" splines "MASS, Matrix, SuppDists, methods, stats4" stats stats4 NA survival NA tcltk NΑ "codetools, methods, xml2, curl" tools utils "methods, XML" Enhances License "Part of R 3.4.4" base NA boot NA "Unlimited" class "GPL-2 | GPL-3" NA "GPL (>= 2)" cluster NA codetools "GPL" NA "Part of R 3.4.4" compiler NA "Part of R 3.4.4" datasets NA foreign "GPL (>= 2)" NA "Part of R 3.4.4" graphics NA grDevices "Part of R 3.4.4" NA "Part of R 3.4.4" NΑ grid KernSmooth NA "Unlimited" lattice "chron" "GPL (>= 2)" "GPL-2 | GPL-3" MASS NA "MatrixModels, graph, SparseM, sfsmisc" "GPL (>= 2) | file LICENCE" Matrix "Part of R 3.4.4" methods NA mgcv NA "GPL (>= 2)" nlme "GPL (>= 2) | file LICENCE" NΔ nnet NA "GPL-2 | GPL-3" parallel "snow, nws, Rmpi" "Part of R 3.4.4" "GPL-2 | GPL-3" rpart NA "GPL-2 | GPL-3" spatial NA splines "Part of R 3.4.4" NA "Part of R 3.4.4" stats NA stats4 "Part of R 3.4.4" NA "LGPL (>= 2)" survival NA tcltk NA "Part of R 3.4.4" "Part of R 3.4.4" tools NA utils NA "Part of R 3.4.4" License_is_FOSS License_restricts_use OS_type MD5sum base NA NA NA NA NA boot NA NA NA class NA NA NA NA cluster NA NA NA NA codetools NΑ NA NΑ NΑ compiler NA NA NA NA datasets NA NA NA NA foreign NA NA NΔ NA graphics NA NA NA NA grDevices NA NA NA NA grid NA NA NA NA

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NA

NA

NA

NA NA

NA

NA

KernSmooth	NA	NA
lattice	NA	NA
MASS	NA	NA
Matrix	NA	NA
methods	NA	NA
mgcv	NA	NA
nlme	NA	NA
nnet	NA	NA
parallel	NA	NA
rpart	NA	NA
spatial	NA	NA
splines	NA	NA
stats	NA	NA
stats4	NA	NA
survival	NA	NA
tcltk	NA	NA
tools	NA	NA
utils	NA	NA
	NeedsCompilation	Built
base	NA	"3.4.4"
boot	"no"	"3.4.2"
class	"yes"	"3.4.2"
cluster	"ves"	"3.4.2"
codetools	"no"	"3.4.2"
compiler	NA	"3.4.4"
datasets	NA	"3.4.4"
foreign	"yes"	"3.4.4"
graphics	"ves"	"3.4.4"
grDevices	"ves"	"3.4.4"
grid	"ves"	"3.4.4"
KernSmooth	"ves"	"3.4.2"
lattice	"yes"	"3.4.2"
MASS	"ves"	"3.4.3"
Matrix	"ves"	"3.4.2"
methods	"yes"	"3.4.4"
macv	"ves"	"3.4.3"
nÎme	"ves"	"3.4.2"
nnet	"ves"	"3.4.2"
parallel	"ves"	"3.4.4"
rpart	"ves"	"3.4.3"
spatial	"ves"	"3.4.2"
splines	"ves"	"3.4.4"
stats	"ves"	"3.4.4"
stats4	NA	"3.4.4"
survival	"ves"	"3.4.2"
tcltk	"ves"	"3.4.4"
tools	"ves"	"3.4.4"
utils	"ves"	"3.4.4"
	,	.

6. Citing R in papers and publications

Credit where credit is due, when using *R* or any of its packages it is important to credit the work of the developers. *R* has a built-in function citation() to get the BibTex entry for reference management software or simply take the reference directly from the terminal as text.

6.1 Cite R

```
> citation()
R Core Team (2018). R: A language and environment for statistical
computing. R Foundation for Statistical Computing, Vienna, Austria.
URL https://www.R-project.org/.
A BibTeX entry for LaTeX users is
@Manual{,
    title = {R: A Language and Environment for Statistical Computing},
    author = {{R Core Team}},
    organization = {R Foundation for Statistical Computing},
    address = {Vienna, Austria},
    year = {2018},
    url = {https://www.R-project.org/},
  }
```

We have invested a lot of time and effort in creating R, please cite it when using it for data analysis. See also 'citation("pkgname")' for citing R packages.

6.2 Cite individual R packages

> citation(package = "lme4")

To cite lme4 in publications use:

```
Douglas Bates, Martin Maechler, Ben Bolker, Steve Walker (2015).
Fitting Linear Mixed-Effects Models Using lme4. Journal of
Statistical Software, 67(1), 1-48. doi:10.18637/jss.v067.i01.
```

```
A BibTeX entry for LaTeX users is
```

```
@Article{,
    title = {Fitting Linear Mixed-Effects Models Using {lme4}},
    author = {Douglas Bates and Martin M{\"a}chler and Ben Bolker and Steve Walker},
    journal = {Journal of Statistical Software},
    year = {2015},
    volume = {67},
    number = {1},
    pages = {1--48},
    doi = {10.18637/jss.v067.i01},
}
```

7. Vectors, the basic data structure in R

The *vector* is the basic data structure in R. It contains element of the same type. The data types can be logical, integer, double, character, complex or raw. It is considered the fundamental data type in R.

- logical TRUE and FALSE are reserved words denoting logical constants, whereas T and F are global variables whose initial values set to these.
- integer Whole number (not a fraction) that can be positive, negative, or zero.
- **double** Creates a double-precision vector of the specified length. The elements of the vector are all equal to 0. It is identical to numeric.
- character Type of indexing is useful when dealing with named vectors.
- complex The vector can be specified either by giving its length, its real and imaginary parts, or modulus and argument.
- raw Type is intended to hold raw bytes. It is possible to extract sub-sequences of bytes, and to replace elements.

```
> x = 3
> y = 5
> meatballs = 7
> meatballs + x - y
[1] 5
> 4 + 3
[1] 7
> a = 4 + 3
> a
[1] 7
```

The vector is the fundamental data type in *R*.

e.g. [1] 1 2 3 4 5 6 7

Numbers being put together in a vector must be done in one of these four ways.

<pre>> x = c(1,2,3,4,5,6,7)</pre>	# concatenate
> y = 1:7	# colon operator
<pre>> z = seq(1,7,1)</pre>	<pre># sequence, 1 to 7 in intervals of 2 i.e. 1 3 5 7</pre>
> m = rep(1:7,2)	# repeat i.e. 1,2,3,4,5,6,7,1,2,3,4,5,6,7

Now consider each.

```
> x = c(1,2,3,4,5,6,7)
> y = 1:7
> z = seq(1,7,1)
> m = rep(1:7,2)
> x
[1] 1 2 3 4 5 6 7
> y
[1] 1 2 3 4 5 6 7
> z
[1] 1 2 3 4 5 6 7
> m
[1] 1 2 3 4 5 6 7
```

Data Analytics

Try some variations.

```
> d = seq(1,12,3)
> d
[1] 1 4 7 10
> e = rep(1:4,3)
> e
[1] 1 2 3 4 1 2 3 4 1 2 3 4
> f = rep(d,4)
> f
[1] 1 4 7 10 1 4 7 10 1 4 7 10 1 4 7 10
```

Note:

٠	c():	- Combines or concatenates its arguments
•	<pre>seq(from, to, by=):</pre>	- Generate regular sequences.
٠	rep(x,):	- replicates the values in 'x'.

Combining these.

```
> x = 1:7
> y = seq(1,7,2)
> z = c(x,y)
> z
[1] 1 2 3 4 5 6 7 1 3 5 7
```

Any time more than one number must be given to *R*, then it MUST be created using one of these functions.

7.1 Exercise: Generate vectors

Answer:

```
5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23
> a = 5:23
1.0 1.3 1.6 1.9 2.2 2.5 2.8 3.1 3.4 3.7 4.0 4.3 4.6 4.9
> c = seq(1,4.9,0.3)
> c = rep(7, 18)
14 12 10 8 6 4 2 0
> c = seq(14, 0, -2)
5 7 6 10 4 3 17
> c = c(5,7,6,10,4,3,17)
1 2 3 4 1 2 3 4 1 2 3 4
> rep(1:4,3)
1 2 3 4 1 2 3 4 1 2 3 4 85
> c = c(rep(1:4,3), 85)
2.00 2.17 2.34 2.51 2.68 2.85 3.02 3.19 3.36 3.53 3.70 3.87 4.04 4.21 4.38 4.55 4.72
4.89 5.06 5.23 5.40 5.57 5.74 5.91 6.08
> c = seq(2,length.out=25,by=0.17)
"bird" "cat" "ferret"
> c = c("bird","cat","ferret")
5 5 12 12 13 13 20 20
> c = rep(c(5,12,13,20),each=2)
```

7.2 Length() of a vector

The length of a vector can be obtained by the *length()* function.

```
> c = (seq(1, 1000, 0.34))
> length(c)
[1] 2939
> c = length(seq(1, 1000, 0.34))
> c
[1] 2939
```

7.3 Vector principles

3 vector principles central to R programming.

- 1. Recycling
- 2. Vectorisation
- 3. Filtering [indexing]

7.3.1 Recycling

When applying an operation to two vectors that requires them to be the same length, R automatically recycles or repeats the shorter vector until it is the same length as the longer one.

> x = c(1,2,3)
> y = c(3,4,5)
> z = 15
> a = c(8,9)
> b = c(10,20)

Looking at *x*+*y*, *1*+*3*=*4*, *2*+*4*=*6* and *3*+*5*=*8*.

```
> x+y
[1] 4 6 8
```

Now looking at x+a, 1+8=9, 2+9=11 and 3+?. As *a* has ran out of elements the first element is recycled again so 3+8=11. The following warning message is received.

```
> x+a
[1] 9 11 11
Warning message:
In x + a : longer object length is not a multiple of shorter object length
```

Now looking at y^*b , $3^*10=30$, $4^*20=80$ and as *a* has ran out of elements the first is recycled again so $5^*10=50$ and a warning message is received.

```
> y*b
[1] 30 80 50
Warning message:
In y * b : longer object length is not a multiple of shorter object length
```

And finally y^*z , $3^*15=45$ and as there are no more elements in z, 15 must be recycled and the final calculations are $4^*15=60$ and $5^*15=75$.

```
> y*z
[1] 45 60 75
Warning message:
In y * b : longer object length is not a multiple of shorter object length
```

The last example above is something that happens all the time, where all the elements in y = c(3,4,5) are multiplied by the single element in b=15.

- 3 x 15 = 45
- 4 x (15 recycled) = 60
- 5 x (15 recycled) = 75

7.3.2 Vectorisation

Scalars are vectors, therefore most functions that you can apply to a single value, you can apply to a vector of values.

```
> sqrt(16)
[1] 4
> x = c(3,6,9,12,15)
> sqrt(x)
[1] 1.732051 2.449490 3.000000 3.464102 3.872983
> sqrt(x/2)
[1] 1.224745 1.732051 2.121320 2.449490 2.738613
> x^2
[1] 9 36 81 144 225
> x/3
[1] 1 2 3 4 5
```

Note here the *modulus*.

> x %% 4 # modulus (x mod y) 5%%2 is 1 [1] 3 2 1 0 3 Breaking this down further.

> 3 %% 4
[1] 3
> 6 %% 4
[1] 2
> 9 %% 4
[1] 1
> 12 %% 4
[1] 0
> 15 %% 4
[1] 3

Looking at a different modulus example, 4 and 5 are continuously recycled until all the elements of x are calculated.

```
> x %% c(4,5)
[1] 3 1 1 2 3
Warning message:
In x%%c(4, 5) :
    longer object length is not a multiple of shorter object length
```

Exercise: write code to produce this vector

```
1 4 9 16 25 36 49 64 81 100
```

> a = seq(1,10,1)
> a^2
[1] 1 4 9 16 25 36 49 64 81 100
> seq(1,10,1)^2
[1] 1 4 9 16 25 36 49 64 81 100
> (1:10)^2
[1] 1 4 9 16 25 36 49 64 81 100

Note the importance of the brackets. Leaving out the brackets gives a completely different answer.

> 1:10^2
[1] 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18
[19] 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36
[37] 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54
[55] 55 56 57 58 59 60 61 62 63 64 65 66 67 68 69 70 71 72
[73] 73 74 75 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90
[91] 91 92 93 94 95 96 97 98 99 100

7.3.3 Filtering [Indexing] of vectors

vector1[vector2]

The elements of vector2 select the elements of vector1 via the [] brackets.

> y = c(10, 1, 16, 3, 8, 5, 13, 55, 34, 13)

Access the third element of the concatenation y.

> **y[3]** [1] 16 Now access the first four elements of the concatenation y.

```
> y[1:4]
[1] 10 1 16 3
So what happens for y[3,4,6,8]?
> y[3,4,6,8]
Error in y[3, 4, 6, 8] : incorrect number of dimensions
```

Well [3,4,6,8] is not a vector. Remember that a vector must be developed as concatenation, via a colon operator, a sequence or a repeat. So rewrite as:

```
> y[c(3,4,6,8)]
[1] 16 3 5 55
```

So what happened?, remember y = c(10, 1, 16, 3, 8, 5, 13, 55, 34, 13). The second vector selects elements from the first vector. *3* selects the third element of *y* which is *16*, *4* selects the forth element *3*, etc..

Exercise (a)

Extract the 3rd, 4th and 7th numbers in this vector.

```
y = c(1,1,2,3,5,8,13,21,34,55)
> y = c(1,1,2,3,5,8,13,21,34,55)
> x = c(3,4,7)
> y[x]
[1] 2 3 13
> y[c(3,4,7)]
[1] 2 3 13
```

Exercise (b)

Extract the first 6 numbers of the vector.

```
> y = c(1,1,2,3,5,8,13,21,34,55)
> x = 1:6
> y[x]
[1] 1 1 2 3 5 8
> y[1:6]
[1] 1 1 2 3 5 8
```

Exercise (c)

Extract the final number of this vector by using the *length()* function.

> y = c(1,1,2,3,5,8,13,21,34,55)

> y[length(y)]
[1] 55

Another way to achieve the same result is to:

> max(y) [1] 55

7.3.4 Other Filtering [Indexing] examples

> y = c(7,8,3,2,4,5,6,3,4,5)		
> min(y) [1] 2	# Returns the smallest element	
> max(y) [1] 8	# Returns the largest element	
> y[2:4] [1] 8 3 2	# Returns elements 2 - 4	
> y[3:6] [1] 3 2 4 5	# Returns elements 3 - 6	

7.3.5 Ordering elements in a vector

The *order()* function returns a permutation which rearranges its first argument into ascending or descending order, breaking ties by further arguments. As can be seen from the example below it returns the positions of the elements in y based on the sequential size starting with position 2 which is 1, then position 4 for the next lowest number 3, etc..

> y = c(10,1,16,3,8,5,13,55,34,13)
> order(y)
[1] 2 4 6 5 1 7 10 3 9 8

If the actual ordered list of the elements instead of the relevant element positions is required then use the following.

> y[order(y)]
[1] 1 3 5 8 10 13 13 16 34 55

Reversing the order. The *rev()* function provides a reversed version of its argument or alternatively negate the *y* in the order function.

```
> y[rev(order(y))]
[1] 55 34 16 13 13 10 8 5 3 1
> y[order(-y)]
[1] 55 34 16 13 13 10 8 5 3 1
```

7.3.6 Extracting elements from a vector

How do I extract numbers from a vector?

- Write down the name of the vector.
- Put square brackets after it.
- Put something inside the square brackets.

```
> y = c(10, 1, 16, 3, 8, 5, 13, 55, 34, 13)
```

> y[5]
[1] 8
> y[7]
[1] 13

What if one wants to extract certain numbers?

All numbers > 7. > y = c(10,1,16,3,8,5,13,55,34,13) > y[y < 7] [1] 1 3 5

All the numbers from 1 to 6.

> y[c(1,2,3,4,5,6)]
[1] 10 1 16 3 8 5
> y[1:6]
[1] 10 1 16 3 8 5

Only numbers equal to 3.

> y = c(10,1,16,3,8,5,13,55,34,13)
> y[y = 3]
[1] 16

Using negatives to eliminate numbers from a list.

```
> y = c(10,1,16,3,8,5,13,55,34,13)
> y[c(-2,-2)]
[1] 10 16 3 8 5 13 55 34 13
> y[-1:-4]
[1] 8 5 13 55 34 13
```

7.4 Booleans

Returns True or False (1 or 0).

7.4.1 Exercise

What happens for: mean(y==3)?

Note: Generic function for the (trimmed) arithmetic mean.

Well the mean of *y*, in other words sum(5)/length(5).

```
> mean(y)
[1] 14.3
> sum(y)/length(y)
[1] 14.3
```

```
'==' means exactly equal to. Remember the output of y == 3?
```

```
> y == 3
[1] FALSE FALSE FALSE TRUE FALSE FALSE FALSE FALSE FALSE FALSE
```

So that is 0,0,0,1,0,0,0,0,0,0.

> sum(c(0,0,0,1,0,0,0,0,0,0)/10)
[1] 0.1

It is the *mean* of the booleans returned from the logic y==3 statement or a single one in 10 elements.

```
> mean(y==3)
[1] 0.1
```

Now put the logic boolean statements inside the square brackets.

```
• T (TRUE) = include

    F (FALSE) = exclude

> y = c(1, 1, 2, 3, 5, 8, 13, 21, 34, 55)
> y[y <= 10]
[1] 1 1 2 3 5 8
> y[y == 3]
[1] 3
> y[y <=10]
[1] 1 1 2 3 5 8
> y[y >= 9]
[1] 13 21 34 55
> y[y > 9]
[1] 13 21 34 55
> y[y != 8]
[1] 1 1 2 3 5 13 21 34 55
> y[y^2 < 10]
[1] 1 1 2 3
```

or map True (T) and False (F) to the values in the vector.

```
> a = c(1,3,5,7,9)
> a[c(T,F,F,T,F)]
[1] 1 7
```

In this case there are not enough boolean statements so they are recycled such that it is the same as: a[c(T,F,T,F,T)].

```
> a[c(T,F)]
[1] 1 5 9
> a[c(T,F,T,F,T)]
[1] 1 5 9
```

7.4.2 Combining Booleans (and (&), or (|))

> x = c(3,6,9,12,15)

Both conditions must be True.

> x > 3 & x < 10
[1] FALSE TRUE TRUE FALSE FALSE</pre>

Either condition is True.

> x == 12 | x < 5
[1] TRUE FALSE FALSE TRUE FALSE</pre>

7.4.3 Boolean operators

- == equals
- < is less than
- > is greater than
- <= less than or equal to
- >= greater than or equal to
- != is not equal to
- & and
- or

Exercise 1

For the vector f = c(1,2,3,6,10,15,21,25,29,30)

- 1. Find all numbers equal to 15.
- 2. Find all numbers greater than 9
- 3. Find all numbers not equal to 10
- 4. Find 6th to 10th vector elements
- 5. Find all except the final element
- 6. Find all numbers that are multiples of 5
- 7. Find all numbers less than or equal to 15
- 8. Find all numbers between 7 and 24

```
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```

```
Answer:
 # Find all numbers equal to 15.
 > f[f==15]
 [1] 15
 # Find all numbers greater than 9.
 > f[f > 9]
 [1] 10 15 21 25 29 30
 # Find all numbers not equal to 10.
 > f[f != 10]
 [1] 1 2 3 6 15 21 25 29 30
 # Find 6th to 10th vector elements.
 > f[f = 6:10]
 [1] 15 21 25 29 30
 # Find all except the final element.
 > f[-length(f)]
 [1] 1 2 3 6 10 15 21 25 29
 # Find all numbers that are multiples of 5.
 > f[(f %% 5) == 0]
 [1] 10 15 25 30
 # Find all numbers less than or equal to 15.
 > f[f <= 15]
 [1] 1 2 3 6 10 15
 # Find all numbers between 7 and 24.
 > f[(f > 7) & (f < 24)]
 [1] 10 15 21
```

Exercise 2

For the vector f = c(1,2,3,6,10,15,21,25,29,30)

- 1. Find all numbers greater than 9
- 2. Find all numbers not equal to 10
- 3. Find 6th to 10th vector elements
- 4. Find all except the final element
- 5. Find all numbers that are multiples of 5
- 6. Find all numbers less than or equal to 15
- 7. Find all numbers between 7 and 24

```
Answer:
 > f = c(1, 2, 3, 6, 10, 15, 21, 25, 29, 30)
  # Find all numbers greater than 9
 > f[f > 9]
  [1] 10 15 21 25 29 30
  # Find all numbers not equal to 10
 > f[f != 9]
  [1] 1 2 3 6 10 15 21 25 29 30
 # Find 6th to 10th vector elements
  > f[6:10]
  [1] 15 21 25 29 30
  # Find all except the final element
  > f[-length(f)]
  [1] 1 2 3 6 10 15 21 25 29
  # Find all numbers that are multiples of 5
  > f[f %% 5 == 0]
  [1] 10 15 25 30
  # Find all numbers less than or equal to 15
 > f[f <= 15]
  [1] 1 2 3 6 10 15
  # Find all numbers between 7 and 24
 > f[(f > 7) & (f < 16)]
  [1] 10 15
```

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8. Building on Vectors, other data structures in R

8.1 Basics of structures

vector[row]

matrix[*row,column*]

array[*row,column,level*]

8.1.1 Vectors

• The vector is the fundamental data type in *R*, scalars and matrices are just special types of vectors and that all things that apply to vectors, apply to these.

8.1.2 Matrix

- A matrix is a collection of data elements arranged in a two-dimensional rectangular layout. The following is an example of a matrix with 2 rows and 3 columns.
- The elements must be of the same type.
- Here is an example.

```
> a.matrix = matrix(c(2, 4, 3, 1, 5, 7), # the data elements
                                    # number of rows
                  nrow=2,
                  ncol=3,
                                    # number of columns
                 )
> a.matrix
    [,1] [,2] [,3]
[1,] 2 3
                5
[2,]
      4
                7
           1
> a.matrix = matrix(c(2, 4, 3, 1, 5, 7), # the data elements
                  nrow=2,
                                  # number of rows
                  ncol=3,
                                    # number of columns
                  byrow = TRUE
                                   # fill matrix by rows
                 )
> a.matrix
   [,1] [,2] [,3]
[1,] 2 4 3
[2,]
      1
           5
                7
```

Indexing Matrices

matrix.name[Row,Column]

 So to access row 2 and column 3 - matrix.name[2,3] or to access all elements in row 2 - matrix.name[2,] or all the elements in column 3 - matrix.name[,3].

```
> a.matrix[2,3]
[1] 7
> a.matrix[2,]
[1] 1 5 7
```

```
> a.matrix[,3]
[1] 3 7
> a.matrix[1:2,2:3]
     [,1] [,2]
[1,]
     4 3
[2,]
       5
            7
> a.matrix[-2,]
[1] 2 4 3
> a.matrix[,-2]
   [,1] [,2]
    2 3
[1,]
[2,]
       1
            7
```

• How about finding all the rows where column 3 is greater than 6?

```
> a.matrix [a.matrix[,3] > 6,]
[1] 1 5 7
```

Apply boolean questions to matrix

For a matrix of four rows and three columns with the data 1,5,9,2,6,10,3,7,11,4,8,12 get the output of the following *R* commands.

- z.matrix == 3
- z.matrix[,3] <= 10
- z.matrix != 3
- any(z.matrix[1,] == 3)
- any(z.matrix > 15)
- all(z.matrix == 3)
- all(z.matrix < 20)
- complete.cases(z.matrix)

```
> z.matrix = matrix(c(1,5,9,2,6,10,3,7,11,4,8,12),
                     nrow=4,
                     ncol=3,
                     byrow=TRUE
                    )
> z.matrix
     [,1] [,2] [,3]
[1,] 1 5 9
[2,] 2 6 10
[3,] 3 7 11
[3,]
[4,]
           8
      4
                12
> z.matrix == 3
     [,1] [,2] [,3]
[1,] FALSE FALSE FALSE
[2,] FALSE FALSE FALSE
[3,] TRUE FALSE FALSE
[4,] FALSE FALSE FALSE
> z.matrix[,3] <= 10</pre>
[1] TRUE TRUE FALSE FALSE
```

> z.matrix != 3 [,1] [,2] [,3] [1,] TRUE TRUE TRUE [2,] TRUE TRUE TRUE [3,] FALSE TRUE TRUE [4,] TRUE TRUE TRUE > any(z.matrix[1,] == 3) [1] FALSE > any(z.matrix > 15) [1] FALSE > all(z.matrix == 3) [1] FALSE > all(z.matrix < 20)</pre> [1] TRUE > complete.cases(z.matrix) [1] TRUE TRUE TRUE TRUE

8.1.3 Array

Arrays are the R data objects which can store data in more than two dimensions. An array is created using the *array()* function. It takes vectors as input and uses the values in the *dim* (dimensions) parameter to create an array. The *dim* parameters define matrices of three rows, four columns and two deep, i.e. two matrices.

array[row,column,level]

Here is an example.



Illustration 3: Array

> v1 = c(5,9,3) > v2 = c(10,11,12,13,14,15) > v3 = c(21,23,24,26,27,28,29) > v4 = c(32,34,35,37,31,34,45,46) > p = array(c(v1,v2,v3,v4),dim = c(3,4,2))

```
> p
, , 1
    [,1] [,2] [,3] [,4]
[1,] 5 10 13 21
[2,] 9 11 14 23
[3,] 3 12 15 24
, , 2
    [,1] [,2] [,3] [,4]
[1,] 26 29 35 34
[2,] 27 32 37 45
[3,] 28 34 31 46
```

• Here is another example.

```
> v1 = c(5,9,3)
> v2 = c(10,11,12,13,14,15)
> p = array(c(v1,v2),dim = c(3,3,2))
> p
, , 1
       [,1] [,2] [,3]
[1,] 5 10 13
[2,] 9 11 14
[3,] 3 12 15
, , 2
       [,1] [,2] [,3]
[1,] 5 10 13
[2,] 9 11 14
[3,] 3 12 15
```

Names can be given to the rows, columns and matrices in the array by using the *dimnames* parameter.

Here is example.

			СС	DL1	СС	DL2	СС	DL3		
ROW1		ļ	5	1	.0	0 1		2		
	RC	W2		со	L1	со	L2	со	L3	
	RO	RO	W1	5	5	1	0	1	3	
l		RO	W2	ç)	1	1	1	4	
		RO	W3	3	3	1	2	1	5	1

Illustration 4: Array 2

> v1 = c(5, 9, 3)> v2 = c(10,11,12,13,14,15) > column.names = c("COL1", "COL2", "COL3") > row.names = c("ROW1", "ROW2", "ROW3") > matrix.names = c("Matrix1","Matrix2") > p = array(c(v1,v2),dim = c(3,3,2),dimnames = list(row.names,column.names,matrix.names)) > p , , Matrix1 COL1 COL2 COL3 ROW151013ROW291114 3 12 15 R0W3 , , Matrix2 COL1 COL2 COL3 ROW151013ROW291114 ROW3 3 12 15

Exercise (a): Matrix

	[,1]	[,2]	[,3]	[,4]	[,5]
[1,]	1	3	5	7	9
[2,]	11	13	15	17	19
[3,]	21	23	25	27	29
[4,]	31	33	35	37	39

Create a matrix like the one above, containing the sequence of odd numbers starting at 1 and counting up to fill the matrix.

Answer:

Exercise (b): Matrix

- 1. Create a matrix (called d1) with 6 rows and 4 columns (*byrow=F*) using the sequence of odd numbers starting at 1.
- 2. Extract the number from the 3rd column, 4th row and assign it to the variable name g1.
- 3. Extract the 6th row
- 4. Extract columns 2 & 4 from d1 and call the new matrix d2
- 5. Create a new matrix (d3), by doing something to d1, that contains the sequence of EVEN numbers starting at 2
- 6. Change d3 element 3rd row, 2nd column so that it equals 500. Look at *d3*. Change 3rd row, 2nd column to *Harry*. Look at *d3* again. Has anything changed? Why? Change *Harry* back to *500*. Now what has happened?

```
> d1 = matrix(seq(1,length.out=24,by=2),
              nrow=6,ncol=4,
              byrow=FALSE
             )
> d1
    [,1] [,2] [,3] [,4]
[1,]
      1 13
               25
                    37
[2,]
      3 15
              27
                    39
[3,]
     5 17 29 41
[4,]
      7 19 31 43
      9 21
[5,]
              33 45
[6,]
     11 23
               35
                   47
> g1 = d1[4,3]
> g1
[1] 31
> d1[6,]
[1] 11 23 35 47
> c(d1[,2], d1[,4])
[1] 13 15 17 19 21 23 37 39 41 43 45 4
> d2 = matrix(c(d1[,2], d1[,4]),
            nrow=6, ncol=2,
            byrow=FALSE
            )
> d3 = d1 + 1
> d3
    [,1] [,2] [,3] [,4]
[1,]
          14
               26
                    38
      2
[2,]
      4
         16
               28
                   40
                    42
[3,]
      6 18
              30
[4,]
      8 20
              32 44
[5,]
     10 22
               34
                   46
[6,] 12 24
               36
                   48
> d3[3,2] = 500
```

```
> d3
    [,1] [,2] [,3] [,4]
[1,] 2 14 26 38
[2,] 4 16 28 40
[3,] 6 500 30 42
[4,] 8 20 32 44
[5,] 10 22 34 46
[6,] 12 24 36 48

> d3[3,2] = 'Harry'
> d3
    [,1] [,2] [,3] [,4]
[1,] "2" "14" "26" "38"
[2,] "4" "16" "28" "40"
[3,] "6" "Harry" "30" "42"
[4,] "8" "20" "32" "44"
[5,] "10" "22" "34" "46"
[6,] "12" "24" "36" "48"
```

All values became strings because in a matrix all values must be of the same type.

```
> d3[3,2] = 500
> d3
       [,1] [,2] [,3] [,4]
[1,] "2" "14" "26" "38"
[2,] "4" "16" "28" "40"
[3,] "6" "500" "30" "42"
[4,] "8" "20" "32" "44"
[5,] "10" "22" "34" "46"
[6,] "12" "24" "36" "48"
```

String type has been maintained.

Exercise (c): Matrix filtering

```
> chick = matrix(c(seq(1,5,1),
               c(10,15,12,13,15,8,11,9,12,13)),
               nrow=5,ncol=3,byrow=FALSE
              )
> colnames(chick) = c('Individual', 'Weight', 'Age')
> chick
   Individual Weight Age
[1,] 1 10 8
[2,]
                15 11
           2
[3,]
           3 12 9
           4 13 12
5 15 13
[4,]
           5
[5,]
                 15 13
```

From the chick matrix:

- 1. All rows where *weight* is less than 15g.
- 2. Using the mean() function, calculate the mean age of chicks >10g in weight.
- 3. Add an extra column (2,5,3,5,6) to the *chick* matrix using the *cbind()* function.

cbind(): - Take a sequence of vector, matrix or data-frame arguments and combine by columns or rows, respectively.

> chick[chick[,2] < 15,]</pre> Individual Weight Age 1 10 8 [1,]3 [2,] 12 9 4 13 12 [3,] > mean(chick[chick[,2] > 10, 'Age']) [1] 11.25 > chick = cbind(chick, c(2, 5, 3, 5, 6)) > chick Individual Weight Age 1 10 8 2 [1,] 2 15 11 5 [2,] 3 12 9 3 [3,] 4 13 12 5 [4,] [5,] 5 15 13 6

8.1.4 The 'apply' family of functions

apply(): returns a vector or array or list of values obtained by applying a function to margins of an array or matrix.

apply(X, MARGIN, FUN, ...)

- X: an array, including a matrix.
- **MARGIN:** 1 = rows, 2 = columns, c(1,2) = both.
- FUN: the function.

```
> v1 = c(5, 9, 3)
> v2 = c(10, 11, 12, 13, 14, 15, 16, 17, 18)
> v3 = c(21, 23, 24, 26, 27, 28, 29, 30)
> p = matrix(c(v1, v2, v3), nrow = 4, ncol = 5)
> p
     [,1] [,2] [,3] [,4] [,5]
[1,]
      5 11 15 21 27
[2,]
       9 12 16
                  23 28
      3 13 17 24 29
[3,]
     10 14 18 26 30
[4,]
# Sum of each column
> apply(p, 2, sum)
[1] 27 50 66 94 114
# Get the mean of each row
> apply(p, 1, mean)
[1] 15.8 17.6 17.2 19.6
> apply(p, c(1,2), mean)
    [,1] [,2] [,3] [,4] [,5]
[1,]
    5 11 15 21 27
[2,]
      9 12 16 23 28
[3,]
      3 13 17
                   24 29
[4,]
     10
          14
              18
                   26
                        30
```

While it is possible to supply a vector c(1,2) to the *MARGIN* argument it makes little sense with a *matrix*. However with an *array* it operates between the matrices within the *array*.

> v1 = c(5, 9, 3)> v2 = c(10,11,12,13,14,15) > v3 = c(21, 23, 24, 26, 27, 28, 29)> v4 = c(32, 34, 35, 37, 31, 34, 45, 46)> a = array(c(v1, v2, v3, v4), dim = c(3, 4, 2))> a , , 1 [,1] [,2] [,3] [,4] [1,] 5 10 13 21 [2,] 9 11 14 23 [3,] 3 12 15 24 , , 2 [,1] [,2] [,3] [,4] [1,] 26 29 35 34 32 37 45 [2,] 27 [3,] 34 31 46 28 > apply(a, c(1,2), mean) [,1] [,2] [,3] [,4] [1,] 15.5 19.5 24.0 27.5 [2,] 18.0 21.5 25.5 34.0 [3,] 15.5 23.0 23.0 35.0

Other apply() functions

Other apply() functions exists, some of which are described later.

- lapply(): For lists and data-frames.
- **sapply():** A simplified wrapper function for *lapply()*.
- **sapply():** The multivariate apply which can vectorise arguments to a function that is not usually accepting vectors as arguments.

In short, *mapply()* applies a function to multiple list or multiple vector arguments.

8.2 Recap exercises

1. Create the following vectors:

 Answer > seq(3, 48, 3) [1] 3 6 9 12 15 18 21 24 27 30 33 36 39 42 45 48 > rep(3,25) > c(20,13,16,17,18,20) [1] 20 13 16 17 18 20 > seq(100, 61, -3) [1] 100 97 94 91 88 85 82 79 76 73 70 67 64 61 > seq(1, 20, 1) [1] 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 > seq(13, 29, 1) [1] 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 > seq(1, 19, 2) [1] 1 3 5 7 9 11 13 15 17 19 > c('bird','fish','cricket') [1] "bird" "fish" "cricket"

- 2. From the following vector
 - 3 6 9 12 15 18 21 24 27 30 33 36 39 42 45 48
 - Multiply each element by 10.
 - Take the square root of each element.
 - Add three to each element.
 - Standardise each element to the mean of the vector. (i.e. divide each element by the vector's mean).
 - Multiply every second element by the vector's length.
 - Answer

```
> d = seq(3, 48, 3)
> d
 [1] 3 6 9 12 15 18 21 24 27 30 33 36 39 42 45 48
> d * 10
 [1] 30 60 90 120 150 180 210 240 270 300 330 360 390 420 450 480
> sqrt(d)
 [1] 1.732051 2.449490 3.000000 3.464102 3.872983 4.242641 4.582576
 [8] 4.898979 5.196152 5.477226 5.744563 6.000000 6.244998 6.480741
 [15] 6.708204 6.928203
> d + 3
 [1] 6 9 12 15 18 21 24 27 30 33 36 39 42 45 48 51
> d/(mean(d))
  [1] \ 0.1176471 \ 0.2352941 \ 0.3529412 \ 0.4705882 \ 0.5882353 \ 0.7058824 \\
 [7] 0.8235294 0.9411765 1.0588235 1.1764706 1.2941176 1.4117647
 [15] 1.5294118 1.6470588 1.7647059 1.8823529
> d * c(1, length(d))
 [1] 3 96
              9 192 15 288 21 384 27 480 33 576 39 672 45 768
```

3. Using vector indexing extract the following vectors from the *a* vector where:

```
a = c(5, 1, 6, 10, 2, 7, 13, 12, 8)
516
5 5 5 13 5 5 1 6 10 2 7 13 12
1 2 5 7 8 10 12 13
numbers less than 10
numbers equal to 7
all even numbers

    Answer

       > a[1:3]
       [1] 5 1 6
       > c(rep(a[1],3),a[7],a[1])
       [1] 5 5 5 13 5
       > a[1:length(a) - 1]
       [1] 5 1 6 10 2 7 13 12
       > c(a[2],a[5],a[1],a[3],a[6],a[9],a[4],a[8],a[7])
       [1] 1 2 5 6 7 8 10 12 13
       > a[a < 10]
       [1] 5 1 6 2 7 8
       > a[a == 7]
       [1] 7
       > a[a %% 2 == 0]
       [1] 6 10 2 12 8
```

4. Create the following matrix

	[,1]	[,2]	[,3]	[,4]
[1,]	2	4	6	8
[2,]	10	12	14	16
[3,]	18	20	22	24
[4,]	26	28	30	32
[5,]	34	36	38	40
[6,]	42	44	46	48

- Extract the 3rd column.
- Extract the 5th row.
- Create a new matrix from the first three rows and last two columns.
- Extract all numbers greater than 25.
- Extract all numbers greater than 35 from row five.
- Extract all numbers greater than 35 from column four.
- Extract all numbers between 10 & 30 from column one.

Answer > x.matrix = matrix(c(seq(2,48,2)), nrow=6,ncol=4,byrow=TRUE) > x.matrix [,1] [,2] [,3] [,4] 2 4 [1,]6 8 [2,] 10 12 14 16 18 20 22 [3,] 24 [4,] 28 30 32 26 [5,] 34 36 38 40 44 46 48 [6,] 42 > x.matrix[,3] [1] 6 14 22 30 38 46 > x.matrix[5,] [1] 34 36 38 40 > x.matrix[1:3,3:4] [,1] [,2] [1,]6 8 [2,] 14 16 [3,] 22 24 > x.matrix[c(x.matrix[]) > 25] [1] 26 34 42 28 36 44 30 38 46 32 40 48 > x.matrix[5,][x.matrix[5,] > 35] [1] 36 38 40 > x.matrix[,4][x.matrix[,4] > 35] [1] 40 48 > x.matrix[,1][x.matrix[,1] > 10 & x.matrix[,1] < 30]</pre> [1] 18 26

5. Create the following matrix

	individual	weight	age
[1,]	1	4	13
[2,]	2	7	15
[3,]	3	2	12
[4,]	4	9	14
[5,]	5	2	20

- Give the columns the names above: use *colnames()*.
- Extract which individuals are older than five years old.
- What is the mean weight of all individuals.
- What is the mean weight of individuals older than five.
- Add a new row i.e. c(6, 5, 15) using *rbind()* function.

```
    Answer

    > y.matrix = matrix(c(seq(1,5,1),c(4,7,2,9,2,13,15,12,14,20)),
                        nrow=5,ncol=3,byrow=FALSE
                       )
    > colnames(y.matrix) = c('individual', 'weight', 'age')
    > v.matrix
        individual weight age
    [1,]
                 1
                        4 13
                        7 15
    [2,]
                  2
                        2 12
    [3,]
                  3
    [4,]
                  4
                         9 14
    [5,]
                        2 20
                  5
    > y.matrix[y.matrix[,3] > 5,'individual']
    [1] 1 2 3 4 5
    or
    > y.matrix[y.matrix[,3] > 5,1]
    [1] 1 2 3 4 5
    > mean(y.matrix[y.matrix[,1],2])
    [1] 4.8
    > mean(y.matrix[y.matrix[,3] > 5,2])
    [1] 4.8
    > y.matrix = rbind(y.matrix, c(6,5,15))
    > y.matrix
         individual weight age
    [1,]
                1
                      4 13
                        7 15
    [2,]
                 2
    [3,]
                  3
                        2 12
    [4,]
                  4
                        9 14
                         2 20
     [5,]
                  5
    [6,]
                         5 15
                  6
```

8.3 Operators as functions

Operators can be used like functions. Place the values to supply to the operator function and the result will return as expected.

```
> '+'(1,2)
[1] 3
> '='(x,2)
> x
[1] 2
> '<-'(y,3)
> y
[1] 3
```

Even arguments can be used as functions.

> '**^'(y,2)** [1] 9

9. Output to standard out

9.1 print() and cat()

print(): prints its argument.

cat(): Concatenate and print. This function outputs the objects, concatenating the representations.

cat() performs much less conversion than *print()*.

```
> string = 'My string'
> numbers = 1:12
> t_date = date()
> chain = c(string, numbers, t_date)
> cat(chain)
My string 1 2 3 4 5 6 7 8 9 10 11 12 Tue Sep 18 13:00:07 2018
> print(chain)
                                 "1"
 [1] "My string"
 [3] "2"
                                 "3"
 [5] "4"
                                 "5"
 [7] "6"
                                 "7"
 [9] "8"
                                 "9"
[11] "10"
                                 "11"
[13] "12"
                                 "Tue Sep 18 13:00:07 2018"
> cat(string, numbers, t_date)
My string 1 2 3 4 5 6 7 8 9 10 11 12 Tue Sep 18 13:00:07 2018
> print(string, numbers, t_date)
Error in print.default(string, numbers, t_date) :
  invalid 'quote' argument
> print(c(string, numbers, t_date))
 [1] "My string"
                                 "1"
 [3] "2"
                                 "3"
 [5] "4"
                                 "5"
 [7] "6"
                                 "7"
[9] "8"
                                 "9"
[11] "10"
                                 "11"
[13] "12"
                                 "Tue Sep 18 13:00:07 2018"
```

9.2 sprintf()

A wrapper for the C function *sprintf*, that returns a character vector containing a formatted combination of text and variable values. The format string is a character string, beginning and ending in its initial shift state, if any. The format string is composed of zero or more directives: ordinary characters (not %), which are copied verbatim to the output stream; and conversion specifications, each of which results in fetching zero or more subsequent arguments. Each conversion specification is introduced by the character %, and ends with a conversion specifier. In between there may be zero or more flags, an optional minimum *field width*, an optional *precision* and an optional *length modifier*.

Notation	Description
%s	a string
%d	an integer
%0xd	an integer padded with x leading zeros
%f	decimal notation with six decimals
%.xf	floating point number with x digits after decimal point
%e	compact scientific notation, e in the exponent
%E	compact scientific notation, E in the exponent
%g	compact decimal or scientific notation (with e)

9.3 paste() and paste0() functions

paste(): Concatenates vectors after converting to character with a single space as a separator.

paste0(): Concatenates vectors after converting to character with no separator.

- *sep*: The element which separates every term. It should be specified with character string format.
- *collapse*: The element which separates every result. It should be specified with character string format and it is optional.

The difference between *paste()* and *paste0()* is that the argument *sep* by default is " " (paste) and "" in (paste0).

```
> string = 'My string'
> numbers = 1:12
> t_date = date()
> chain = c(string, numbers, t_date)
> p_string = paste(string, numbers, t_date)
> chain
[1] "My string"
                                "1"
[3] "2"
                                "3"
[5] "4"
                                "5"
[7] "6"
                                "7"
[9] "8"
                                "9"
[11] "10"
                                "11"
[13] "12"
                                "Tue Sep 18 13:00:07 2018"
> p_chain
[1] "My string 1 Tue Sep 18 13:00:07 2018"
 [2] "My string 2 Tue Sep 18 13:00:07 2018"
[3] "My string 3 Tue Sep 18 13:00:07 2018"
[4] "My string 4 Tue Sep 18 13:00:07 2018"
[5] "My string 5 Tue Sep 18 13:00:07 2018"
[6] "My string 6 Tue Sep 18 13:00:07 2018"
 [7] "My string 7 Tue Sep 18 13:00:07 2018"
 [8] "My string 8 Tue Sep 18 13:00:07 2018"
 [9] "My string 9 Tue Sep 18 13:00:07 2018"
[10] "My string 10 Tue Sep 18 13:00:07 2018"
[11] "My string 11 Tue Sep 18 13:00:07 2018"
[12] "My string 12 Tue Sep 18 13:00:07 2018"
```

9.4 Numbers

Double precision value, in *fixed point* decimal notation.

```
> number = 1234567.894567
> number1 = sprintf("%.2f", number)
> print(number1)
[1] "1234567.89"
> number1 = sprintf("%e", number)
> print(number1)
[1] "1234567.89"
# Exponential output
> number2 = sprintf("%e", number)
> print(number2)
[1] "1.234568e+06"
# Scientific notation
> number3 = sprintf("%a", number)
> print(number3)
[1] "0x1.2d687e50257c9p+20"
```

Note that with integers, if an non-integer is given R will give an error. Either give the input as an integer. Optionally print using the floating point number with zero digits after the decimal point to get an integer from a non integer input.

```
# Integer
> number4 = sprintf("%d", number)
Error in sprintf("%d", 1234567.894567) :
    invalid format '%d'; use format %f, %e, %g or %a for numeric objects
> number1 = sprintf("%d", 1234567)
> print(number1)
[1] "1234567"
> number5 = sprintf("%.0f", number)
> print(number5)
[1] "1234568"
```

9.5 Character string

> word = "R programming"

```
> word1 = sprintf("%s is fun.", word)
> print(word1)
[1] "R programming is fun."
```

10. Flow control & Data frames

10.1 if & else conditionals

Used to create flexibility in programming.

Write an *if()* statement that tells if a number (*x*) is positive (the console returns the word *positive*).

```
> x = 5
> y = -4
> if (x > 0) print('positive') else print('negative')
[1] "positive"
> if (y > 0) print('positive') else print('negative')
[1] "negative"
> if (x > 0){
    print('positive')
}else{
    print('negative')
}
[1] "positive"
> if (y > 0){
    print('positive')
}else{
    print('negative')
3
[1] "negative"
```

• *if & if else* are not vectorised, they only take single values.

10.2 ifelse conditional

ifelse(boolean condition,output if TRUE,output if FALSE)

```
> ifelse((x > 0), 'positive', 'negative')
[1] "positive"
> ifelse((y > 0), 'positive', 'negative')
[1] "negative"
```

ifelse is a vectorised function.

10.2.1 Exercise: ifelse

age = c(3,5,7,4,9,3,5,4,8,6)

• Create a new variable (age.cat) where ages four and below are 0 and above four are 1.

```
> age = c(3,5,7,4,9,3,5,4,8,6)
> age.cat = ifelse((age < 5),0,1)
> age.cat
[1] 0 1 1 0 1 0 1 0 1 1
```

• Create a new variable (age.limit) where ages six and above are included in a single age category of *6*.

```
> age.limit = ifelse((age < 6), age, 6)
> age.limit
[1] 3 5 6 4 6 3 5 4 6 6
```

• Create a new variable (age.3cat) where ages are in three categories 1=(1 to 4), 2=(5 to 6), and 3=(7 to 9)

```
> age.3cat = ifelse(
              (age < 5),
              print ('1'),
              ifelse(
                (age > 4 & age < 7),
                print ('2'),
                ifelse(
                  (age > 6),
                  print ('3'),"NA"
                )
              )
            )
[1] "1"
[1] "2"
[1] "3"
> age.3cat
[1] "1" "2" "3" "1" "3" "1" "2" "1" "3" "2"
```

10.3 Lists

- Lists are the *R* objects which contain elements of different types like numbers, strings, vectors and another list inside it. A list can also contain a matrix or a function as its elements. List is created using *list()* function.
- Here is an example.

```
> list_data = list("Orange", "Green", c(43,13,61), TRUE, 42.18, 218.2)
> list_data
[[1]]
[1] "Orange"
[[2]]
[1] "Green"
[[3]]
[1] 43 13 61
[[4]]
[1] TRUE
[[5]]
[1] 42.18
[[6]]
[1] 218.2
```

Accessing elements in the list. Here accessing the forth element in the list.

```
> list_data[4]
[[1]]
[1] TRUE
```

• The *R* List can combine objects of any mode into a single object.

• Use *unclass()* to view the list.

```
> unclass(w)
$site
[1] 5
$species
[1] 2 3 4 5 6 7 8 9
$names
[1] "Captain Kirk" "Richard Dawkins"
```

 Here are the various tools to extract from the list. These tools all index the second sub-vector of the list.

```
> w$site
[1] 5
> w$species
[1] 2 3 4 5 6 7 8 9
> w[['species']]
[1] 2 3 4 5 6 7 8 9
```

```
> w[[1]]
[1] 5
> w[3]
$names
[1] "Captain Kirk" "Richard Dawkins"
```

· Adding to the list.

```
> w$shopping.list = c('milk','eggs')
```

```
> w
$site
[1] 5
$species
[1] 2 3 4 5 6 7 8 9
$names
[1] "Captain Kirk" "Richard Dawkins"
$shopping.list
[1] "milk" "eggs"
```

To remove *shopping.list* from our list *w*.

```
> w$shopping.list = NULL
> w
$site
[1] 5
$species
[1] 2 3 4 5 6 7 8 9
$names
[1] "Captain Kirk" "Richard Dawkins"
```

Most statistical outputs (objects) are contained within an R list.

Im(): - is used to fit linear models. **mtcars:** - is a stored dataset.

```
> data(mtcars)
> a = lm(mpg~wt, data=mtcars)
> summary(a)
Call:
lm(formula = mpg ~ wt, data = mtcars)
Residuals:
             1Q Median
                              ЗQ
   Min
                                      Мах
-4.5432 -2.3647 -0.1252 1.4096 6.8727
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 37.2851 1.8776 19.858 < 2e-16 ***
wt -5.3445 0.5591 -9.559 1.29e-10 ***
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 3.046 on 30 degrees of freedom
Multiple R-squared: 0.7528, Adjusted R-squared: 0.7446
F-statistic: 91.38 on 1 and 30 DF, p-value: 1.294e-10
```

> unclass(a) \$coefficients (Intercept) wt 37.285126 -5.344472 \$residuals Mazda RX4 Mazda RX4 Waq Datsun 710 Hornet 4 Drive -2.2826106 -0.9197704 -2.0859521 1.2973499 Hornet Sportabout Valiant Duster 360 Merc 240D -0.2001440 -0.6932545 -3.9053627 4.1637381 Merc 230 Merc 280 Merc 280C Merc 450SE 2.3499593 0.2998560 -1.1001440 0.8668731 Merc 450SLC Cadillac Fleetwood Lincoln Continental Merc 450SL -1.8830236 -0.05024721.1733496 2.1032876 Chrysler Imperial Fiat 128 Honda Civic Toyota Corolla 5.9810744 6.8727113 1.7461954 6.4219792 Toyota Corona Dodge Challenger AMC Javelin Camaro Z28 -2.6110037 -2.9725862 -3.7268663 -3.4623553 Pontiac Firebird Fiat X1-9 Porsche 914-2 Lotus Europa 0.3564263 2,4643670 0.1520430 1.2010593 Ford Pantera L Ferrari Dino Maserati Bora Volvo 142F -4.5431513 -2.7809399 -3.2053627 -1.0274952 \$effects (Intercept) wt -113.6497374 -29.1157217 -1.6613339 1.6313943 0.1111305 -0.3840041 -3.6072442 4.5003125 2.6905817 0.6111305 -0.7888695 1.1143917 0.2316793 -1.60615711.3014525 2.2137818 6.0995633 7.3094734 2.2421594 6.8956792 -2.2010595 -2.6694078 -3.4150859 -3.1915608 2.7346556 0.8200064 0.5948771 1.7073457 -4.2045529-2.4018616-2.9072442 -0.6494289 \$rank [1] 2 \$fitted.values Hornet 4 Drive Mazda RX4 Mazda RX4 Wag Datsun 710 23.282611 21.919770 24.885952 20.102650 Hornet Sportabout Valiant Duster 360 Merc 240D 18.900144 18.793255 18.205363 20.236262 Merc 230 Merc 280 Merc 280C Merc 450SE 20.450041 18.900144 18.900144 15.533127 Merc 450SL Merc 450SLC Cadillac Fleetwood Lincoln Continental 17.350247 17.083024 9.226650 8.296712 Chrysler Imperial Fiat 128 Honda Civic Toyota Corolla 25.527289 28.653805 27.478021 8,718926 Dodge Challenger Toyota Corona AMC Javelin Camaro Z28 24.111004 18.472586 18.926866 16.762355 Pontiac Firebird Fiat X1-9 Porsche 914-2 Lotus Europa 16.735633 26.943574 25.847957 29.198941 Ford Pantera L Ferrari Dino Maserati Bora Volvo 142E 20.343151 22,480940 18,205363 22,427495 \$assign [1] 0 1 \$ar \$qr (Intercept) wt Mazda RX4 -5.6568542 -18.199514334 Mazda RX4 Wag 0.1767767 5.447820482

Datsun 710

0.148230003

0.1767767

0.1767767 -0.016055881 Hornet 4 Drive Hornet Sportabout 0.1767767 -0.057356801 Notice oportabout0.1/67767-0.057356801Valiant0.1767767-0.061027994Duster 3600.1767767-0.081219555Merc 240D0.1767767-0.004124504Merc 2800.1767767-0.004124504Merc 280C0.1767767-0.057356801Merc 450SE0.1767767-0.172999378Merc 450SLC0.1767767-0.119767081Cadilloo Flootuation0.1767767-0.119767081 Lincoln Continental 0.1767767 -0.421539139 Chrysler Imperial 0.1767767 -0.407037927 Fiat 128 0.1767767 0.170257160 Honda Civic 0.1767767 0.277639553 Toyota Corolla 0.1767767 0.237256431 Toyota Corona 0.1767767 0.237256431 Dodge Challenger 0.1767767 0.121613854 Dodge Challenger 0.1767767 -0.072041573 AMC Javelin 0.1767767 -0.056439003 Camaro Z28 0.1767767 -0.130780659 Pontiac Firebird 0.1767767 -0.131698458 Fiat X1-9 0.1767767 0.218900467 Porsche 914-2 0.1767767 0.181270739 Lotus Europa 0.1767767 0.296362637 Ford Pantera L 0.1767767 -0.007795696 Ferrari Dino 0.1767767 -0.081219555 Lincoln Continental 0.1767767 -0.421539139 Maserati Bora 0.1767767 -0.081219555 Volvo 142E 0.1767767 0.063792566 attr(,"assign") [1] 0 1 \$qraux [1] 1.176777 1.046354 \$pivot [1] 1 2 \$tol [1] 1e-07 \$rank [1] 2 attr(,"class") [1] "qr" \$df.residual [1] 30 \$xlevels named list() \$call lm(formula = mpg ~ wt, data = mtcars) \$terms mpg ~ wt attr(,"variables") list(mpg, wt) attr(,"factors") wt mpg 0 wt 1 attr(,"term.labels") [1] "wt" attr(,"order")

[1] 1 attr(,"intercept") [1] 1 attr(,"response") [1] 1 attr(,".Environment") <environment: R GlobalEnv> attr(,"predvars") list(mpg, wt) attr(,"dataClasses") mpg wt "numeric" "numeric" \$model mpg wt Mazda RX4
 Mazda RX4
 21.0
 2.620

 Mazda RX4 Wag
 21.0
 2.875
 Datsun 71022.8 2.320Hornet 4 Drive21.4 3.215 Hornet Sportabout 18.7 3.440 18.1 3.460 14.3 3.570 Valiant Duster 360 Merc 240D 24.4 3.190 22.8 3.150 Merc 230 19.2 3.440 17.8 3.440 16.4 4.070 Merc 280 Merc 280C Merc 450SE
 Merc
 450SL
 17.3
 3.730

 Merc
 450SLC
 15.2
 3.780
 Cadillac Fleetwood 10.4 5.250 Lincoln Continental 10.4 5.424 Chrysler Imperial 14.7 5.345 Fiat 128 32.4 2.200 Fiat 12832.4 2.200Honda Civic30.4 1.615
 Honda Civic
 30.4
 1.615

 Toyota Corolla
 33.9
 1.835

 Toyota Corona
 21.5
 2.465

 Dodge Challenger
 15.5
 3.520
 AMC Javelin 15.2 3.435 Camaro Z28 13.3 3.840 Pontiac Firebird 19.2 3.845 Pontiac Files Fiat X1-9 27.3 1.000 bo 014-2 26.0 2.140 co 4 1 513

 Porsche 914-2
 26.0
 2.140

 Lotus Europa
 30.4
 1.513

 Ford Pantera L
 15.8
 3.170

 Ferrari Dino
 19.7
 2.770

 Maserati Bora
 15.0
 3.570

 Volvo 142E
 21.4
 2.780

 Volvo 142E 21.4 2.780

10.4 Table

• Tables return the number of each element in a vector. The vector contains one *a*, three *b* and one *c*.

```
> vector = c("a", "a", "b", "b", "b", "c")
> table(vector)
vector
a b c
2 3 1
> z = table(vector)
> z[2]
b
3
```

10.5 for() and while()

for(): is used to repeat a set of instructions, and it is used when you know in advance the values that the loop variable will have each time it goes through the loop.

while(): is be used to repeat a set of instructions, and it is often used when you do not know in advance how often the instructions will be executed.

```
> for (element in seq(0:10)){
        print(element);
 }
 [1] 1
[1] 2
[1] 3
[1] 4
[1] 5
[1] 6
[1] 7
[1] 8
[1] 9
[1] 10
[1] 11
> numbers = c(1,2,4,8,16,32,64)
> for (loop in numbers){
       cat("Loop number: ",loop,"\n");
 }
Loop number: 1
Loop number: 2
Loop number: 4
Loop number:
              8
Loop number: 16
Loop number: 32
Loop number: 64
> cars = list('Ford', 'Saab', 'Toyota', 'Nissan', 'Volvo', 'Renault')
> position = 1
> for (loop in cars){
    cat("Car number: ",position, ':', loop,"\n");
    position = position + 1;
 }
Car number: 1 : Ford
Car number: 2 : Saab
Car number: 3 : Toyota
```

```
Car number: 4 : Nissan
Car number: 5 : Volvo
Car number: 6 : Renault
> position = 1
> while(position < 5){</pre>
     print (cars[position]);
     position = position + 1;
 }
[[1]]
[1] "Ford"
[[1]]
[1] "Saab"
[[1]]
[1] "Toyota"
[[1]]
[1] "Nissan"
```

10.6 Data-frame

A data frame is a table or a two-dimensional array-like structure in which each column contains values of one variable and each row contains one set of values from each column. A spreadsheet if you will. There MUST be the same number of values in each row and column. Cells cannot be empty and must al least have a logical constant which contains the missing value indicator *NA*.

Following are the characteristics of a data frame.

- The column names should be non-empty.
- The row names should be unique.
- The data stored in a data frame can be of numeric, factor or character type.
- Each column should contain same number of data items.

Here is an example.

```
> employee.data = data.frame(employee_id = c (1:5),
    employee_name = c("Áine","Dónal","Siobhán","Sinéad","Donnacha"),
    salary = c(623.3,515.2,611.0,729.0,843.25),
    start_date = as.Date(c("2012-01-01", "2013-09-23", "2014-11-15", "2014-05-11",
    "2015-03-27")),
    stringsAsFactors = FALSE
    )
```

• Print the data frame.

. .

>	employee.dat	ta		
	<pre>employee_id</pre>	employee_name	salary	<pre>start_date</pre>
1	1	Áine	623.30	2012-01-01
2	2	Dónal	515.20	2013-09-23
3	3	Siobhán	611.00	2014-11-15
4	4	Sinéad	729.00	2014-05-11
5	5	Donnacha	843.25	2015-03-27

•	Acce	ess elements from the frame.	
	> (1 2 3 4 5	data.frame(employee.data\$employee_name, employee.data.employee_name employee.da Áine 2012-01 Dónal 2013-09 Siobhán 2014-11 Sinéad 2014-09 Donnacha 2015-03	employee.data\$start_date) ta.start_date -01 -23 -15 -11 -27
	> (employee.data[1:2]	
	(employee_id employee_name	
	1	1 Alne 2 Dónal	
	3	3 Siobhán	
	4	4 Sinéad	
	5	5 Donnacha	
•	Extra	act first two rows.	
	~	employee data[1:2]	
	(employee_id employee_name salary start_	date
	1	1 Áine 623.3 2012-0	1-01
	2	2 Dónal 515.2 2013-0	9-23
•	Con	sider the following <i>R</i> dataframe.	
	> :	individual = c("a1", "a2", "a3", "a4")	
	> (age = c(15,13,16,12)	
	> 1	weight.class = c("high", "high", "low",	"high")
	> (df = data.frame(individual, age, weight	.class)
	> (df	
	-	individual age weight.class	
	1	al 15 high	
	2	a2 13 high	
	4	a4 12 high	
•	Use	<i>class()</i> or <i>str()</i> to look at how each variation	able is classified.
	> (class(individual)	
	[1]] "character"	
	> ([1]	class(age)] "numeric"	
	> ([1]	class(weight.class)] "character"	
	> (class(df)	

```
ass(at)
[1] "data.frame"
> str(individual)
chr [1:4] "a1" "a2" "a3" "a4"
> str(age)
num [1:4] 15 13 16 12
> str(weight.class)
    chr [1:4] "high" "high" "low" "high"
```

10.7 Indexing Data-frames

read.csv(): Imports data from a Comma Separated File (CSV) file.

head() and **tail()**: Return the first or last parts of a vector, matrix, table, data frame or function.

For demonstration purposes, import the *bird_egg.csv* file.

>	bird_egg =	read.	csv('Da	atas	ets/b	ird_egg.cs	v', header=TR	UE)
>	head(bird_e	egg)						
	individual	year	clutch	age	eggs	dist_food	fail_fledge	
1	rm∕bg	101	3	2	4	149	1	
2	wm/rb	97	3	1	3	63	Θ	
3	rb/bkm	107	3	2	4	NA	Θ	
4	bbk/bkm	108	3	7	3	NA	Θ	
5	wbk/ym	106	3	2	3	NA	1	
6	o/ym	103	3	2	4	164	Θ	
>	tail(bird_e	egg)						
	individua	al yea	ar cluto	ch ag	ge egg	gs dist_fo	od fail_fledg	е
82	25 rr/ <u>5</u>	jm 10	98	1	2	5	NA	1
82	26 vm/k	ov 10	9	1	1	4	NA	1
82	27 m/wy	/2 10	98	1	3	5	NA	1
82	28 hm/	/y 10	9	1	2	4	NA	1
82	29 /ł	nm 10	98	1	1	4	NA	0
83	80 wm/y	/r 10	06	1	4	4	NA	1

Reviewing the column for year is (Note: *head()* function used to limit the output to a useable output):

```
> head(bird_egg$year)
[1] 101 97 107 108 106 103
> head(bird_egg[,2])
[1] 101 97 107 108 106 103
```

The first five rows of age is either of these (Note: *head()* function used to limit the output to a useable output):

> bird_egg\$age[1:5]
[1] 2 1 2 7 2
> bird_egg[1:5,4]
[1] 2 1 2 7 2

10.8 Add a new column

Add a column of squared values (e.g. age two) to bird_egg?. It is a simple matter of naming it and assign values to it.

>	head(bird_e	egg)						
	individual	year	clutch	age	eggs	dist_food	fail_fledge	age.square
1	rm/bg	101	3	2	4	149	1	4
2	wm/rb	97	3	1	3	63	Θ	1
3	rb/bkm	107	3	2	4	NA	Θ	4
4	bbk/bkm	108	3	7	3	NA	Θ	49
5	wbk/ym	106	3	2	3	NA	1	4
6	o/ym	103	3	2	4	164	Θ	4

It is also possible to add to the data frame based on an ifelse() decision rule.

This example:

- Adds new column to the *bird_egg* data.frame and calls it *egg.factor*.
- Creates a 2-level categorical variable for number of eggs.

> bird_egg\$age.square = bird_egg\$age^2

```
> bird_egg$eggs.factor = ifelse(bird_egg$eggs > 3, 'few', 'many')
```

> h	ead(bird_e	gg)							
ind	ividual ye	ar clutch	age	eggs	dist_fo	od fail_fledge	age.square	eggs.f	actor
1	rm/bg	101	3	2	4	149	1	4	few
2	wm/rb	97	3	1	3	63	Θ	1	many
3	rb/bkm	107	3	2	4	NA	Θ	4	few
4	bbk/bkm	108	3	7	3	NA	Θ	49	many
5	wbk/ym	106	3	2	3	NA	1	4	many
6	o/ym	103	3	2	4	164	Θ	4	few

10.9 Single or double brackets for indexing?

Single and double square brackets. As is demonstrated below the single brackets extract a list whereas double brackets extract in a numeric vector style format.

```
> a = seq(1, 10, length.out = 10)
> b = seq(1, 10, length.out = 10)
> c = seq(1.75, 3.2, length.out = 10)
> d = seq(0.43, 1.6, length.out = 10)
> a = data.frame(w = a, x = b, y = c, z = d)
> m = a[1]
> n = a[[2]]
> 0 = a[3]
> p = a[[4]]
> class(m)
[1] "data.frame"
> class(n)
[1] "numeric"
> class(o)
[1] "data.frame"
> class(p)
[1] "numeric"
```

> typeof(m) [1] "list"	
> typeof(n) [1] "double"	
<pre>> typeof(o) [1] "list"</pre>	
> typeof(p) [1] "double"	
> m W 1 1 2 2 3 3 4 4 5 5 6 6 7 7 8 8 9 9 10 10	
> n [1] 1 2 3 4 5 6 7 8 9 10	0
<pre>> 0</pre>	
> p	

. [1] 0.43 0.56 0.69 0.82 0.95 1.08 1.21 1.34 1.47 1.60 67

10.10 Exercise: Data frame 1

Create a new third variable in the data frame (called *age.cat*) which is a categorical variable based on age where all individuals four or less are *young* and those greater than four are considered *old*.

• Answer:

```
> individual = 1:10
> age = c(3, 5, 7, 4, 9, 3, 5, 4, 8, 6)
> age.df = data.frame (individual, age)
> age.df$age.cat = ifelse(age.df$age < 5, 'young', 'old')</pre>
> age.df
  individual age age.cat
              3 young
1
          1
2
           2 5
                   old
3
           37
                    old
           4 4 young
4
              9
3
5
           5
                   old
           6
6
                  young
           75
7
                   old
                  young
8
          8 4
           98
9
                   old
10
          10 6
                     old
```

10.11 Changing names within the data

It is often necessary to adjust the names within the data. Follow the example below.

```
> names(age.df)
[1] "Individual" "age" "age.cat"
> names(age.df)[3] = "age2"
> age.df
  Individual age age2
     1 3 young
1
2
          2 5
                  old
             7
3
          3
                  old
4
          4
             4 young
5
          5
              9
                 old
             3 young
6
          6
7
          7
             5 old
8
          8
             4 young
9
          9
              8 old
10
         10
             6
                  old
```

10.12 Read files into R

read.csv() and read.csv2():

• *read.csv* and *read.csv2* are identical to *read.table* except for the defaults. They are intended for reading *comma separated value* files (*.csv*) or (*read.csv2*) the variant used in countries that use a comma as decimal point and a semicolon as field separator.

read.table():

• Reads a file in table format and creates a data frame from it, with cases corresponding to lines and variables to fields in the file.

read.delim(): - This is a wrapper function for *read.table()* with default argument values that are convenient when reading in tab-separated data.

10.13 English / European

Many European computer settings are different to the original *.csv* delimited file settings because the comma is used to denote fractions of numbers (decimals).

- 3.14 (English setting)
- 3,14 (European setting)

Thus European computers use semi-colons in their *comma separated* files and commas for decimals. This causes great confusion to *R* if import it the wrong way Open your csv file in a text editor and check how it looks.

- Use read.csv() for English settings
- Use read.csv2() for European settings

10.14 Set a working directory

Set a working directory or set file pathway to read in data files.

- setwd(): Is used to set the working directory to dir.
- get.wd(): Check what the working directory is set to.
- file.choose(): Choose a File Interactively.

```
> setwd('~/course_datasets')
```

```
> ind = read.csv('individual.csv')
```

```
> head(ind)
  individual trait
1
          а
                  3
2
           а
                  4
3
           g
                  2
4
                  6
           g
5
           g
                  5
6
           q
                  4
```

10.15 Checks after importing Data-frame

The first steps after importing data.frame are:

- 1. Check it looks like it should use *head()* & *tail()*.
- 2. Use *summary()* to get an overview of the data.
- 3. Use *str()* to check the structure of each variable.
- Fix variable classes if necessary with as.factor() or as.character(), as.numeric() or as.Date() functions.
- 5. Get the names of each variable using *names()* (tip: paste names in to your code for future reference).
- 6. Create the datasets you want to use for analysis by using *data.frame* indexing & filtering.

10.16 Removing missing values from a data set

Complete cases is a logical (boolean) function that returns *TRUE* for each observation (vectors) or row (data frame) that is complete (i.e. has no missing value / NA) for a data.frame called *data*.

```
> complete.data = data[complete.cases(data),]
```

0

10.16.1 Example:

Use the dataset *bird_egg.csv*.

```
> df = read.csv('bird_egg.csv')
```

```
> head(df)
```

	individual	year	clutch	age	eggs	dist_food	fail_fledge
1	rm/bg	101	3	2	4	149	1
2	wm/rb	97	3	1	3	63	Θ
3	rb/bkm	107	3	2	4	NA	Θ
4	bbk/bkm	108	3	7	3	NA	Θ
5	wbk/ym	106	3	2	3	NA	1

o/ym 103 3 2 4 164

>	summary(df)	
-	Summary (ur)	

6

individual		year			clutch		age		eggs		
rm/bg	:	21	Min.	: 9	7.0	Min.	:1.000	Min.	: 1.000	Min.	:1.000
bm/bw	:	20	1st	Qu.:10	2.0	1st Qu.	:1.000	1st Qu	.: 1.000	1st Qu.	:4.000
m∕ro	:	19	Medi	an :10	5.0	Median	:1.000	Median	: 2.000	Median	:4.000
bb/m	:	17	Mean	:10	4.7	Mean	:1.396	Mean	: 2.667	Mean	:4.032
bb/rm	:	16	3rd	Qu.:10	7.0	3rd Qu.	:2.000	3rd Qu	.: 4.000	3rd Qu.	:5.000
g/m	:	15	Max.	:10	9.0	Max.	:3.000	Max.	:10.000	Max.	:6.000
(Other)	:7	22						NA's	:1	NA's	:5
dist_	fo	od		fail_f	ledge						
Min.	:	8.00) M	in.	:0.000	0					
1st Qu.	:	10.00) 1	st Qu.	:0.000	0					
Median	:	70.00) M	edian	:1.000	0					
Mean	:	90.69	M	ean	:0.667	9					
3rd Qu.	:1	35.00) 3	rd Qu.	:1.000	0					
Max.	:4	34.00) M	ax.	:1.000	0					
NA's	:4	81	N	A's	:2						

```
> str(df)
```

'data.frame': 830 obs. of 7 variables:

\$ individual : Factor w/ 252 levels "","/gm","/hm",..: 162 214 152 11 204 130 234
238 55 214 ...

\$ year : int 101 97 107 108 106 103 97 97 100 98 ...

\$ clutch : int 3 3 3 3 3 3 3 3 3 3 3 3 ... \$ age : int 2 1 2 7 2 2 4 1 2 2 ... \$ eggs : int 4 3 4 3 3 4 3 3 2 3 ... \$ dist_food : int 149 63 NA NA NA 164 18 191 112 12 ... \$ fail_fledge: int 1 0 0 0 1 0 0 0 0 0 ...

```
> df$year = as.factor(df$year)
```

```
> df$fail_fledge = as.factor(df$fail_fledge)
```

```
> df$clutch = as.factor(df$clutch)
```

[6] "dist_food" "fail_fledge"

```
> str(df)
  'data.frame': 830 obs. of 7 variables:
   $ individual : Factor w/ 252 levels "","/gm","/hm",..: 162 214 152 11 204 130 234
238 55 214 ...
               : Factor w/ 13 levels "97","98","99",..: 5 1 11 12 10 7 1 1 4 2 ...
  $ year
  $ clutch : Factor w/ 3 levels "1", "2", "3": 3 3 3 3 3 3 3 3 3 3 ...
              : int 2127224122...
  $ age
  $ eggs : int 4 3 4 3 3 4 3 3 2 3 ...
$ dist_food : int 149 63 NA NA NA 164 18 191 112 12 ...
  $ fail_fledge: Factor w/ 2 levels "0","1": 2 1 1 1 2 1 1 1 1 1 ...
 > names(df)
  [1] "individual" "year"
                                  "clutch"
                                                 "age"
                                                               "eggs"
```

10.17 Data.frame object classes

Integer	as.integer()
Numeric	as.numeric()
Character	as.character()
Factor	as.factor()
Date	as.Date()
Logical	as.logical()

Examples to change object class.

```
> df$clutch = as.factor(df$clutch)
```

```
> df$individual = as.character(df$individual)
```

as.matrix() and as.data.frame() can be used for 2-dimensional objects.

10.18 Saving tables & Data Frames

write.table(): - prints its required argument 'x' (after converting it to a data frame if it is not one nor a matrix) to a file or connection.

write.csv(): - Does the same as write.table() except in comma delimited format.

These will save the data-frame or matrix to a file in the current working directory (use *get.wd()* to check where that is and *set.wd()* to change it.)

```
> write.csv(df.egg, "bird_egg2.csv")
```

10.19 subset() function

It is regular that a subset of a frame is necessary. The *subset()* function handles this event.

10.19.1 Columns subset

```
> names(df.egg)
[1] "individual"
                "year"
                             "clutch"
                                         "age"
                                                      "eggs"
[6] "dist_food"
                "fail_fledge"
> df.sub1 = subset(df.egg, select = c(individual, clutch, age, eggs))
> head(df.sub1)
 individual clutch age eggs
1
     rm/bq
              32
                        4
2
     wm/rb
               3 1
                        3
    rb/bkm
               32
3
                       4
  bbk/bkm
4
                3
                   7
                        3
                3 7
3 2
5
    wbk/ym
                        3
                3 2
                      4
6
      o/ym
```

Obviously the same could be achieved by indexing.

```
> df.sub2 = df.egg[, c(1,3,4,5)]
```

```
> head(df.sub2)
 character factors age eggs
1
    rm/bg 3 2 4
              3 1
3 2
2
    wm/rb
                      3
3
    rb/bkm
                      4
              3 7
 bbk/bkm
                     3
4
              32
5
   wbk/ym
                     3
               3 2
6
     o/ym
                      4
```

10.20 Date and time

Date and time for timestamps or dates and time within datasets can be useful. First looking at getting the time and date from the system with the *date()* function. The example demonstrates getting the date and time plus formatting them using the format types documented in the table below.

```
> t = Sys.time()
> print(t)
[1] "2018-09-28 10:45:41 EAT"
> str(t)
POSIXct[1:1], format: "2018-09-28 10:45:41
> t.short = format( x = t, format = '%d %b %Y')
> print(t.short)
[1] "28 Sep 2018"
> str(t.short)
chr "28 Sep 2018"
```

Strings of data that are in *character* or other formats can be converted to a *date* format. First ensure the data is in character format, then apply the *as.Date()* function to change to a date format.

```
> c.date = 'Fri 02 Sep 2018 - 11:02:12'
> c.date = as.character('Fri 02 Sep 2018 - 11:02:12')  # Must be 'chr' format
> str(c.date)
    chr "Fri 02 Sep 2018 - 11:02:12"
> c.date = as.Date(t, format = '%a %d %b %Y - %X')
> print(c.date)
[1] "2018-09-28"
> str(c.date)
Date[1:1], format: "2018-09-28"
> c.date.short = format( x = c.date, format = '%d %b %Y')
> print(c.date.short)
[1] "28 Sep 2018"
> str(c.date.short)
chr "28 Sep 2018"
```
For more granularity with time there is a specific *time()* function that creates the vector of times at which a time series was sampled.

Format types

- %a Locale's abbreviated weekday name.
- %A Locale's full weekday name.
- %b Locale's abbreviated month name.
- %B Locale's full month name.
- %c Locale's appropriate date and time representation.
- %C Century (a year divided by 100 and truncated to an integer) as a decimal number [00,99].
- %d Day of the month as a decimal number [01,31].
- %D Date in the format mm/dd/yy.
- %e Day of the month as a decimal number [1,31] in a two-digit field with leading space character fill.
- %h A synonym for %b.
- %H Hour (24-hour clock) as a decimal number [00,23].
- %I Hour (12-hour clock) as a decimal number [01,12].
- %j Day of the year as a decimal number [001,366].
- %m Month as a decimal number [01,12].
- %M inute as a decimal number [00,59].
- %n A <newline>.
- %p Locale's equivalent of either AM or PM.
- %r 12-hour clock time [01,12] using the AM/PM notation; in the POSIX locale, this shall be equivalent to %I : %M : %S %p.
- %S Seconds as a decimal number [00,60].
- %t A <tab>.
- %T 24-hour clock time [00,23] in the format HH:MM:SS.
- %u Weekday as a decimal number [1,7] (1=Monday).
- Week of the year (Sunday as the first day of the week) as a decimal number
 [00,53]. All days in a new year preceding the first Sunday shall be considered to be in week 0.

%V Week of the year (Monday as the first day of the week) as a decimal number [01,53]. If the week containing January 1 has four or more days in the new year, then it shall be considered week 1; otherwise, it shall be the last week of the previous year, and the next week shall be week 1.

- %w Weekday as a decimal number [0,6] (0=Sunday).
- Week of the year (Monday as the first day of the week) as a decimal number
 W [00,53]. All days in a new year preceding the first Monday shall be considered to be in week 0.
- %x Locale's appropriate date representation.
- %X Locale's appropriate time representation.
- %y Year within century [00,99].
- %Y Year with century as a decimal number.
- %Z Timezone name, or no characters if no timezone is determinable.

10.21 lapply()

Another function from the *apply()* functions.

The list apply *lapply()* function operates like apply *()* with similar output. The *lapply()* function can operate on other objects like dataframes and lists. It returns a list of the same length as X, each element of which is the result of applying *FUN* to the corresponding element of X.

- **lapply():** returns a vector or array or list of values obtained by applying a function to margins of an array or matrix.
- apply(X, FUN, ...)
 - X: an array, including a matrix.
 - *FUN*: the function.

This example shows the *mean()* function applied to the elements of the vector at position three of the list.

```
> l = list("Orange", "Green", c(43,13,61), TRUE, 42.18, 218.2)
(1,"[", , 3)
> str(1)
List of 6
$ : chr "Orange"
$ : chr "Green"
$ : num [1:3] 43 13 61
$ : logi TRUE
$ : num 42.2
$ : num 218
> lapply(1[3],mean)
[[1]]
[1] 39
> df = data.frame(employee_id = c (1:5),
       employee_name = c("Áine", "Dónal", "Siobhán", "Sinéad", "Donnacha"),
       salary = c(623.3,515.2,611.0,729.0,843.25),
      start_date = as.Date(c("2012-01-01", "2013-09-23", "2014-11-15", "2014-05-
      11", "2015-03-27")),
      stringsAsFactors = FALSE
      )
> df
  employee_id employee_name salary start_date
1
           1 Áine 623.30 2012-01-01
           2
                     Dónal 515.20 2013-09-23
2
                  Siobhán 611.00 2014-11-15
3
           3
4
           4
                   Sinéad 729.00 2014-05-11
                 Donnacha 843.25 2015-03-27
5
           5
> lapply(df[3], sum)
$salary
[1] 3321.75
```

10.22 User-defined Functions

One of the great strengths of any programming language is the ability to add functions, R is no different. A function should be considered if there is any block of code that is being repeated in a script. Instead put the block of code in a function, feed the function values and return results.

```
# Defining function above main block of code
new_function_name = function(arg1, arg2, ...){
    statements
    return(object)
}
# Call function from main block of code
new_function_name(arg1, arg2, ...)
```

The following script demonstrates how a user-defined function works. Create the *my_function_demo.R* file, make it executable and run it. The steps breakdown:

- 1. The existing set of objects in *R* are cleared.
- 2. Five vector objects are created and the last four are added to a dataframe.
- 3. The user-defined function is defined.
- 4. The main program follows:
 - A loop of four sends the v vector plus the each vector from the dataframe in turn as a list to the user-defined function.
 - The function processes and returns the linear model to where it was called from.
 - This response is assigned to the object model<loop #>. In this way four models are defined.
 - The *model<loop #>* names are extracted from the list of objects.
 - A loop through these models and their summary is output to standard out.

```
$ cat << EOM >> my_function_demo.R
#!/usr/bin/Rscript
```

```
# Clear objects from R
rm(list = ls())
# Define objects
v = 1:8
v1 = c(51, 19, 43, 74, 45, 26, 83, 42)
v2 = c(101, 111, 112, 123, 141, 152, 193, 141)
v3 = c(214, 233, 234, 226, 237, 248, 269, 276)
v4 = c(322, 354, 385, 377, 381, 314, 425, 416)
df = data.frame(v1, v2, v3, v4)
# Function 'realmod'
realmod = function(arg1, arg2){
    x = as.numeric(unlist(arg1));
    y = as.numeric(unlist(arg2));
    return(lm(y ~ x))
}
# Main program (calling the realmod function)
for (x in 1:4){
    assign(paste0('model',x), realmod(v,df[x]));
}
```

```
# Get list objects
obj_list = ls()
mod_list = grep ('model', obj_list)
# Loop through models and output to stdout
for (x in mod_list){
    cat(sprintf('%s',obj_list[x]),'\n');
    cat(strrep("=",6), "\n");
    print(summary(get(obj_list[x])))
}
# End script
quit(status = 0)
EOM
$ chmod +x my_function_demo.R
$ ./my_function_demo.R
model1
======
Call.
lm(formula = y \sim x)
Residuals:
            1Q Median
                            3Q
   Min
                                   Мах
-25.036 -15.839 -2.821 14.670 29.857
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 38.393 17.764 2.161 0.0739.
                        3.518 0.599 0.5711
              2.107
х
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 22.8 on 6 degrees of freedom
Multiple R-squared: 0.05643, Adjusted R-squared: -0.1008
F-statistic: 0.3588 on 1 and 6 DF, p-value: 0.5711
model2
======
Call:
lm(formula = y \sim x)
Residuals:
           1Q Median
                        3Q
   Min
                                   Мах
-27.750 -6.607 1.321 2.107 34.107
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 89.893 14.384 6.250 0.000778 ***
              9.857
                         2.848 3.461 0.013459 *
Х
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 18.46 on 6 degrees of freedom
Multiple R-squared: 0.6662, Adjusted R-squared: 0.6106
F-statistic: 11.98 on 1 and 6 DF, p-value: 0.01346
model3
======
Call:
lm(formula = y \sim x)
```

Data Analytics

```
Residuals:
Min 1Q Median 3Q Max
-12.155 -6.801 1.726 6.319 10.726
Coefficients:
      Estimate Std. Error t value Pr(>|t|)
(Intercept) 206.393 6.989 29.531 1e-07 ***
                       1.384 5.737 0.00122 **
             7.940
х
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 8.97 on 6 degrees of freedom
Multiple R-squared: 0.8458, Adjusted R-squared: 0.8201
F-statistic: 32.91 on 1 and 6 DF, p-value: 0.001218
model4
======
Call:
lm(formula = y \sim x)
Residuals:
  Min
          1Q Median 3Q
                                 Мах
-72.107 -0.714 8.107 14.964 29.321
Coefficients:
         Estimate Std. Error t value Pr(>|t|)
(Intercept) 328.679 27.290 12.044 1.99e-05 ***
            9.571
                       5.404 1.771
                                        0.127
х
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 35.02 on 6 degrees of freedom
Multiple R-squared: 0.3433, Adjusted R-squared: 0.2339
F-statistic: 3.137 on 1 and 6 DF, p-value: 0.1269
```

10.23 Recap exercises

10.23.1 Exercise: Data-frames 2

- 1. Import the <u>bird_egg.csv</u> data and call the data frame df.egg.
- 2. Explore the data frame and check structure change year, fail_fledge and clutch to factors, & individual to character.
- 3. Create new column in *df.egg* called *constant* consisting of a column of *1*s.
- 4. Create a new data frame (by indexing) consisting of *individual*, *clutch*, *age* & *egg* and call it *df.sub1*.
- 5. Create a new data.frame (called *df.sub2*) which consists of all data from first clutches for birds less than three years old.
- 6. Create a new data.frame (called df.sub3) which contains all data for which there are no missing values...
 - HINT: complete.cases().
- Calculate the mean distance to food for failed versus fledged nests. Do this for all data.frames (df.sub1, 2 & 3)....
 - HINT: check *help(mean)* if having problems.
- 8. Create a new column (*dist.cat*) that is a three-level distance category based on the *dist_food* column. Where <100 is *near*, 100-200 is *mid*, and >200 is *far*.
 - HINT: *ifelse(..., ..., ifelse(..., ..., ...)*).

 Create *df.sub4* which contains all records without missing values in order of bird age (youngest to oldest)

• HINT: df.sub3[order(),].

- 10.Rename the *individual* column to *id* & rename the *dist_food* column to *distance.food*.
- 11.Save the new data.frames as *.csv* files in your working directory folder (e.g. *df.sub1.csv* etc).

Answer:

```
clutch
                                             age
                                                            eggs
             Min. : 97.0 Min. :1.000 Min. : 1.000 Min. :1.000
bm/bw : 20 1st Qu.:102.0 1st Qu.:1.000 1st Qu.: 1.000
                                                        1st Qu.:4.000
m/ro : 19 Median :105.0 Median :1.000 Median : 2.000 Median :4.000
bb/m : 17 Mean :104.7
                         Mean :1.396 Mean : 2.667 Mean :4.032
                                         3rd Qu.: 4.000
bb/rm : 16
            3rd Qu.:107.0
                          3rd Qu.:2.000
                                                        3rd Qu.:5.000
            Max. :109.0 Max. :3.000
g/m
      : 15
                                         Max. :10.000
                                                        Max. :6.000
                                         NA's :1
(Other):722
                                                        NA's
                                                              :5
  dist_food
                fail_fledge
Min. : 8.00 Min. :0.0000
1st Qu.: 10.00 1st Qu.:0.0000
Median : 70.00
               Median :1.0000
Mean : 90.69
               Mean :0.6679
3rd Qu.:135.00
               3rd Qu.:1.0000
Max. :434.00
               Max. :1.0000
NA's
       :481
               NA's
                    :2
> str(df.egg)
'data.frame': 830 obs. of 7 variables:
 $ individual : Factor w/ 252 levels "","/gm","/hm",..: 162 214 152 11 204 130 234
238 55 214 ...
           : int 101 97 107 108 106 103 97 97 100 98 ...
$ year
$ clutch
           : int 3333333333...
$ age
           : int 2127224122...
            : int 4343343323...
$ eggs
$ dist_food : int 149 63 NA NA NA 164 18 191 112 12 ...
$ fail_fledge: int 1000100000...
> names(df.egg)
[1] "individual"
               "year"
                                        "age"
                                                    "eggs"
                            "clutch"
```

```
[6] "dist_food"
               "fail_fledge"
> head(df.egg)
 individual year clutch age eggs dist_food fail_fledge
1
     rm/bg 101
                32
                         4
                                 149
                                             1
                  31
2
     wm/rb
           97
                          3
                                  63
                                             0
                  32
    rb/bkm 107
                                             0
3
                         4
                                  NA
                 3 7 3
4
    bbk/bkm 108
                                 NA
                                             0
                 32
32
                         3
5
    wbk/ym 106
                                 NΑ
                                             1
      o/ym 103
6
                          4
                                 164
                                             0
```

```
###### // Change year, fail fledge and clutch to factors //
> str(df.egg)
'data.frame': 830 obs. of 8 variables:
 $ individual : Factor w/ 252 levels "","/gm","/hm",..: 162 214 152 11 204 130 234
238 55 214 ...
            : int 101 97 107 108 106 103 97 97 100 98 ...
$ year
             : int 3333333333...
$ clutch
$ age
             : int 2127224122...
$ eggs
             : int 4343343323...
$ dist_food : int 149 63 NA NA NA 164 18 191 112 12 ...
$ fail_fledge: int 1000100000...
> df.egg[, 'year'] = as.factor(df.egg[, 'year'])
> str(df.egg$year)
Factor w/ 13 levels "97", "98", "99", ..: 5 1 11 12 10 7 1 1 4 2 ...
> df.egg[, 'fail_fledge'] = as.factor(df.egg[, 'fail_fledge'])
> str(df.egg$fail_fledge)
Factor w/ 2 levels "0", "1": 2 1 1 1 2 1 1 1 1 1 ...
###### // Change individual to character //
> df.egg[, 'individual'] = as.character(df.egg[, 'individual'])
> str(df.egg$individual)
chr [1:830] "rm/bg" "wm/rb" "rb/bkm" "bbk/bkm" "wbk/ym" "o/ym" "yg/m" ...
###### // Create new column in df.egg called 'constant' made of 1s //
> df.egg$constant = 1
> head(df.egg)
 individual year clutch age eggs dist_food fail_fledge constant
1
      rm/bg 101
                      3
                         2
                               4
                                       149
                                                    1
                                                             1
                      3 1
2
      wm/rb
             97
                               3
                                       63
                                                    0
                                                             1
                      3 2
3
     rb/bkm 107
                                                    0
                                       NA
                                                             1
                               4
4
    bbk/bkm 108
                      3 7
                               3
                                       NA
                                                    0
                                                             1
5
     wbk/ym 106
                      3
                        2
                               3
                                       NA
                                                    1
                                                             1
6
       o/ym 103
                      3 2
                               Δ
                                       164
                                                    0
                                                             1
###### // Create a new data frame df.sub1 with individual, clutch, age & eggs //
> names(df.egg)
[1] "individual"
                "year"
                               "clutch"
                                             "age"
                                                          "eggs"
[6] "dist_food"
                 "fail_fledge" "constant"
> df.sub1 = df.egg[, c(1,3,4,5)]
> head(df.sub1)
 individual clutch age eggs
1
      rm/bg
                 3
                    2
                          4
2
      wm/rb
                 3
                     1
                          3
3
     rb/bkm
                 3
                     2
                          4
4
    bbk/bkm
                 3
                     7
                          3
5
     wbk/ym
                 3
                     2
                          3
6
       o/ym
                 3
                     2
                          4
```

// df.sub2 to consists of all data for birds < 3 //

```
> df.sub2 = df.egg[df.egg$age < 3,]</pre>
```

```
> head(df.sub2)
  individual year clutch age eggs dist_food fail_fledge constant
       rm/bg 101
                      32
1
                                 Λ
                                         149
                                                        1
                                                                 1
2
       wm/rb
               97
                       3
                                 3
                                          63
                                                        0
                                                                 1
                            1
3
      rb/bkm
              107
                       3
                            2
                                 4
                                          NA
                                                        0
                                                                 1
5
      wbk/ym
              106
                       3
                            2
                                 3
                                          NA
                                                        1
                                                                 1
6
        o/ym
              103
                       3
                                         164
                                                        0
                                                                 1
                            2
                                 4
8
        ym/b
               97
                       3
                            1
                                 3
                                         191
                                                        0
                                                                 1
###### // df.sub3 to contain all data with no missing values //
> df.sub3 = df.egg[complete.cases(df.egg),]
> head(df.sub3)
  individual year clutch age eggs dist_food fail_fledge constant
1
       rm/bg 101
                       3 2
                                 4
                                         149
                                                        1
                                                                 1
2
       wm/rb
               97
                       3
                            1
                                 3
                                          63
                                                        0
                                                                 1
6
        o/ym
              103
                       3
                            2
                                 4
                                         164
                                                        0
                                                                 1
7
        yg/m
               97
                       3
                            4
                                 3
                                          18
                                                        0
                                                                 1
8
        ym/b
               97
                       3
                            1
                                 3
                                         191
                                                        0
                                                                 1
9
        bw/m
             100
                       3
                            2
                                 2
                                         112
                                                        0
                                                                 1
###### // Calculate the mean distance to food for failed versus fledged nests //
Note: cannot do df.dub1 as it doesn't have a 'fail fledge' column.
> mean(df.sub2[df.sub2$fail_fledge == 0 & complete.cases(df.sub2$dist_food),6])
[1] 118.2319
> mean(df.sub2[df.sub2$fail_fledge == 1 & complete.cases(df.sub2$dist_food),6])
[1] 93.60135
> mean(df.sub3[df.sub3$fail_fledge == 0 & complete.cases(df.sub3$dist_food),6])
[1] 96.70642
> mean(df.sub3[df.sub3$fail_fledge == 1 & complete.cases(df.sub3$dist_food),6])
[1] 87.9625
###### // 3-level distance category based on the 'dist_food' column //
> df.egg$dist.cat = ifelse(
                      (df.egg$dist_food < 100),</pre>
                      print ('near'),
                      ifelse(
                        (df.egg$dist_food > 100 & df.egg$dist_food < 200),</pre>
                        print ('mid'),
                        ifelse(
                          (df.egg$dist_food > 200),
                          print ('far'),
                       "NA"
                      )
                    )
                 )
[1] "near"
[1] "mid"
[1] "far"
> head(df.egg)
  individual year clutch age eggs dist_food fail_fledge constant dist.cat
1
       rm/bg
             101
                       3
                            2
                                 4
                                         149
                                                        1
                                                                 1
                                                                        mid
2
       wm/rb
               97
                                 3
                                          63
                                                                 1
                       3
                            1
                                                        Θ
                                                                        near
3
      rb/bkm
              107
                       3
                                                        0
                                                                 1
                                                                        <NA>
                            2
                                 4
                                          NA
     bbk/bkm 108
                       3
                            7
                                 3
                                          NA
                                                        0
                                                                        <NA>
4
                                                                 1
5
                       3 2
                                                                        <NA>
      wbk/ym 106
                                 3
                                          NA
                                                        1
                                                                 1
        o/ym 103
6
                       3
                            2
                                 4
                                         164
                                                        0
                                                                 1
                                                                        mid
```

// df.sub4 contains all records without missing values in order of bird age //

```
> temp = df.egg[complete.cases(df.egg),]
> df.sub4 = temp[order(temp$age),]
> head(df.sub4)
  individual year clutch age eggs dist_food fail_fledge constant dist.cat
2
       wm/rb 97
                     3 1 3
                                       63
                                                     0
                                                                   near
                                                             1
                      3 1
                                       191
                                                     0
                                                             1
        ym/b 97
                               3
                                                                    mid
8
                         1 4
15
        m/ro
              99
                      3
                                       127
                                                     1
                                                             1
                                                                    mid
27
        bg/m
              100
                       2
                          1
                               4
                                       135
                                                     0
                                                             1
                                                                    mid
                       2 1
28
       rm/bg 100
                               4
                                       150
                                                     Θ
                                                             1
                                                                    mid
                       2 1
31
       wm/rb
              97
                               4
                                        32
                                                     0
                                                             1
                                                                   near
###### // rename 'individual' --> 'id', 'dist_food' --> 'distance.food' //
> names(df.eqq)
[1] "individual"
                 "year"
                               "clutch"
                                            "age"
                                                          "eggs"
[6] "dist_food"
                 "fail_fledge" "constant"
                                            "dist.cat"
> names(df.egg)[1] = "id"
> names(df.egg)[6] = "distance.food"
> names(df.egg)
[1] "id"
                   "vear"
                                                  "age"
                                  "clutch"
[5] "eggs"
                   "distance.food" "fail_fledge"
                                                  "constant"
[9] "dist.cat"
> write.csv(df.sub1, file = "df.sub1.csv")
> write.csv(df.sub2, file = "df.sub2.csv")
> write.csv(df.sub3, file = "df.sub3.csv")
> write.csv(df.sub4, file = "df.sub4.csv")
```

10.23.2 I

Exercise: Data-frames 3

- 1. Create a new folder that contains the *owl_data.csv* data file.
- 2. Set your working directory to the new folder.
- 3. Import the *owl_data.csv* data and call the data frame owl.
- 4. Explore the data frame and check structure [e.g. by using summary(), str(), names(), head()].
- 5. Create a column of 1's and 0's called *sex10* where *male=0* & *female=1* (from the *sex* column).
- 6. Change the sex10 column to a factor
 - HINT: as.factor().
- 7. Subset the data.frame to only include data from broods with five chicks.
- 8. Add 2 columns to the *owl data.frame*.
 - food.category: where food is a 2-category variable *low* (food less than 25) & *high* (food greater than or equal to 25).
 - begging.3: where begging is a 3-category variable 1 (between 0-10), 2 (from 10-20) and 3 (above 20).
 - HINT: you can nest *ifelse* functions within *ifelse* functions.
- 9. Save the new data.frame as a .csv file called owl_2.csv.

```
###### // Inport the owl.csv file //
> setwd('~/datasets/owl_data')
> owl = read.csv('owl_data.csv')
###### // Explore the data frame //
> summary(owl)
         nest
                     sex
                                  food
                                                begging
                                                                brood
         : 52 Female:245
                             Min. :21.71 Min. : 0.00 Min. :1.000
Oleyes
           : 41 Male :354
                              1st Qu.:23.11 1st Qu.: 0.00
Moutet
                                                            1st Qu.:4.000
Etrabloz : 34
                              Median :24.38 Median : 5.00
                                                            Median :4.000
Yvonnand
         : 34
                              Mean :24.76 Mean : 6.72 Mean :4.392
                              3rd Qu.:26.25
Champmartin: 30
                                             3rd Qu.:11.00
                                                            3rd Qu.:5.000
        : 29
Lucens
                              Max. :29.25 Max. :32.00 Max. :7.000
 (Other)
           :379
> str(owl)
'data.frame': 599 obs. of 5 variables:
$ nest : Factor w/ 27 levels "AutavauxTV", "Bochet", ...: 1 1 1 1 1 1 1 1 1 1 ...
         : Factor w/ 2 levels "Female", "Male": 2 2 2 2 2 2 2 1 2 1 ...
$ Sex
$ food
        : num 22.2 22.4 22.5 22.6 22.6 ...
$ begging: int 4 0 2 2 2 2 18 4 18 0 ...
$ brood : int 555555555...
> names(owl)
             "sex"
                      "food"
                                "begging" "brood"
[1] "nest"
> head(owl)
       nest sex food begging brood
1 AutavauxTV Male 22.25
                           4
                                 5
2 AutavauxTV Male 22.38
                            0
                                  5
3 AutavauxTV Male 22.53
                            2
                                  5
4 AutavauxTV Male 22.56
                            2
                                 5
5 AutavauxTV Male 22.61
                            2
                                  5
6 AutavauxTV Male 22.65
                            2
                                  5
##### // Create the 'sex10' column //
> owl$sex10 = ifelse(owl$sex == 'Male', 0, 1)
> head(owl)
       nest sex food begging brood sex10
1 AutavauxTV Male 22.25 4 5
                                       0
2 AutavauxTV Male 22.38
                           0
                                 5
                                       0
                          2
3 AutavauxTV Male 22.53
                                5
                                       0
                          2
                                5
4 AutavauxTV Male 22.56
                                       0
5 AutavauxTV Male 22.61
                            2
                                 5
                                       0
6 AutavauxTV Male 22.65
                            2
                                  5
                                       0
###### // Change the 'sex10' column values to factors //
> owl[, 'sex10'] = as.factor(owl[, 'sex10'])
> str(owl$sex10)
Factor w/ 2 levels "0", "1": 1 1 1 1 1 1 1 2 1 2 ...
###### // Subset to only include data from broods with 5 chicks //
> owl.5chicks = owl[owl$brood == 5,]
```

```
###### // add 'food.category' and 'begging.3' columns //
> owl$food.category = ifelse(owl$food < 25, 'low', 'high')</pre>
> head(owl)
         nest sex food begging brood sex10 food.category
1 AutavauxTV Male 22.25 4 5 0
                                                                  low
2 AutavauxTV Male 22.38
                                  0 5 0
                                                                  low

      3 AutavauxTV Male 22.53
      2
      5
      0

      4 AutavauxTV Male 22.56
      2
      5
      0

      5 AutavauxTV Male 22.61
      2
      5
      0

      6 AutavauxTV Male 22.65
      2
      5
      0

3 AutavauxTV Male 22.53
                                                                 low
                                                                  low
                                                                  low
                                                                  low
> owl$begging.3 = ifelse(
                     (owl$begging < 10),
                     print ('1'),
                     ifelse(
                       (owl$begging > 9 & owl$begging < 20),
                       print ('2'),
                       ifelse(
                          (owl$begging > 19),
                         print ('3'),
                       "NA"
                       )
                     )
                 )
[1] "1"
[1] "2"
[1] "3"
> head(owl)
         nest sex food begging brood sex10 food.category begging.3
1 AutavauxTV Male 22.25 4 5 0
2 AutavauxTV Male 22.38 0 5 0
                                                                 low
                                                                                1
                                                                 low
                                                                                1
                                  2 5 0
2 5 0
2 5 0
2 5 0
2 5 0
3 AutavauxTV Male 22.53
                                                                 low
                                                                               1
4 AutavauxTV Male 22.56
                                                                low
                                                                               1
5 AutavauxTV Male 22.61
                                                                 low
                                                                                1
6 AutavauxTV Male 22.65
                                                                  low
                                                                                 1
##### // Dave the data //
> write.csv(owl, file = "owl_2.csv")
```

10.23.3 Exercise: Data-frames 4

- 1. Create a new folder that contains the *individual.csv* data file.
- 2. Set your working directory to the new folder.
- 3. Import the *individual.csv* data and call the data frame ind.
- 4. Explore the data frame and check structure. [e.g. by using *summary()*, *str()*, *names()*, *head()*].
- 5. Create a new column (*id*) where the individual names from the first column are replaced by numbers starting at one and increasing sequentially. Note that individuals are repeated throughout the dataset so you'll have to make sure each number always matches the same individual.
 - HINT: *str()* & *as.numeric()*.
- 6. Save the new data.frame as a .csv file called ind_2.csv.

```
###### // Inport the individual.csv file //
> setwd('~/datasets/individual')
> ind = read.csv('individual.csv')
###### // Explore the data frame and check structure.
> summary(ind)
  individual
                 trait
 g
       :9
             Min. :2.000
             1st Qu.:4.000
 n
       :6
           Median :6.000
 ee
       :5
 h
           Mean :5.513
       :5
 а
       :3
           3rd Qu.:7.000
       :3
             Max. :9.000
 q
 (Other):8
> str(ind)
'data.frame': 39 obs. of 2 variables:
 $ individual: Factor w/ 10 levels "a","ee","g","h",..: 1 1 3 3 3 3 3 3 3 4 ...
            : int 3426546754...
 $ trait
> names(ind)
[1] "individual" "trait"
> head(ind)
 individual trait
1
          а
                3
2
                4
          а
3
                2
          g
4
          g
                6
5
                5
          g
6
          g
                4
##### // Create column 'id' //
> ind$id = as.numeric(ind[,1])
> ind
   individual trait id
1
           а
               3 1
2
                 4
           а
                    1
3
                 2
                    3
           g
                    3
4
           g
                 6
5
                 5
                    3
           g
6
                 4
                    3
           g
                 63
7
           g
8
                 7
                    3
           g
9
                 5
                    3
           g
10
           h
                 4
                    4
                 5
                    6
11
           n
12
                 7
                    1
           а
13
           n
                 2
                    6
14
           n
                 6
                    6
                    6
15
                 7
           n
                 2
                    6
16
           n
17
           n
                 9
                    6
18
          ee
                 8
                    2
19
          ee
                 6
                    2
20
          ee
                 7
                    2
21
                 8 2
          ee
22
           S
                 9
                    8
23
           h
                 6
                    4
                    4
24
           h
                 4
```

25	g	3	3	
26	g	6	3	
27	ee	7	2	
28	h	8	4	
29	h	6	4	
30	q	5	7	
31	q	4	7	
32	q	3	7	
33	t	6	9	
34	t	7	9	
35	У	8	10	
36	У	8	10	
37	t	5	9	
38	m	4	5	
39	m	3	5	

//Save the new as 'ind_2.csv' //

> write.csv(ind, file = "ind_2.csv")

10.23.4 Exercise: Data-frames 5

- 1. Create a new folder that contains the *RIKZ.csv* data file.
- 2. Set your working directory to the new folder.
- 3. Import the data (rikz), explore and check structure.
- 4. reorder the rows of the entire data.frame by increasing values of NAP
 HINT: rikz[order()].
- 5. create a data.frame called *rikz2* that only contains data from beaches one to four for the first week of data collection.
- 6. what is the mean chalk value for Beach 5 and the mean grain size for beach eight?.
- 7. save the new data.frame as .csv file called rikz_2.csv.

// Inport the RIKZ.csv dataset file //

```
> setwd('~/datasets/RIKZ')
```

```
> rikz = read.csv('RIKZ.csv')
```

// Import the data (rikz), explore and check structure //

```
> summary(rikz)
```

Sample	Richness	Week	angle1	angle2
Min. : 1	Min. : 0.000	Min. :1.000	Min. : 6.00	Min. :21.00
1st Qu.:12	1st Qu.: 3.000	1st Qu.:2.000	1st Qu.: 22.00	1st Qu.:32.00
Median :23	Median : 4.000	Median :2.000	Median : 32.00	Median :42.00
Mean :23	Mean : 5.689	Mean :2.333	Mean : 50.31	Mean :57.78
3rd Qu.:34	3rd Qu.: 8.000	3rd Qu.:3.000	3rd Qu.: 55.00	3rd Qu.:89.00
Max. :45	Max. :22.000	Max. :4.000	Max. :312.00	Max. :96.00
exposure	salinity	temperature	NAP	
Min. : 8.00) Min. :26.4	Min. :15.80	Min. :-1.3360	
1st Qu.:10.00) 1st Qu.:27.1	1st Qu.:17.50	1st Qu.:-0.3750	
Median :10.00) Median :27.9	Median :18.77	Median : 0.1670	
Mean :10.22	2 Mean :28.1	Mean :18.77	Mean : 0.3477	
3rd Qu.:11.00) 3rd Qu.:29.4	3rd Qu.:20.00	3rd Qu.: 1.1170	
Max. :11.00) Max. :29.9	Max. :20.80	Max. : 2.2550	
penetrability	/ grainsize	humus	chalk	
Min. :151.8	3 Min. :186.0	Min. :0.000	00 Min. : 0.8	50
1st Qu.:237.1	L 1st Qu.:222.5	1st Qu.:0.000	00 1st Qu.: 2.20	90
Median :256.1	L Median :266.0	Median :0.050	00 Median : 4.7	50
Mean :289.4	4 Mean :272.5	Mean :0.050	28 Mean : 7.90	61

3rd Qu.: 9.150

```
Max.
        :624.0
                  Max. :405.5
                                  Max. :0.30000
                                                    Max.
                                                          :45.750
     sorting1
                       Beach
  Min. : 53.08
                   Min. :1
  1st Qu.: 72.75
                   1st Qu.:3
  Median : 89.03
                   Median :5
  Mean : 97.82
                   Mean :5
  3rd Qu.:115.44
                   3rd Qu.:7
  Max.
         :248.60
                   Max.
                         :9
  > str(rikz)
  'data.frame': 45 obs. of 15 variables:
  $ Sample
                : int 12345678910...
  $ Richness
                 : int 11 10 13 11 10 8 9 8 19 17 ...
  $ Week
                 : int 1111111111...
  $ angle1
                 : int 32 62 65 55 23 129 126 52 26 143 ...
  $ angle2
                 : int 96 96 96 96 96 89 89 89 89 89 ...
  $ exposure
                 : int 10 10 10 10 10 8 8 8 8 8 ...
  $ salinity
                 : num
                        29.4 29.4 29.4 29.4 29.4 29.6 29.6 29.6 29.6 29.6 ...
  $ temperature : num 17.5 17.5 17.5 17.5 17.5 20.8 20.8 20.8 20.8 20.8 ...
  $ NAP
                 : num 0.045 -1.036 -1.336 0.616 -0.684 ...
  $ penetrability: num 254 227 237 249 252 ...
  $ grainsize
               : num 222 200 194 221 202 ...
  $ humus
                 : num 0.05 0.3 0.1 0.15 0.05 0.1 0.1 0.1 0.15 0 ...
  $ chalk
                 : num 2.05 2.5 3.45 1.6 2.45 2.5 1.85 1.7 2.3 2.6 ...
  $ sorting1
                  : num 69.8 59 59.2 67.8 57.8 ...
                 : int 1111122222...
  $ Beach
  > names(rikz)
                      "Richness"
                                      "Week"
                                                      "angle1"
  [1] "Sample"
   [5] "angle2"
                      "exposure"
                                      "salinity"
                                                      "temperature"
  [9] "NAP"
                       "penetrability" "grainsize"
                                                      "humus"
  [13] "chalk"
                      "sorting1"
                                      "Beach"
 > head(rikz)
    Sample Richness Week angle1 angle2 exposure salinity temperature
                                                                       NAP
                                   96
                                                   29 4
                                                               17.5 0.045
 1
        1
                11
                      1
                            32
                                           10
 2
        2
                10
                            62
                                   96
                                            10
                                                   29.4
                                                               17.5 -1.036
                      1
 3
        3
                13
                      1
                            65
                                   96
                                            10
                                                   29.4
                                                               17.5 -1.336
 4
        4
                11
                      1
                            55
                                   96
                                            10
                                                   29.4
                                                               17.5 0.616
                                            10
                                                               17.5 -0.684
 5
                10
                      1
                            23
                                   96
                                                   29.4
        5
 6
        6
                 8
                      1
                           129
                                   89
                                             8
                                                   29.6
                                                               20.8 1.190
   penetrability grainsize humus chalk sorting1 Beach
 1
           253.9
                     222.5 0.05 2.05
                                         69.830
                                                    1
 2
           226.9
                     200.0 0.30
                                  2.50
                                         59.000
                                                    1
 3
           237.1
                     194.5 0.10
                                  3.45
                                         59.220
                                                    1
 4
           248.6
                     221.0 0.15
                                  1.60
                                         67.750
                                                    1
 5
           251.9
                     202.0 0.05
                                 2.45
                                         57.760
                                                    1
  6
           250.1
                     192.5 0.10 2.50
                                         53.075
                                                    2
  ###### // Reorder the rows of the entire data.frame by increasing values of NAP.
HINT: rikz[order()] //
 > rikz = rikz[order(rikz$NAP),]
 > head(rikz)
    Sample Richness Week angle1 angle2 exposure salinity temperature
                                                                        NAP
  3
         3
                 13
                       1
                             65
                                    96
                                             10
                                                    29.4
                                                                17.5 -1.336
  10
         10
                 17
                       1
                            143
                                    89
                                              8
                                                    29.6
                                                                20.8 -1.334
  2
                             62
                                    96
                                             10
                                                    29 4
                                                                17.5 -1.036
         2
                 10
                       1
  38
                  7
                             55
                                    32
                                             10
                                                    26.4
                                                               20.0 -1.005
        38
                       3
                       2
                             41
                                    42
                                                    27.9
                                                               15.8 -0.976
 11
        11
                  6
                                             11
                       3
                                    36
  29
        29
                  6
                             48
                                             11
                                                    27.1
                                                                17.4 -0.893
    penetrability grainsize humus chalk sorting1 Beach
                      194.5 0.10
  3
            237.1
                                   3.45
                                          59.220
                                                     1
            257.9
                      197.0 0.00
  10
                                   2.60
                                          59.575
                                                     2
```

3rd Ou.:0.10000

3rd Qu.:272.9

3rd Qu.:316.5

2 226.9 200.0 0.30 2.50 59.000 1 38 382.8 355.0 0.00 16.70 133.085 8 268 4 330.5 0.05 3.40 101.330 11 З 29 179.3 336.0 0.05 15.50 151.665 6 ###### // Create 'rikz2' that only contains Beaches 1-4 from week 1 // > names(rikz) [1] "Sample" "Richness" "Week" "angle1" [5] "angle2" "exposure" "salinity" "temperature" [9] "NAP" "penetrability" "grainsize" "humus" "sorting1" "Beach" [13] "chalk" > rikz2 = rikz[rikz\$Week == 1,][rikz[rikz\$Week == 1,]\$Beach > 0 & rikz[rikz\$Week == 1,]\$Beach < 5,] > head(rikz2) Sample Richness Week angle1 angle2 exposure salinity temperature NAP З 17.5 -1.336 3 13 1 65 96 10 29.4 10 17 143 89 8 29.6 20.8 -1.334 10 1 2 2 10 62 96 10 29.4 17.5 -1.036 1 5 23 96 10 5 10 1 29.4 17.5 -0.684 96 10 29.4 1 11 32 17.5 0.045 1 1 9 9 19 1 26 89 8 29.6 20.8 0.061 penetrability grainsize humus chalk sorting1 Beach 237.1 3 194.5 0.10 3.45 59.220 1 257.9 197.0 0.00 2.60 59.575 2 10 2 226.9 200.0 0.30 2.50 59.000 1 5 251.9 202.0 0.05 2.45 57.760 1 253.9 222.5 0.05 2.05 1 69.830 1 248.9 205.5 0.15 2.30 9 58.810 2 ###### // The mean chalk value for Beach 5 // > rikz[rikz\$Beach == 5,][rikz[rikz\$Beach == 5,]\$chalk,] Sample Richness Week angle1 angle2 exposure salinity temperature NAP 22 22 22 4 22 21 10 29.9 19.8 -0.503 25 25 6 4 18 21 10 29.9 19.8 0.054 19.8 0.054 29.9 25.1 25 6 4 18 10 21 penetrability grainsize humus chalk sorting1 Beach 22 256.1 265.0 0.0 1.6 89.035 5 254.5 25 231.1 0.1 2.1 86.170 5 25.1 231.1 254.5 0.1 2.1 86.170 5 ##### // The mean grain size for Beach 8 // > mean(rikz[rikz\$Beach == 8,][,'grainsize']) [1] 327.9 ##### // Save the new data.frame 'rikz_2.csv' //

```
> write.csv(rikz, file = "rikz_2.csv")
```

11. Linear Models, predictions and probability distributions

11.1 Some terms

• **Mean** - is a set of values is the quantity commonly called the mean or the average. To get the mean sum the number of values and divide the result by the number of values.



Illustration 5: The mean

Im(y ~ 1) will return the mean value.

- **Regression** A method for fitting a curve or a straight line through a set of points using some goodness-of-fit criterion. The most common type of regression is linear regression.
- Linear Regression A regression that is linear in the unknown parameters used in the fit. It is defined by the *intercept point (a)* of the best fit line where the x axis is equal to zero plus the *slope (β)* of the best fit line times the value of x. The *intercept point (a)* is the mean.



Illustration 6: Linear regression model

- **Multiple Regression** A regression giving conditional expectation values of a given variable in terms of two or more other variables.
- **Null Hypothesis (H₀)** A H₀ is a statistical hypothesis that is tested for possible rejection under the assumption that it is true (usually that observations are the result of chance). It means that there is no expectation of change.
- Alternative Hypothesis (H_1) is the hypothesis used in hypothesis testing that is contrary to the H_0 . It is usually taken to be that the observations are the result of a real effect (with some amount of chance variation superposed). In the p-test values less than 0.05 indicate a significance and indicate contrary to the H_0 .
- **Estimate** Mean expectation of the mean, the Null Hypothesis (H_0) .
- Standard Error This gives one standard deviation (δ) of the certainty of the accuracy of the Estimate (mean). 2δ gives the certainty of accuracy for 95% of values. mean = 6.7195 and 2δ = 0.55, therefore there is a 95% chance of values giving a mean in the range of 6.17 7.27.
- t-value The coefficient divided by its standard error. The standard error is an estimate of the standard deviation of the coefficient, the amount it varies across cases. It can be thought of as a measure of the precision with which the regression coefficient is measured. This value says how many standard deviations from zero. For example:

the *Estimate / Std. Error = t-value ==>* 6.7195 / 0.2726 = 24.64967

- p-value Shows the expectation that the *Estimate* is accurate for values. A small p-value (typically ≤ 0.05) indicates strong evidence against the H₀, so you reject the H₀. A large p-value (> 0.05) indicates weak evidence against the H₀, so you shouldn't reject H₀. The p-value of 0.05 means that there is less than a 5% chance of the results arising due to random chance.
- t-test Refers to any statistical hypothesis test in which the test statistic follows a t-distribution under the H₀. A t-test is most commonly applied when the test statistic would follow a normal distribution if the value of a scaling term in the test statistic were known. Here is an example with the built-in *mtcars* dataset. It shows a t-test to compare vehicles in rows 1 to 10 with vehicles in rows 11 to 20 for column 1 which is miles per gallon (mpg). The H₀ is that there there is little to no statistical difference between the groups in terms of *mpg*. As the p-value from the test is 0.8744, it concurs with the H₀ and sets the expectation that there is indeed little statistical expectation of a significant difference.

```
> data(mtcars)
> head(mtcars[1])
                   mpg
Mazda RX4
                  21.0
Mazda RX4 Wag
                  21.0
Datsun 710
                  22 8
Hornet 4 Drive
                  21.4
Hornet Sportabout 18.7
Valiant
                  18.1
> a = t.test(mtcars[1:10,1],mtcars[11:20,1])
> print (a)
      Welch Two Sample t-test
data: mtcars[1:10, 1] and mtcars[11:20, 1]
t = 0.16185, df = 10.892, p-value = 0.8744
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
-6.055477 7.015477
sample estimates:
mean of x mean of y
    20.37
              19.89
> str(a)
list of 9
 $ statistic : Named num 0.162
 ... attr(*, "names")= chr "t"
$ parameter : Named num 10.9
  ... attr(*, "names")= chr "df"
$ p.value : num 0.874
$ conf.int : atomic [1:2] -6.06 7.02
  ... attr(*, "conf.level")= num 0.95
 $ estimate : Named num [1:2] 20.4 19.9
  ..- attr(*, "names")= chr [1:2] "mean of x" "mean of y"
 $ null.value : Named num 0
 ..- attr(*, "names")= chr "difference in means"
 $ alternative: chr "two.sided"
 $ method : chr "Welch Two Sample t-test"
 $ data.name : chr "mtcars[1:10, 1] and mtcars[11:20, 1]"
 - attr(*, "class")= chr "htest"
```

```
> unclass(a)
 $statistic
         t
 0.1618478
 $parameter
       df
 10.89198
 $p.value
 [1] 0.8743889
 $conf.int
 [1] -6.055477 7.015477
 attr(,"conf.level")
 [1] 0.95
 $estimate
 mean of x mean of y
     20.37
              19.89
 $null.value
 difference in means
                   Θ
 $alternative
 [1] "two.sided"
 $method
 [1] "Welch Two Sample t-test"
 $data.name
 [1] "mtcars[1:10, 1] and mtcars[11:20, 1]"
 # Extract the mean of y
 > a$estimate[2]
 mean of y
     19.89
 # Extract the 'p' value
 > a$p.value
 [1] 0.8743889
```

- Dependent Variable (DV) The variable being tested and measured in an experiment.
- Independent Variable (IV) The variable that is changed or controlled in a scientific experiment to test the effects on the DV.
- Analysis of Variance (ANOVA) is a collection of statistical models and their associated estimation procedures used to analyse the differences among group means in a sample.

$$y = a + \beta x_1 + \beta x_2$$

 Multivarite ANOVA (MANOVA) - is a procedure for comparing multivariate sample means. As a multivariate procedure, it is used when there are two or more dependent variables, and is typically followed by significance tests involving individual dependent variables separately. Analysis of covariance (ANCOVA) - is a general linear model which blends ANOVA and regression. ANCOVA evaluates whether the means of a DV are equal across levels of a categorical IV.

- Generalised Linear Model (GLM) is a flexible generalisation of ordinary linear regression that allows for response variables that have error distribution models other than a normal distribution.
- Generalised Linear Mixed Model (GLMM) is an extension to GLM in which the linear predictor contains random effects in addition to the usual fixed effects. They also inherit from GLMs the idea of extending linear mixed models to nonnormal data.

11.2 Demonstration

Working with the *owl_data.csv* file once more to demonstrate linear models, predictions and probability distributions.

```
> setwd('~/datasets/owl_data_2')
> owl = read.csv('owl_data.csv')
###### // Inspect the data //
> str(owl)
'data.frame': 599 obs. of 5 variables:
 $ nest : Factor w/ 27 levels "AutavauxTV", "Bochet",..: 1 1 1 1 1 1 1 1 1 1 ...
 $ sex : Factor w/ 2 levels "Female", "Male": 2 2 2 2 2 2 2 1 2 1 ...
          : num 22.2 22.4 22.5 22.6 22.6 ...
 $ food
 $ begging: int 4 0 2 2 2 2 18 4 18 0 ...
$ brood : int 5 5 5 5 5 5 5 5 5 5 ...
> summary(owl)
         nest
                                           food begging brood
                        sex

      Oleyes
      : 52
      Female:245
      Min. :21.71
      Min. : 0.00
      Min. :1.000

      Moutet
      : 41
      Male :354
      1st Qu.:23.11
      1st Qu.: 0.00
      1st Qu.:4.000

      Etrabloz
      : 34
      Median :24.38
      Median : 5.00
      Median :4.000

 Yvonnand : 34
                                      Mean :24.76 Mean : 6.72 Mean :4.392
 Champmartin: 30
                                     3rd Qu.:26.25 3rd Qu.:11.00 3rd Qu.:5.000
                                    Max. :29.25 Max. :32.00 Max. :7.000
 Lucens : 29
 (Other) :379
> head(owl)
        nest sex food begging brood
1 AutavauxTV Male 22.25 4 5
2 AutavauxTV Male 22.38
                                   0
                                          5
3 AutavauxTV Male 22.53
                                 2 5
4 AutavauxTV Male 22.56
5 AutavauxTV Male 22.61
6 AutavauxTV Male 22.65
                                  2 5
                                  2 5
6 AutavauxTV Male 22.65
                                  2
                                          5
```

The purpose of this is to explain the *begging rate* of the chicks in the nest. Response variable = y = begging **Im()**: is used to fit linear models. It can be used to carry out regression, single stratum analysis of variance and analysis of covariance.

11.3 The mean()

Question 1: What is the begging rate of the sampled population?





Now the statistics are given in the *summary()* of the data. *Estimate*, *Standard error*, *t*-*value* and *p*-*value*.

> mod = lm(begging ~ 1, data = owl)

```
> summary(mod)
Call:
lm(formula = begging ~ 1, data = owl)
Residuals:
    Min    1Q Median    3Q    Max
    -6.72    -6.72    -1.72    4.28    25.28
Coefficients:
        Estimate Std. Error t value Pr(>|t|)
(Intercept)    6.7195     0.2726    24.65    <2e-16 ***
---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 6.671 on 598 degrees of freedom</pre>
```

11.4 The t-test

Question 2: What is the begging rate of males versus females?

In this case *R* coded *females* = 0 and *males* = 1. So therefore females indicate the intercept (* 0) and males the difference from the intercept. This is indicated by *sexMale* in the second line of the output, males have been labelled internally as = 1, therefore *sexFemale* must be = 0. So the intercept of females 6.0122 plus the difference 1.1968 gives the intercept value for males = 7.209.

In *R* the linear modelling is carried out using the Im() function. The DV is placed to the left side of the tilde and the IV or IVs are placed to the right. As such the DV are the *y* axis and the IVs are on the *x* axis of associated graphs.

Im(DV ~ IV(s), data=<dataset>)

```
> lm(begging ~ sex, data = owl)
Call:
lm(formula = begging ~ sex, data = owl)
Coefficients:
(Intercept) sexMale
        6.012 1.197
> mod2 = lm(begging ~ sex, data = owl)
```

```
> summary(mod2)
Call:
lm(formula = begging ~ sex, data = owl)
Residuals:
         10 Median
  Min
                     30
                             Мах
-7.209 -6.012 -1.209 4.791 24.791
Coefficients:
          Estimate Std. Error t value Pr(>|t|)
(Intercept) 6.0122 0.4249 14.151 <2e-16 ***
                       0.5527 2.165 0.0307 *
sexMale
             1.1968
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Residual standard error: 6.65 on 597 degrees of freedom Multiple R-squared: 0.007793, Adjusted R-squared: 0.006131 F-statistic: 4.689 on 1 and 597 DF, p-value: 0.03075



Illustration 8: Mean begging rate based on sex

11.5 ANOVA, the Analysis of Variance

Comparing the means of greater than two groups.

Question 3: How does the begging rate change as nest size increases?

Important: First change *owl\$brood* to a factor.

This is an ANOVA. the *lm*(*begging* ~ *brood*, *data* = *owl*) doesn't give much information but the *summary*(*lm*(*begging* ~ *brood*, *data* = *owl*)) gives more useful information with *brood1* indicating the intercept and all other broods are indicated as the difference from *brood1*.

Can't be certain that the birds didn't beg at a negative rate. Obviously this isn't possible.

t-value and p-values demonstrate the certainty of difference between each brood and *brood1*. However there is a high level of uncertainty of H_0 .

For example *brood7* p-value is less than 0.05 and therefore indicates that it doesn't have the same begging rate as *brood1*, i.e. reject the H_0 .

This test is only comparing each brood relative to *brood1*. To review this further posthoc tests which will carry out statistics between each brood group.

```
> lm(begging ~ brood, data = owl)
Call:
lm(formula = begging ~ brood, data = owl)
Coefficients:
(Intercept)
                brood2
                            brood3
                                          brood4
                                                       brood5
                                                                    brood6
                -0.5263
                             0.4237
                                                                    4.5385
     4.0000
                                          3.0905
                                                       2.6404
     brood7
     7.5000
> mod3 = lm(begging ~ brood, data = owl)
```

```
> summary(mod3)
Call:
lm(formula = begging ~ brood, data = owl)
Residuals:
   Min
            10 Median
                            30
                                    Мах
-11.500 -5.090 -1.500
                         4.743 25.360
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept)
            4.0000
                        3.2477
                                 1.232
                                         0.2186
             -0.5263
                        3.4144 -0.154
brood2
                                         0.8775
             0.4237
brood3
                        3.3560
                                0.126
                                         0.8996
brood4
             3.0905
                        3.2785
                                 0.943
                                         0.3462
brood5
             2.6404
                        3.2761
                                 0.806
                                         0.4206
                        3.4886
brood6
             4.5385
                                 1.301
                                         0.1938
                        3.4334
                                         0.0293
brood7
             7.5000
                                 2.184
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 6.495 on 592 degrees of freedom
```

Multiple R-squared: 0.06141, Adjusted R-squared: 0.0519 F-statistic: 6.456 on 6 and 592 DF, p-value: 1.317e-06

 $lm(y \sim x1 + x2 + ...)$

```
y = a + \beta x_1 + \beta x_2 + \beta x_n
```



Illustration 9: ANOVA

anova() - Compute analysis of variance (or deviance) tables for one or more fitted model objects.

aov() - Fit an analysis of variance model by a call to Im for each stratum.

tukey() - Create a set of confidence intervals on the differences between the means of the levels of a factor with the specified family-wise probability of coverage. The intervals are based on the Studentised range statistic, *Tukey Honest Significant Difference* method.

```
> post = aov(mod3)
> TukeyHSD(post)
 Tukey multiple comparisons of means
    95% family-wise confidence level
Fit: aov(formula = mod3)
$brood
          diff
                       lwr
                                  upr
                                          p adj
2-1 -0.5263158 -10.6277301 9.575099 0.9999989
3-1 0.4237288 -9.5049990 10.352457 0.9999997
4-1 3.0904762 -6.6089619 12.789914 0.9654242
5-1 2.6403509 -7.0519281 12.332630 0.9843572
6-1 4.5384615 -5.7825744 14.859498 0.8514925
7-1 7.5000000 -2.6578473 17.657847 0.3055212
3-2 0.9500446 -3.0470773 4.947167 0.9923990
4-2 3.6167920 0.2291015 7.004482 0.0276208
5-2 3.1666667 -0.2004714 6.533805 0.0809328
6-2 5.0647773 0.1738541 9.955701 0.0368230
7-2 8.0263158 3.4898846 12.562747 0.0000048
4-3 2.6667474 -0.1647738 5.498269 0.0800918
5-3 2.2166221 -0.5902774 5.023521 0.2284480
6-3 4.1147327 -0.4087831 8.638249 0.1022524
7-3 7.0762712 2.9386060 11.213936 0.0000116
5-4 -0.4501253 -2.2880991 1.387848 0.9910760
6-4 1,4479853 -2,5472191 5,443190 0,9360239
7-4 4.4095238 0.8570970 7.961951 0.0048667
6-5 1.8981107 -2.0796816 5.875903 0.7955024
7-5 4.8596491 1.3268162 8.392482 0.0010456
7-6 2.9615385 -2.0448994 7.967976 0.5824679
```

11.6 Regression

Compare the means of a continuous variable.

Question 4: How does begging rate change as food increases?

This demonstrates that the expectation line intercepts the y axis at a 26.9 begging rate. This may or may not have an actual biological meaning as the samples taken may not actually have gone down to zero food. The same can be said of the line crossing the xaxis. However it is useful mathematically. The slope of negative (-0.81) means the expectation is that *begging* reduces by 0.81 for every 1 unit of *food* added. The very low *p*-value means that there is a very high confidence that more food decreases the begging rate.

```
> lm(begging ~ food, data = owl)
Call:
lm(formula = begging ~ food, data = owl)
Coefficients:
(Intercept) food
    26.9738 -0.8181
> mod4 = lm(begging ~ food, data = owl)
```

```
> summary(mod4)
Call:
lm(formula = begging ~ food, data = owl)
Residuals:
          10 Median
  Min
                       30
                              Мах
-9.155 -5.253 -1.298 4.454 25.544
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
                     3.4431 7.834 2.17e-14 ***
(Intercept) 26.9738
                        0.1387 -5.900 6.09e-09 ***
            -0.8181
food
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Residual standard error: 6.49 on 597 degrees of freedom Multiple R-squared: 0.0551, Adjusted R-squared: 0.05351 F-statistic: 34.81 on 1 and 597 DF, p-value: 6.092e-09



 $lm(y \sim x1 + x2 + ...)$

 $y = a + \beta x_1 + \beta x_2 + \beta x_n$

Illustration 10: Regression

11.7 Multiple regression

Compare the means of a continuous variable plus a group variable.

Question 5a: How does begging rate change as food and sex increase?

```
> lm(begging ~ food + sex, data = owl)
Call:
lm(formula = begging \sim food + sex, data = owl)
Coefficients:
(Intercept) food
26.4475 -0.8276
                        sexMale
1.2892
                            1.2892
> mod5a = lm(begging ~ food + sex, data = owl)
> summary(mod5a)
Call:
lm(formula = begging ~ food + sex, data = owl)
Residuals:
  Min 1Q Median 3Q
                             Мах
-9.711 -5.244 -1.597 4.700 25.020
Coefficients:
          Estimate Std. Error t value Pr(>|t|)
(Intercept) 26.4475 3.4365 7.696 5.85e-14 ***
food -0.8276 0.1382 -5.990 3.63e-09 ***
sexMale 1.2892 0.5374 2.399 0.0168 *
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 6.464 on 596 degrees of freedom
Multiple R-squared: 0.06413, Adjusted R-squared: 0.06099
F-statistic: 20.42 on 2 and 596 DF, p-value: 2.642e-09
```

 $lm(y \sim x1 + x2 + ...)$ $y = a + \beta x_1 + \beta x_2 + \beta x_n$



Illustration 11: Multiple Regression

Question 5b: How does begging rate change as food and brood increase?

The data is a multidimensional model with both food and brood. In this case brood is now continuous data showing brood size (as.numeric(owl\$brood)).

```
> owl$brood = as.numeric(owl$brood)
> lm(begging ~ food + brood, data = owl)
Call:
lm(formula = begging ~ food + brood, data = owl)
Coefficients:
(Intercept)
                    food
                                brood
    21.0447
                 -0.7877
                               1.1786
> mod5b = lm(begging ~ food + brood, data = owl)
> summary(mod5b)
Call:
lm(formula = begging ~ food + brood, data = owl)
Residuals:
   Min
             1Q Median
                             ЗQ
                                    Мах
-10.981 -5.250 -1.300
                         4.514 24.818
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
                         3.5561 5.918 5.50e-09 ***
(Intercept) 21.0447
food
             -0.7877
                         0.1358 -5.799 1.08e-08 ***
                                  5.220 2.47e-07 ***
brood
             1.1786
                         0.2258
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 6.352 on 596 degrees of freedom
Multiple R-squared: 0.09641, Adjusted R-squared: 0.09338
F-statistic: 31.79 on 2 and 596 DF, p-value: 7.583e-14
```



Illustration 12: Another multiple regression

11.8 ANCOVA, the Analysis of Covariance

Comparing the means of multiple variables and allow for interactions between them.

Question 5c: How does the begging rate change as food & sex increase, while allowing for the food effect of each sex to differ?

The interaction of food & sex: an ANCOVA. If the effect of both food and sex change relative to each other then the *interaction* of both must be accounted for. This allows the relationship between males and females to change. As the *p*-value is quite large there is no reason to reject the H_0 . In other words there is little evidence that there is a divergence between the begging rate of males and females given greater levels of food.

```
> lm(begging ~ food + sex + food*sex, data = owl)
Call:
lm(formula = begging ~ food + sex + food * sex, data = owl)
Coefficients:
  (Intercept) food sexMale food:sexMale
    23.6642 -0.7149 6.0699 -0.1933
> mod5c = lm(begging ~ food + sex + food*sex, data = owl)
> summart(mod5c)
Error in summart(mod5c) : could not find function "summart"
```

```
> summary(mod5c)
Call:
lm(formula = begging \sim food + sex + food * sex, data = owl)
Residuals:
   Min
           1Q Median
                         ЗQ
                               Мах
-9.955 -5.241 -1.461 4.515 25.042
Coefficients:
             Estimate Std. Error t value Pr(>|t|)
                                  4.462 9.71e-06 ***
(Intercept)
              23.6642
                          5.3035
                                  -3.338 0.000895 ***
food
              -0.7149
                          0.2141
sexMale
               6.0699
                          6.9569
                                   0.873 0.383286
                          0.2804 -0.689 0.490936
food:sexMale -0.1933
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Residual standard error: 6.467 on 595 degrees of freedom Multiple R-squared: 0.06488, Adjusted R-squared: 0.06016 F-statistic: 13.76 on 3 and 595 DF, p-value: 1.094e-08



 $lm(y \sim x1 + x2 + x1 * x2)$

Illustration 13: ANCOVA

Function	R function	Result
$y = \alpha + \beta x$	Im(begging ~ 1)	The mean of y
$y = \alpha + \beta x$	Im(begging ~ sex)	t-test
$y = \alpha + \beta x_1 + \beta x_2$	Im(begging ~ brood size)	ANOVA
$y = \alpha + \beta x$	Im(begging ~ food)	Simple regression
$y = \alpha + \beta x_1 + \beta x_2$	Im(begging ~ food + sex)	Multiple regression
$y = \alpha + \beta x_1 + \beta x_2$	Im(begging ~ food + brood)	Multiple regression
$y = \alpha + \beta x_1 + \beta x_2$	Im(begging ~ food + sex + food*sex)	ANCOVA

11.9 Linear model summary

All linear models are about calculating the mean of a particular group or groups. The *statistics* simply tell you how confident you are that the means differ between groups.

11.10 Exercise: Linear models

- 1. Import the *RIKZ.csv* data into an object called rikz (inspect the data etc).
- 2. What is the average species richness across all samples?
- 3. How does species richness vary with sand grainsize?
 - HINT: simple regression.
- 4. How does species richness vary at the different beaches?
 - HINT: ANOVA.
- 5. How does species richness vary with sampling time (week 1&2 versus week 3&4)?
 - HINT: t-test (will need to create a 2-category variable from week).
- 6. How does species richness vary with NAP and beach angle; is there any evidence of an interaction effect between them?
 - HINT: multiple regression.

Question 1: Import the 'RIKZ.csv' data into an object called rikz.

- > setwd('~/datasets/RIKZ_2')
- > rikz = read.csv('RIKZ.csv')

Explore the data frame.

<pre>> summary(rikz)</pre>				
Sample	Richness	Week	angle1	angle2
Min. : 1 Mi	.n. : 0.000	Min. :1.000	Min. : 6.00	Min. :21.00
1st Qu.:12 1s	st Qu.: 3.000	1st Qu.:2.000	1st Qu.: 22.00	1st Qu.:32.00
Median :23 Me	dian : 4.000	Median :2.000	Median : 32.00	Median :42.00
Mean :23 Me	an : 5.689	Mean :2.333	Mean : 50.31	Mean :57.78
3rd Qu.:34 3r	d Qu.: 8.000	3rd Qu.:3.000	3rd Qu.: 55.00	3rd Qu.:89.00
Max. :45 Ma	x. :22.000	Max. :4.000	Max. :312.00	Max. :96.00
exposure	salinity	temperature	NAP	
Min. : 8.00	Min. :26.4	Min. :15.80) Min. :-1.3360)
1st Qu.:10.00	1st Qu.:27.1	1st Qu.:17.50) 1st Qu.:-0.3750)
Median :10.00	Median :27.9	Median :18.77	′ Median : 0.1670)
Mean :10.22	Mean :28.1	Mean :18.77	′ Mean : 0.3477	,
3rd Qu.:11.00	3rd Qu.:29.4	3rd Qu.:20.00) 3rd Qu.: 1.1170)
Max. :11.00	Max. :29.9	Max. :20.80	Max. : 2.2550)
penetrability	grainsize	humus	chalk	
Min. :151.8	Min. :186.0	Min. :0.00	0000 Min. : 0.8	50
1st Qu.:237.1	1st Qu.:222.5	1st Qu.:0.00	0000 1st Qu.: 2.2	00
Median :256.1	Median :266.0	Median :0.05	000 Median : 4.7	50
Mean :289.4	Mean :272.5	Mean :0.05	028 Mean : 7.9	61
3rd Qu.:272.9	3rd Qu.:316.5	3rd Qu.:0.10	0000 3rd Qu.: 9.1	.50
Max. :624.0	Max. :405.5	Max. :0.30	0000 Max. :45.7	50
sorting1	Beach			
Min. : 53.08	Min. :1			
1st Qu.: 72.75	1st Qu.:3			
Median : 89.03	Median :5			
Mean : 97.82	Mean :5			
3rd Qu.:115.44	3rd Qu.:7			
Max. :248.60	Max. :9			
> str(r1kz)				
'data.trame': 45	005. OF 15 V	ariables:		
\$ Samp⊥e	: 1nt 1 2 3 4	5678910.		
\$ Richness	: int 11 10 1	3 11 10 8 9 8 1	.9 17	
\$ Week	: int 1111	111111.		
\$ angle1	: 1nt 32 62 6	5 55 23 129 126	52 26 143	
\$ anglez	: 1nt 96 96 9	6 96 96 89 89 8 0 10 10 0 0 0 0	9 89 89	
\$ exposure	: int 10 10 1	0 10 10 8 8 8 8	8	~ ~ ~ ~
\$ salinity	: num 29.4 29	.4 29.4 29.4 29	0.4 29.6 29.6 29.6	29.6 29.6
\$ temperature	: num 17.5 17	.5 17.5 17.5 17	.5 20.8 20.8 20.8	20.8 20.8
\$ NAP	: num 0.045 -	1.036 -1.336 0.	616 -0.684	
\$ penetrability	': num 254 227	237 249 252	•	
\$ grainsize	: num 222 200			0
\$ numus	: num 0.05 0.	3 0.1 0.15 0.05		0
\$ Chaik	: num 2.05 2.	5 3.45 1.6 2.45	2.5 1.85 1.7 2.3	2.6
\$ SOTTING1	: num 69.8 59	59.2 6/ 8 57.8	5	
\$ Beach	: int 1111	122222.		
> names(rikz)				
[1] "Samnlo"	"Richness	"Wook"	"1010"	
[5] "angle?"	"exposure	" "salinit	v" "temperatu	Ire"
[9] "NAP"	"nenetrah	Jarrinto ilitv" "arainei	יז נכוווטביומנט. דפי "humus"	
[13] "chalk"	"sorting1	" "Reach"	.20 1101103	
LTO] ONUTI	301 C±119±	Deach		

>	> head(rikz)								
	Sample	Richness	Week ar	ngle1 a	ngle2	exposure	salinity	temperature	NAP
1	1	11	1	32	96	10	29.4	17.5	0.045
2	2	10	1	62	96	10	29.4	17.5	-1.036
3	3	13	1	65	96	10	29.4	17.5	-1.336
4	4	11	1	55	96	10	29.4	17.5	0.616
5	5	10	1	23	96	10	29.4	17.5	-0.684
6	6	8	1	129	89	8	29.6	20.8	1.190
	penetra	ability g	rainsize	e humus	chalk	sorting1	Beach		
1		253.9	222.5	5 0.05	2.05	69.830	1		
2		226.9	200.0	0.30	2.50	59.000	1		
3		237.1	194.5	5 0.10	3.45	59.220	1		
4		248.6	221.0	0.15	5 1.60	67.750	1		
5		251.9	202.0	0.05	5 2.45	57.760	1		
6		250.1	192.5	5 0.10	2.50	53.075	2		

Question 2: What is the average species richness across all samples?

Mean : 5.689 # Extracted from the summary(rikz)
Alternatively extract from the intercept (α) where y = 0
> lm (Richness ~ 1, rikz)
Call:
lm(formula = Richness ~ 1, data = rikz)
Coefficients:
(Intercept)
5.689

Question 3: How does species richness vary with sand grainsize?

```
> rikz 1 = lm(Richness ~ grainsize, data=rikz)
> summary(rikz1)
Call:
lm(formula = Richness ~ grainsize, data = rikz)
Residuals:
   Min
           1Q Median
                         ЗQ
                                  Мах
-8.4833 -3.6733 -0.2693 2.1872 16.0701
Coefficients:
          Estimate Std. Error t value Pr(>|t|)
(Intercept) 14.49529 3.40326 4.259 0.000109 ***
grainsize -0.03232
                     0.01222 -2.644 0.011386 *
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 4.694 on 43 degrees of freedom
Multiple R-squared: 0.1399, Adjusted R-squared: 0.1198
F-statistic: 6.991 on 1 and 43 DF, p-value: 0.01139
```

Answer, Very little as the slope of the line is almost zero with a p-value < 0.05 means that the model demonstrates a high probability of H_0 .



Illustration 14: Richness/grainsize

Question 4: How does species richness vary at the different beaches?

```
> rikz$Beach = as.factor(rikz$Beach)
> rikz2 = lm(Richness ~ Beach, data=rikz)
> summary(rikz2)
Call:
lm(formula = Richness ~ Beach, data = rikz)
Residuals:
            10 Median
   Min
                             30
                                     Мах
  -7.4
          -1.4 -0.2
                            1.0
                                    14.6
Coefficients:
             Estimate Std. Error t value Pr(>|t|)
(Intercept) 11.000 1.761 6.245 3.27e-07 ***

        Beach2
        1.200
        2.491
        0.482
        0.63289

        Beach3
        -7.600
        2.491
        -3.051
        0.00426

        Beach4
        -8.600
        2.491
        -3.453
        0.00144

                              2.491 -3.051 0.00426 **
2.491 -3.453 0.00144 **
Beach4
                -8.600
                -3.600
                             2.491 -1.445 0.15703
Beach5
                             2.491 -2.810 0.00796 **
Beach6
                -7.000
Beach7
              -8.800
                             2.491 -3.533 0.00115 **
              -7.000
-6.400
                             2.491 -2.810 0.00796 **
Beach8
               -6.400
                               2.491 -2.569 0.01448 *
Beach9
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 3.938 on 36 degrees of freedom
Multiple R-squared: 0.4931, Adjusted R-squared: 0.3805
F-statistic: 4.378 on 8 and 36 DF, p-value: 0.0009179
```
As the p-value is less than 0.05 there is a high level of confidence that at least one of the pairs of beaches vary from each other in species richness. Carrying out fit an analysis of variance model test (aov()) test to investigate further.

The resulting p-value of 0.000918 says that with a high level of probability that there is a variance between beaches. Compute the *Tukey Honest Significant Differences* between the beaches.

```
> rikz4 = TukeyHSD(rikz3)
> print(rikz4)
Tukey multiple comparisons of means
95% family-wise confidence level
```

```
Fit: aov(formula = rikz2)
```

\$Beach

	diff	lwr	upr	p adj
2-1	1.200000e+00	-7.01263	9.41262978	0.9999026
3-1	-7.600000e+00	-15.81263	0.61262978	0.0882779
4-1	-8.600000e+00	-16.81263	-0.38737022	0.0342201
5-1	-3.600000e+00	-11.81263	4.61262978	0.8723088
6-1	-7.000000e+00	-15.21263	1.21262978	0.1475693
7-1	-8.800000e+00	-17.01263	-0.58737022	0.0279853
8-1	-7.000000e+00	-15.21263	1.21262978	0.1475693
9-1	-6.400000e+00	-14.61263	1.81262978	0.2347476
3-2	-8.800000e+00	-17.01263	-0.58737022	0.0279853
4-2	-9.800000e+00	-18.01263	-1.58737022	0.0097691
5-2	-4.800000e+00	-13.01263	3.41262978	0.6005725
6-2	-8.200000e+00	-16.41263	0.01262978	0.0506097
7-2	-1.000000e+01	-18.21263	-1.78737022	0.0078509
8-2	-8.200000e+00	-16.41263	0.01262978	0.0506097
9-2	-7.600000e+00	-15.81263	0.61262978	0.0882779
4-3	-1.000000e+00	-9.21263	7.21262978	0.9999757
5-3	4.000000e+00	-4.21263	12.21262978	0.7953899
6-3	6.000000e-01	-7.61263	8.81262978	0.9999995
7-3	-1.200000e+00	-9.41263	7.01262978	0.9999026
8-3	6.000000e-01	-7.61263	8.81262978	0.9999995
9-3	1.200000e+00	-7.01263	9.41262978	0.9999026
5-4	5.000000e+00	-3.21263	13.21262978	0.5484761
6-4	1.600000e+00	-6.61263	9.81262978	0.9991816
7-4	-2.000000e-01	-8.41263	8.01262978	1.0000000
8-4	1.600000e+00	-6.61263	9.81262978	0.9991816
9-4	2.200000e+00	-6.01263	10.41262978	0.9925552
6-5	-3.400000e+00	-11.61263	4.81262978	0.9034383
7-5	-5.200000e+00	-13.41263	3.01262978	0.4968860
8-5	-3.400000e+00	-11.61263	4.81262978	0.9034383
9-5	-2.800000e+00	-11.01263	5.41262978	0.9664876
7-6	-1.800000e+00	-10.01263	6.41262978	0.9981055
8-6	8.881784e-16	-8.21263	8.21262978	1.0000000
9-6	6.000000e-01	-7.61263	8.81262978	0.9999995
8-7	1.800000e+00	-6.41263	10.01262978	0.9981055
9-7	2.400000e+00	-5.81263	10.61262978	0.9869248
9-8	6.000000e-01	-7.61263	8.81262978	0.9999995

Data Analytics

Looking over the p-values there are a number of beach pairs that how a value lower than 0.05, in other words there is a difference in species diversity for these beach pairs. Convert the list to a dataframe and pull out that data.

```
> df.rikz4 = data.frame(unclass(rikz4))
> names(df.rikz4)
                 "Beach.lwr"
[1] "Beach.diff"
                                "Beach.upr"
                                              "Beach.p.adj"
> df.rikz4[df.rikz4$Beach.p.adj < 0.05,]</pre>
   Beach.diff Beach.lwr Beach.upr Beach.p.adj
         -8.6 -16.81263 -0.3873702 0.034220065
4-1
7-1
          -8.8 -17.01263 -0.5873702 0.027985274
         -8.8 -17.01263 -0.5873702 0.027985274
3-2
4-2
         -9.8 -18.01263 -1.5873702 0.009769101
7-2
        -10.0 -18.21263 -1.7873702 0.007850934
```

There is a significant difference in species diversity between beaches:

- beach 4 and beach 1
- beach 7 and beach 1
- beach 3 and beach 2
- beach 4 and beach 2
- beach 7 and beach 2

Question 5: How does species richness vary with sampling time?

Week 1&2 versus week 3&4 t-test (need to create a 2-category variable from 'week").

```
> rikz5 = rikz[rikz$Week < 5,]</pre>
> rikz5$fortnight = ifelse(rikz5$Week < 3, 1, 2)</pre>
> rikz5$fortnight = as.factor(rikz5$fortnight)
> rikz6 = lm(Richness ~ fortnight, data=rikz5)
> summary(rikz6)
Call
lm(formula = Richness ~ fortnight, data = rikz5)
Residuals:
                      3Q
  Min 1Q Median
                                Мах
 -5.24 -3.24 -1.00 2.00 17.00
Coefficients:
         Estimate Std. Error t value Pr(>|t|)
(Intercept) 6.240 1.004 6.212 1.79e-07 ***
fortnight2 -1.240 1.507 -0.823 0.415
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 5.022 on 43 degrees of freedom
Multiple R-squared: 0.01551, Adjusted R-squared: -0.007387
F-statistic: 0.6774 on 1 and 43 DF, p-value: 0.415
```

The p-value of 1.79e⁻⁰⁷ being the intercept does not mean anything significant. The fact that 0.415 is greater that 0.05 indicates that there is little likelihood that there is any difference between species richness *fortnight1* and *fortnight2*.

Question 6: How does species richness vary with NAP and beach angle?

```
> rikz7 = lm(Richness ~ NAP + angle1, data=rikz)
> summary(rikz7)
Call:
lm(formula = Richness ~ NAP + angle1, data = rikz)
Residuals:
           1Q Median 3Q
   Min
                                  Мах
-4.9890 -3.0331 -0.7893 1.5244 14.0487
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 6.376753 0.903875 7.055 1.21e-08 ***
NAP -2.862905 0.636315 -4.499 5.31e-05 ***
          0.006113 0.012145 0.503 0.617
angle1
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 4.197 on 42 degrees of freedom
Multiple R-squared: 0.3286, Adjusted R-squared: 0.2966
F-statistic: 10.28 on 2 and 42 DF, p-value: 0.0002327
> rikz8 = lm(Richness ~ NAP + angle2, data=rikz)
> summary(rikz8)
Call:
lm(formula = Richness ~ NAP + angle2, data = rikz)
Residuals:
   Min 1Q Median 3Q
                                  Max
-5.4611 -2.4319 -0.8159 1.4524 15.7456
Coefficients:
    Estimate Std. Error t value Pr(>|t|)
(Intercept) 3.88471 1.34484 2.889 0.00609 **
NAP -2.72332 0.60297 -4.517 5.03e-05 ***
angle2 0.04761 0.02024 2.353 0.02339 *
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 3.956 on 42 degrees of freedom
Multiple R-squared: 0.4032, Adjusted R-squared: 0.3748
F-statistic: 14.19 on 2 and 42 DF, p-value: 1.961e-05
```

The p-value for *angle1* is not significant however the *p-value* of 0.02339 for *angle2* demonstrates significance in the difference in species richness.

Question 7: Is there any evidence of an interaction effect between them?

```
> rikz9 = lm(Richness ~ NAP + angle1 + NAP * angle1, data=rikz)
> summary(rikz9)
Call:
lm(formula = Richness ~ NAP + angle1 + NAP * angle1, data = rikz)
Residuals:
    Min
             1Q Median
                             3Q
                                     Мах
-5.3839 -2.7098 -0.8666 1.6629 14.6538
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 5.68941 1.13138 5.029 1.02e-05 ***
NAP-2.130580.96476-2.2080.0329 *angle10.019470.017951.0840.2846NAP:angle1-0.014180.01404-1.0100.3186
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 4.196 on 41 degrees of freedom
Multiple R-squared: 0.3449, Adjusted R-squared: 0.2969
F-statistic: 7.195 on 3 and 41 DF, p-value: 0.0005466
> rikz10 = lm(Richness ~ NAP + angle2 + NAP * angle2, data=rikz)
> summary(rikz10)
Call:
lm(formula = Richness ~ NAP + angle2 + NAP * angle2, data = rikz)
Residuals:
           1Q Median 3Q
   Min
                                Мах
-5.608 -2.265 -1.046 1.453 16.257
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 3.56005 1.41559 2.515 0.0159 *
NAP-1.811181.33015-1.3620.1807angle20.052510.021312.4650.0180 *
NAP:angle2 -0.01603 0.02081 -0.770 0.4455
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 3.976 on 41 degrees of freedom
Multiple R-squared: 0.4117, Adjusted R-squared: 0.3687
F-statistic: 9.565 on 3 and 41 DF, p-value: 6.513e-05
```

There does appear to be a significant impact on species richness when the interaction effect between angle2 and NAP is considered. However there does not appear to be a significant impact when the interaction effect between angle1 and NAP is considered.

11.11 Model Predictions

The *predict()* function returns predictions from the results of various model fitting functions. Most prediction methods which are similar to those for linear models have an argument *newdata* specifying the first place to look for explanatory variables to be used for prediction.

Input to the *predict()* function must be in the format of a dataframe.

```
> owl = read.csv('owl_data.csv')
> owl.lm = lm(begging~sex+food+sex*food, data = owl)
> newdata = data.frame(sex=rep('Male',20),food=1:20)
> newdata.2 = data.frame(sex=rep('Female', 20), food=1:20)
> owl.lm_male = predict(owl.lm, newdata)
> owl.lm_female = predict(owl.lm, newdata.2)
> owl.lm_male
      1
              2
                      3
                               4
                                       5
                                                6
                                                        7
                                                                8
28.82591 27.91776 27.00962 26.10147 25.19332 24.28518 23.37703 22.46889
     9
           10 11 12 13 14 15
                                                               16
21.56074 20.65260 19.74445 18.83630 17.92816 17.02001 16.11187 15.20372
     17 18 19 20
14.29558 13.38743 12.47929 11.57114
> owl_lm_female
             2
                      3
                              4
                                       5
                                               6
                                                        7
                                                                8
      1
22.94928 22.23438 21.51949 20.80459 20.08970 19.37480 18.65991 17.94501
     g
            10
                     11 12 13 14 15
                                                               16
17.23012 16.51522 15.80033 15.08543 14.37054 13.65564 12.94075 12.22585
     17
             18
                     19
                              20
11.51096 10.79606 10.08117 9.36627
> colours = c("red","blue", "green",
             "yellow", "purple", "Cyan",
             "pink", "brown"
            )
> title = "Histogram of Owl predictions (Male)"
> title.2 = "Histogram of Owl predictions (Female)"
> hist(owl.lm_male, col = colours, main = title, ylab = "Begging", xlab = "Food")
```

Histogram of Owl predictions (Male)



Illustration 15: owl: Predictions (male)

hist(owl.lm_female, col = colours, main = title.2, ylab = "Begging", xlab = "Food")





Illustration 16: owl: Predictions (female)

12. Distribution models

Thus far the linear modelling assumed that the residuals fell around the line of regression on a Normal or Gaussian distribution. This form of model is a continuous function which approximates the exact binomial distribution of events. The Gaussian distribution shown is normalised so that the sum over all values of x gives a probability of one.

12.1 Standard Deviation

A simple example.

```
> a = c(9, 2, 5, 4, 12, 7, 8, 11, 9, 3, 7, 4, 12, 5, 4, 10, 9, 6, 9, 4)
> a
[1] 9 2 5 4 12 7 8 11 9 3 7 4 12 5 4 10 9 6 9 4
> b = mean(a)
> b
[1] 7
> c = a - b
> c
[1] 2 -5 -2 -3 5 0 1 4 2 -4 0 -3 5 -2 -3 3 2 -1 2 -3
> d = c*c
b <
[1] 4 25 4 9 25 0 1 16 4 16 0 9 25 4 9 9 4 1 4 9
> e = mean(d)
> e
[1] 8.9
> f = sqrt(e)
> f
[1] 2.983287
```

So for the set of data (9, 2, 5, 4, 12, 7, 8, 11, 9, 3, 7, 4, 12, 5, 4, 10, 9, 6, 9, 4) mean (μ) = 7 standard deviation δ = 2.983287

12.1.1 Plotting the Standard Deviation

Note for the plot # - type: the type of plot to be drawn where "n" means do not plot the points

- xlab: the title of the x axis
- ylab: the title of the y axis
- main: the overall title for the plot

 axes: when FALSE it suppresses the axis automatically generated by the high level plotting function so that we can create custom axis

```
# Set the sample mean to 7 and SD to 2.8
> sample_mean = 7
> sample_sd = 2.8
# Fill one SD
> sd_to_fill = 1
> lower_bound = sample_mean - sample_sd * sd_to_fill
> upper_bound = sample_mean + sample_sd * sd_to_fill
# Generates equally spaced values within 4 SD of mean
> x = seq(-4, 4, length = 1000) * sample_sd + sample_mean
# The height of the probability distribution at each point
> y = dnorm(x, sample_mean, sample_sd)
# Generate the plot
> plot(x, y, type="n", xlab = "Samples", ylab = "", main = "Distribution of
  Samples", axes = FALSE)
# Connect the points with each other to form a curve
lines(x, y)
# Returns a vector of boolean values to ensure only x values
# between bounds are allowed
> bounds_filter = x >= lower_bound & x <= upper_bound</pre>
> x_within_bounds = x[bounds_filter]
> y_within_bounds = y[bounds_filter]
# Bordering the area to be filled
> x_polygon = c(lower_bound, x_within_bounds, upper_bound)
> y_polygon = c(0, y_within_bounds, 0)
> polygon(x_polygon, y_polygon, col = "green")
# Returns the probability that a normally distributed random number
# will be less than the given number
> probability_within_bounds = pnorm(upper_bound, sample_mean, sample_sd) -
   pnorm(lower_bound, sample_mean, sample_sd)
# Concatenate the various values to display on the curve
 > text = paste("p(", lower_bound, "< height <", upper_bound, ") =",</pre>
    signif(probability_within_bounds, digits = 3))
# Display the text on the plot
> mtext(text)>
# Add an axis to the plot
> sd_axis_bounds = 5>
> axis_bounds = seq(-sd_axis_bounds * sample_sd + sample_mean, sd_axis_bounds
 * sample sd + sample mean, by = sample sd)
> axis(side = 1, at = axis_bounds, pos = 0)
```

By changing the value of the *sd_fill* from one to two and three the various areas under the curve for each standard deviation can be seen. There is a simple rule canned the *empirical rule* to remember these. 68.27%, 95.45% and 99.73% of the values lie within one, two and three standard deviations of the mean.



Illustration 17: Standard Deviation

Another example

Looking at another example in more detail.

To calculate the standard deviation of a set of random numbers:

- Work out the mean (µ).
- For each number, subtract the mean (x μ).
- Square the result $(x \mu)^2$.
- Get the mean of the squared differences $1/N \Sigma (x \mu)^2$. This is called the variance (v).
- Get the square root of that $\delta = \sqrt{1/N} \Sigma (x \mu)^2$.

```
> data.set = round(runif(100,0,1), 2)
```

```
> data.set
[1] 0.56 0.13 0.27 0.91 0.08 0.49 0.99 0.67 0.74 0.59 0.92 0.55 0.18 0.44 0.64
[16] 0.92 0.15 0.96 0.17 0.67 0.74 0.33 0.52 0.59 0.17 0.59 0.95 0.68 0.71 0.51
[31] 0.73 0.72 0.71 0.57 0.37 0.73 0.57 0.99 0.84 0.06 0.93 0.16 0.23 0.91 0.75
[46] 0.89 0.67 0.17 0.29 0.62 0.56 0.35 0.03 0.55 0.62 0.84 0.30 0.69 0.30 0.88
[61] 0.10 0.03 0.98 0.68 0.38 0.35 0.60 0.82 0.51 0.24 0.58 1.00 0.97 0.65 0.04
[76] 0.93 0.42 0.97 0.03 0.52 0.38 0.83 0.18 0.37 0.10 0.60 0.96 0.96 0.54 0.65
[91] 0.75 0.56 0.27 0.94 0.25 0.66 0.41 0.34 0.21 0.99
```

```
> mean_ds = mean(data.set)
```

```
> mean_ds
[1] 0.556
```

The mean: $\mu = 0.556$

Subtract the Mean from each x to get $(x - \mu)$.

```
> sub_ds = data.set - mean_ds
```

> sub_ds
[1] 0.004 -0.426 -0.286 0.354 -0.476 -0.066 0.434 0.114 0.184 0.034
[11] 0.364 -0.006 -0.376 -0.116 0.084 0.364 -0.406 0.404 -0.386 0.114
[21] 0.184 -0.226 -0.036 0.034 -0.386 0.034 0.394 0.124 0.154 -0.046
[31] 0.174 0.164 0.154 0.014 -0.186 0.174 0.014 0.434 0.284 -0.496
[41] 0.374 -0.396 -0.326 0.354 0.194 0.334 0.114 -0.386 -0.266 0.064
[51] 0.004 -0.206 -0.526 -0.006 0.064 0.284 -0.256 0.134 -0.256 0.324
[61] -0.456 -0.526 0.424 0.124 -0.176 -0.206 0.044 0.264 -0.046 -0.316
[71] 0.024 0.444 0.414 0.094 -0.516 0.374 -0.136 0.414 -0.526 -0.036
[81] -0.176 0.274 -0.376 -0.186 -0.456 0.044 0.404 0.404 -0.016 0.094
[91] 0.194 0.004 -0.286 0.384 -0.306 0.104 -0.146 -0.216 -0.346 0.434

Square each result to get $(x - \mu)^2$.

```
> sq_ds = sub_ds * sub_ds
```

```
> sq_ds
[1] 0.000016 0.181476 0.081796 0.125316 0.226576 0.004356 0.188356 0.012996
[9] 0.033856 0.001156 0.132496 0.000036 0.141376 0.013456 0.007056 0.132496
[17] 0.164836 0.163216 0.148996 0.012996 0.033856 0.051076 0.001296 0.001156
[25] 0.148996 0.001156 0.155236 0.015376 0.023716 0.002116 0.030276 0.026896
[33] 0.023716 0.000196 0.034596 0.030276 0.000196 0.188356 0.080656 0.246016
[41] 0.139876 0.156816 0.106276 0.125316 0.037636 0.111556 0.012996 0.148996
[49] 0.070756 0.004096 0.00016 0.042436 0.276676 0.000036 0.004096 0.088656
[57] 0.065536 0.017956 0.065536 0.104976 0.207936 0.276676 0.179776 0.015376
[65] 0.030976 0.042436 0.001936 0.069696 0.002116 0.099856 0.000576 0.197136
[73] 0.171396 0.008836 0.266256 0.139876 0.018496 0.171396 0.276676 0.001296
[81] 0.030976 0.075076 0.141376 0.034596 0.207936 0.001936 0.163216 0.163216
[89] 0.000256 0.008836 0.037636 0.000016 0.081796 0.147456 0.093636 0.010816
[97] 0.021316 0.046656 0.119716 0.188356
```

Get the mean of these values $1/N \Sigma (x - \mu)^2$.

```
> mean_ds_sd = mean(sq_ds)
```

> mean_ds_sd
[1] 0.081938

Therefore $\delta = 0.081938$.

12.2 Expand Grid - expand.grid()

Create a data-frame from all combinations of the supplied vectors or factors.

>	grid.dat	ta	
	height	weight	sex
1	60	100	Male
2	65	100	Male
3	70	100	Male
4	75	100	Male
5	80	100	Male
6	60	150	Male
7	65	150	Male
8	70	150	Male
9	75	150	Male
10	80	150	Male
11	60	200	Male
12	65	200	Male
13	70	200	Male
14	75	200	Male

15	80	200	Male
16	60	250	Male
17	65	250	Male
18	70	250	Male
19	75	250	Male
20	80	250	Male
21	60	300	Male
22	65	300	Male
23	70	300	Male
24	75	300	Male
25	80	300	Male
26	60	100	Female
27	65	100	Female
28	70	100	Female
29	75	100	Female
30	80	100	Female
31	60	150	Female
32	65	150	Female
33	70	150	Female
34	75	150	Female
35	80	150	Female
36	60	200	Female
37	65	200	Female
38	70	200	Female
39	75	200	Female
40	80	200	Female
41	60	250	Female
42	65	250	Female
43	70	250	Female
44	75	250	Female
45	80	250	Female
46	60	300	Female
47	65	300	Female
48	70	300	Female
49	75	300	Female
50	80	300	Female

12.3 Normal or Gaussian distribution

What has been shown is actually a Normal or Gaussian distribution. This is a very common continuous probability distribution that are often used to represent real-valued random variables whose distributions are not known.

12.4 Poisson distribution

The Poisson distribution is the probability distribution of independent event occurrences in an interval. If *lambda* (λ) is the mean occurrence per interval, then the probability of having x occurrences within a given interval is:

 $f(x) = \lambda^x e^{-\lambda} / x!$ where x = 0, 1, 2, 3, n



Note that the Poisson distribution is for zero and positive values only. This can be useful if the dataset being operated on cannot have negative values.

For example.

Question: If there are five birds spotted landing on a pond per minute on average, what is the probability of eight birds landing on the pond in any minute?

The probability of having eight or *less* birds landing on the pond in any minute is given by the *ppois()* function. This function is the density, distribution function, quantile function and random generation for the Poisson distribution with parameter *lambda* (λ). This is called the *lower tail* of the function.

```
> ppois(8, lambda=5)
[1] 0.9319064
```

Secondly it is necessary to calculate the probability of having eight or *more* birds landing on the pond in any minute is called the *upper tail*. Hence the probability of having seventeen or more cars crossing the bridge in a minute is in the upper tail of the probability density function.

```
> ppois(8, lambda=5, lower=FALSE)
[1] 0.06809363
```

So there is a 93% chance of having eight or less birds landing on the pond while there is a 6.8% chance of having eight or more birds landing on the pond in and minute.

12.4.1 Linear Models based on Poisson Distribution

Returning to an earlier example.

```
> setwd('~/datasets/RIKZ_2')
> rikz = read.csv('RIKZ.csv')
> rikz10 = lm(Richness ~ NAP + angle2 + NAP * angle2, data=rikz)
```

```
> summary(rikz10)
Call
lm(formula = Richness ~ NAP + angle2 + NAP * angle2, data = rikz)
Residuals:
  Min 10 Median 30 Max
-5.608 -2.265 -1.046 1.453 16.257
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 3.56005 1.41559 2.515 0.0159 *
NAP -1.81118 1.33015 -1.362 0.1807
           0.05251 0.02131 2.465 0.0180 *
angle2
NAP:angle2 -0.01603 0.02081 -0.770 0.4455
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 3.976 on 41 degrees of freedom
Multiple R-squared: 0.4117, Adjusted R-squared: 0.3687
F-statistic: 9.565 on 3 and 41 DF, p-value: 6.513e-05
```

Using the Normal Distribution did not show a p-value of much significance.

Try running a linear model using the Poisson Distribution. In *R* to do this it is necessary to run the *Fitting Generalized Linear Models* which offers the *family* option. to run models based on other distributions like poisson.

```
> rikz11 = glm(Richness ~ NAP + angle2 + NAP * angle2, data=rikz, family=poisson)
> summary(rikz11)
Call
glm(formula = Richness ~ NAP + angle2 + NAP * angle2, family = poisson,
     data = rikz)
Deviance Residuals:
                                  ЗQ
     Min
             1Q Median
                                                   Мах
-2.4543 -1.1122 -0.7233 0.7948 5.0552
Coefficients:
               Estimate Std. Error z value Pr(>|z|)
(Intercept) 1.259472 0.157057 8.019 1.06e-15 ***

        NAP
        -0.743224
        0.188056
        -3.952
        7.75e-05
        ***

        angle2
        0.008586
        0.002176
        3.946
        7.95e-05
        ***

        NAP:angle2
        0.003209
        0.002523
        1.272
        0.203

                                            1.272
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
(Dispersion parameter for poisson family taken to be 1)
     Null deviance: 179.753 on 44 degrees of freedom
Residual deviance: 97.026 on 41 degrees of freedom
AIC: 247.02
Number of Fisher Scoring iterations: 5
```

As can be seen the p-values show much more significance for this distribution model.

12.4.2 Linear Models based on other Distributions

There are many other distribution models. Here is a summary and *R* offers the capability to run tests with each by adjusting the *family* value.

Continuous Data	Data range	Link Function
Normal	Any value, positive or negative	1
Gamma	Positive values only	log
Beta	Values between zero and one	logit
Discrete Data	Data range	Link Function
Poisson	Zero and positive values	log
Binomial	Zero and positive values	logit

> rikz12 = glm(Richness ~ NAP, data=rikz, family=binomial)
Error in eval(family\$initialize) : y values must be 0 <= y <= 1</pre>

In this case //Richness/ would have to be yes/no values.

> rikz13 = glm(Richness ~ NAP, data=rikz, family=gamma)
Error in family() : 0 arguments passed to 'gamma' which requires 1

In this case Richness would have to consist of count values.

12.4.3 Link Functions

The link function provides the relationship between the linear predictor and the mean of the distribution function. There are many commonly used link functions, and their choice is informed by several considerations. The table above gives the link functions associated with the various distributions.

12.4.4 Type Response

The type of prediction is required for *predict.glm*. The default is on the scale of the linear predictors; the alternative *response* is on the scale of the response variable. Thus for a default binomial model the default predictions are of log-odds (probabilities on *logit* scale) and *type = "response"* gives the predicted probabilities. The *terms* option returns a matrix giving the fitted values of each term in the model formula on the linear predictor scale.

The alternative type to response is the *type* = "*terms*" option which returns a matrix giving the fitted values of each term in the model formula on the linear predictor scale.

```
> owl = read.csv('owl_data.csv')
```

```
> owl.glm = glm(begging~sex+food+sex*food, family=poisson, data = owl)
```

```
> newdata = data.frame(sex=rep('Male',20),food=1:20)
```

```
> owl.glm_male = predict.glm(owl.glm, newdata, family=poisson)
```

```
> owl.glm_male
               2
                       3
                                4
                                        5
                                                 6
      1
                                                         7
                                                                  8
5.155740 5.020808 4.885877 4.750945 4.616013 4.481082 4.346150 4.211218
      q
            10 11
                             12
                                       13 14
                                                      15
                                                                 16
4.076286 3.941355 3.806423 3.671491 3.536559 3.401628 3.266696 3.131764
     17
            18
                     19
                              20
2.996832 2.861901 2.726969 2.592037
> owl.glm_male = predict.glm(owl.glm, newdata, family=poisson, type='response')
> owl.glm_male
       1
                2
                          3
                                   4
                                            5
                                                      6
                                                                7
                                                                         8
173.42413 151.53377 132.40650 115.69357 101.09021 88.33015 77.18073 67.43864
              10
                       11 12 13 14
                                                         15
                                                                       16
       q
 58,92623 51,48830 44,98922 39,31048 34,34854 30,01291 26,22455 22,91437
      17
             18 19
                                  20
20.02202 17.49475 15.28648 13.35696
> owl.glm_male = predict.glm(owl.glm, newdata, family=poisson, type='terms')
> owl.glm_male
                food
                         sex:food
        sex
1
  0.1482603 3.0438445 0.093027486
  0.1482603 2.9157238 0.086216467
2
  0.1482603 2.7876031 0.079405448
3
4
  0.1482603 2.6594824 0.072594430
  0.1482603 2.5313617 0.065783411
5
  0.1482603 2.4032409 0.058972392
6
  0.1482603 2.2751202 0.052161374
7
8
  0.1482603 2.1469995 0.045350355
9 0.1482603 2.0188788 0.038539336
10 0.1482603 1.8907581 0.031728318
11 0.1482603 1.7626373 0.024917299
12 0.1482603 1.6345166 0.018106280
13 0.1482603 1.5063959 0.011295261
14 0.1482603 1.3782752 0.004484243
15 0.1482603 1.2501545 -0.002326776
16 0.1482603 1.1220338 -0.009137795
17 0.1482603 0.9939130 -0.015948813
18 0.1482603 0.8657923 -0.022759832
19 0.1482603 0.7376716 -0.029570851
20 0.1482603 0.6095509 -0.036381869
```

attr(,"constant") [1] 1.870608

12.5 Exercise: Linear Modelling 1

Using the *insect_spray.csv* data which shows counts of insects after a particular type of insect spray has been used.

- 1. Run a simple linear model comparing the effectiveness of the different sprays (this will be a traditional ANOVA).
- 2. Get predictions and their standard errors from this model for each spray type (using the *predict.Im* function).
- 3. Run the model again as a GLM using the poisson distribution (are the estimates different from model 1? why are they so different?).
- 4. Get predictions and their standard errors (using the *predict.glm* function). How do the SEs compare to model 1. Why are they so different?
- 5. Create a matrix to store the predictions and SEs from both models (i.e. four column matrix).
- 6. Save this matrix table of results as a .csv file in your working directory.

Answer:

Import the data.

```
> insect_spray = read.csv('insect_spray.csv')
 > names(insect_spray)
  [1] "X"
             "count" "spray"
 > head(insect_spray)
   X count spray
 1 1
        10
               А
 2 2
         7
               А
        20
            Α
 3 3
        14
 44
               Α
 55
        14
               А
 6 6
        12
               Α
 > str(insect_spray)
  'data.frame': 72 obs. of 3 variables:
  $ X : int 1 2 3 4 5 6 7 8 9 10 ...
  $ count: int 10 7 20 14 14 12 10 23 17 20 ...
  $ spray: Factor w/ 6 levels "A", "B", "C", "D", ..: 1 1 1 1 1 1 1 1 1 1 ...
 > summary(insect_spray)
        Х
                     count
                                  spray
  Min. : 1.00 Min. : 0.00 A:12
  1st Qu.:18.75 1st Qu.: 3.00 B:12
Median :36.50 Median : 7.00 C:12
  Mean :36.50 Mean : 9.50 D:12
  3rd Qu.:54.25 3rd Qu.:14.25 E:12
  Max. :72.00 Max. :26.00 F:12
Run a simple linear model comparing the effectiveness of the different sprays.
 > mod1 = lm(count~spray, data=insect_spray)
```

```
> summary(mod1)
Call:
lm(formula = count ~ spray, data = insect_spray)
Residuals:
          1Q Median
                       3Q
  Min
                              Мах
-8.333 -1.958 -0.500 1.667 9.333
Coefficients:
           Estimate Std. Error t value Pr(>|t|)
(Intercept) 14.5000 1.1322 12.807 < 2e-16 ***
            0.8333
                       1.6011 0.520
sprayB
                                        0.604
           -12.4167 1.6011 -7.755 7.27e-11 ***
spravC
           -9.5833 1.6011 -5.985 9.82e-08 ***
sprayD
           -11.0000 1.6011 -6.870 2.75e-09 ***
2.1667 1.6011 1.353 0.181
sprayE
sprayF
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 3.922 on 66 degrees of freedom
Multiple R-squared: 0.7244, Adjusted R-squared: 0.7036
F-statistic: 34.7 on 5 and 66 DF, p-value: < 2.2e-16
```

Get predictions and their standard errors from this model for each spray type.

```
> new.data = data.frame(spray=c("A", "B", "C", "D", "E", "F"))
```

```
> pred1 = predict.lm(mod1, new.data, se.fit=T)
> summary(pred1)
              Length Class Mode
fit
              6
                     -none- numeric
se.fit
              6
                     -none- numeric
df
                    -none- numeric
              1
residual.scale 1
                    -none- numeric
> pred1
$fit
                 2
                           3
       1
                                     4
                                               5
14.500000 15.333333 2.083333 4.916667 3.500000 16.666667
$se.fit
               2
                        3
                                 4
                                          5
       1
                                                    6
1.132156 1.132156 1.132156 1.132156 1.132156 1.132156
$df
[1] 66
$residual.scale
[1] 3.921902
```

Run the model again as a GLM using the poisson distribution. Are the estimates different from model one?

```
> mod2 = glm(count~spray, data=insect_spray, family=poisson)
> summary(mod2)
Call:
glm(formula = count ~ spray, family = poisson, data = insect_spray)
Deviance Residuals:
   Min 10 Median
                           30
                                     Max
-2.3852 -0.8876 -0.1482 0.6063 2.6922
Coefficients:
          Estimate Std. Error z value Pr(>|z|)
(Intercept) 2.67415 0.07581 35.274 < 2e-16 ***
                    0.10574 0.528
sprayB
           0.05588
                                       0.597
          -1.94018 0.21389 -9.071 < 2e-16 ***
spravC
sprayD
          -1.08152 0.15065 -7.179 7.03e-13 ***
         -1.42139 0.17192 -8.268 < 2e-16 ***
sprayE
          0.13926 0.10367 1.343
                                       0.179
sprayF
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
(Dispersion parameter for poisson family taken to be 1)
   Null deviance: 409.041 on 71 degrees of freedom
Residual deviance: 98.329 on 66 degrees of freedom
```

```
AIC: 376.59

Number of Fisher Scoring iterations: 5

> anova(mod2)

Analysis of Deviance Table

Model: poisson, link: log

Response: count

Terms added sequentially (first to last)

Df Deviance Resid. Df Resid. Dev

NULL 71 409.04

spray 5 310.71 66 98.33
```

Get predictions and their standard errors (using the *predict.glm* function). How do the SEs compare to model one. Why are they so different?

```
> new.data = data.frame(spray=c("A", "B", "C", "D", "E", "F"))
> pred2 = predict.glm(mod2, new.data, se.fit=T, type="response")
> summary(pred2)
              Length Class Mode
fit
               6
                  -none- numeric
se.fit
               6
                      -none- numeric
                     -none- numeric
residual.scale 1
> pred2
$fit
                  2
                            3
        1
                                      4
                                                5
                                                          6
14.500000 15.333333 2.083333 4.916667 3.500000 16.666667
$se.fit
                  2
                            3
                                      4
                                                5
                                                          6
        1
1.0992422 1.1303883 0.4166664 0.6400955 0.5400617 1.1785113
$residual.scale
[1] 1
```

Save this matrix table of results as a *.csv* file in your working directory.

```
> store = matrix(0, nrow=6, ncol=4)
> store[,1] = pred1$fit
> store[,2] = pred1$se.fit
> store[,3] = pred2$fit
> store[,4] = pred2$se.fit
> write.csv(store, "insect_pred_table.csv")
> quit()
Save workspace image? [y/n/c]: n
$ cat insect_pred_table.csv
"", "V1", "V2", "V3", "V4"
"1",14.5,1.13215550799177,14.5,1.0992421631893
"2", 15.333333333333333, 1.13215550799177, 15.3333333333333, 1.13038833052086
"3"
   , 2.083333333333331, 1.13215550799177, 2.083333333333498, 0.416666404601133
"4"
   ,4.916666666666668,1.13215550799177,4.91666666666666666,0.640095478972524
"5", 3.5000000000001, 1.13215550799177, 3.49999999999999, 0.540061724814324
"6",16.666666666666667,1.13215550799177,16.6666666666666667,1.17851130197669
```

12.6 Exercise: Linear Modelling 2

Using the CO2.csv data which shows the uptake of carbon dioxide of two plant types when kept at different temperatures and at different concentrations of CO_2 .

- 1. Run a simple linear model (regression) comparing the uptake of CO₂ to the concentration of CO₂.
- Run a simple linear model (t-test) comparing the effect of temperature on CO₂ uptake.
- 3. Combine these two explanatory variables into a model that includes them both, and also look at one that includes an interaction between them.
- 4. Get predictions for the two types of plants, for a range of CO_2 concentrations.
- 5. Rerun the models and get predictions for a Gamma distribution.
- 6. Compare the different models and find the best model by looking at their Akaike Information Criterion (AIC).

Note: The AIC is an estimator of the relative quality of statistical models for a given set of data. Given a collection of models for the data, AIC estimates the quality of each model, relative to each of the other models. Thus, AIC provides a means for model selection. The AIC can be used to help decide which model is better at describing your data and making predictions from it. The lower the AIC, the better the model.

HINT: an example of using the function is *AIC(mod1, mod2, mod3)*.

Answer:

Import the data.

```
> co2 = read.csv('C02.csv')
 > names(co2)
 [1] "X"
                 "Plant"
                            "Type"
                                        "Treatment" "conc"
                                                               "uptake"
 > head(co2)
   X Plant
            Type Treatment conc uptake
 1 1 Qn1 Quebec nonchilled 95
                                   16.0
 22
       Qn1 Quebec nonchilled 175
                                   30.4
                                   34.8
      Qn1 Quebec nonchilled 250
 3 3
 44
       Qn1 Quebec nonchilled 350
                                   37.2
 55
       Qn1 Quebec nonchilled 500
                                   35.3
 6 6 Qn1 Quebec nonchilled 675
                                   39.2
 > str(co2)
 'data.frame': 84 obs. of 6 variables:
  $ X : int 1 2 3 4 5 6 7 8 9 10 ...
             : Factor w/ 12 levels "Mc1", "Mc2", "Mc3",..: 10 10 10 10 10 10 10 11 11
   $ Plant
11 ...
            : Factor w/ 2 levels "Mississippi",..: 2 2 2 2 2 2 2 2 2 2 ...
  $ Type
  $ Treatment: Factor w/ 2 levels "chilled", "nonchilled": 2 2 2 2 2 2 2 2 2 2 ...
  $ conc : int 95 175 250 350 500 675 1000 95 175 250 ...
  $ uptake : num 16 30.4 34.8 37.2 35.3 39.2 39.7 13.6 27.3 37.1 ...
```

```
> summary(co2)
                Plant
                                 Туре
                                            Treatment
     Х
                                                         conc
Min. : 1.00 Mc1 : 7 Mississippi:42 chilled :42 Min. : 95
1st Qu.:21.75 Mc2
                   : 7
                         Quebec :42 nonchilled:42 1st Qu.: 175
Median :42.50
              Mc3
                    : 7
                                                      Median : 350
             Mn1
Mean :42.50
                    : 7
                                                      Mean : 435
3rd Qu.:63.25 Mn2
                    : 7
                                                      3rd Qu.: 675
Max. :84.00 Mn3 : 7
                                                      Max. :1000
              (Other):42
   uptake
Min. : 7.70
1st Qu.:17.90
Median :28.30
Mean :27.21
3rd Qu.:37.12
Max. :45.50
```

Run a simple linear model (regression) comparing the uptake of CO_2 to the concentration of CO_2 .

```
> mod1 = lm(uptake~conc, data=co2)
> summary(mod1)
Call:
lm(formula = uptake ~ conc, data = co2)
Residuals:
   Min
           1Q Median
                          ЗQ
                                  Мах
-22.831 -7.729 1.483 7.748 16.394
Coefficients:
            Estimate Std. Error t value Pr(>|t|)
(Intercept) 19.500290 1.853080 10.523 < 2e-16 ***
         0.017731 0.003529 5.024 2.91e-06 ***
conc
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 9.514 on 82 degrees of freedom
Multiple R-squared: 0.2354, Adjusted R-squared: 0.2261
F-statistic: 25.25 on 1 and 82 DF, p-value: 2.906e-06
```

Run a simple linear model (t-test) comparing the effect of temperature on CO_2 uptake.

```
> mod2 = lm(uptake~Treatment, data=co2)
> summary(mod2)
Call:
lm(formula = uptake ~ Treatment, data = co2)
Residuals:
    Min
            1Q Median
                             30
                                      Мах
-20.0429 -8.6530 -0.4429 9.7321 18.6167
Coefficients:
                  Estimate Std. Error t value Pr(>|t|)
                   23.783 1.591 14.948 <2e-16 ***
(Intercept)
Treatmentnonchilled 6.860
                              2.250 3.048 0.0031 **
- - -
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
Residual standard error: 10.31 on 82 degrees of freedom
Multiple R-squared: 0.1018, Adjusted R-squared: 0.09084
```

F-statistic: 9.293 on 1 and 82 DF, p-value: 0.003096

Combine these two explanatory variables into a model that includes them both, and also look at one that includes an interaction between them.

```
> mod3a = lm(uptake~conc + Treatment, data=co2)
 > mod3b = lm(uptake~conc * Treatment, data=co2)
 > summary(mod3a)
 Call:
 lm(formula = uptake \sim conc + Treatment, data = co2)
 Residuals:
             1Q Median
                             30
     Min
                                    Мах
  -19.401 -7.066 -1.168 7.573 17.597
 Coefficients:
                     Estimate Std. Error t value Pr(>|t|)
                   16.070528 1.989746 8.077 5.31e-12 ***
 (Intercept)
                     0.017731 0.003306 5.364 7.55e-07 ***
 conc
 Treatmentnonchilled 6.859524 1.944840 3.527 0.000695 ***
 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
 Residual standard error: 8.912 on 81 degrees of freedom
 Multiple R-squared: 0.3372, Adjusted R-squared: 0.3208
 F-statistic: 20.6 on 2 and 81 DF, p-value: 5.837e-08
 > summary(mod3b)
 Call:
 lm(formula = uptake \sim conc * Treatment, data = co2)
 Residuals:
             1Q Median
     Min
                          ЗQ
                                    Мах
  -18.218 -7.401 -1.117 7.835 17.209
 Coefficients:
                          Estimate Std. Error t value Pr(>|t|)
                          16.981416 2.464160 6.891 1.15e-09 ***
 (Intercept)
                          0.015637 0.004693 3.332 0.00131 **
 conc
 Treatmentnonchilled
                          5.037747
                                     3.484848
                                                1.446 0.15219
 conc:Treatmentnonchilled 0.004188 0.006636 0.631 0.52979
 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
 Residual standard error: 8.946 on 80 degrees of freedom
 Multiple R-squared: 0.3405, Adjusted R-squared: 0.3157
 F-statistic: 13.77 on 3 and 80 DF, p-value: 2.528e-07
Get predictions for the two types of plants, for a range of CO_2 concentrations.
```

8

```
> predict.lm(mod4, new.data, se.fit=T)
 $fit
        1
                2
                        3
                                1
                                         5
                                                 6
                                                          7
 25,81104 16,73566 28,11905 17,97377 30,42705 19,21188 32,73506 20,44999
                       11 12 13 14 15
        9
              10
                                                                 16
 35.04306 21.68811 37.35106 22.92622 39.65907 24.16433 41.96707 25.40245
       17 18 19 20
 44.27508 26.64056 46.58308 27.87867
 $se.fit
                2
                        3
                                4
                                         5
                                                 6
                                                          7
       1
 1.622000 1.622000 1.369811 1.369811 1.177546 1.177546 1.077770 1.077770
       9
          10 11 12 13 14
                                                        15
                                                                 16
 1.096037 1.096037 1.227089 1.227089 1.440463 1.440463 1.705538 1.705538
                               20
       17
              18
                       19
 2.001880 2.001880 2.317526 2.317526
 $df
 [1] 80
 $residual.scale
 [1] 6.935822
Rerun the models and get predictions for a Gamma distribution.
 > mod5 = glm(uptake~conc * Type, data=co2, family=Gamma)
 > summary(mod5)
 Call:
 glm(formula = uptake ~ conc * Type, family = Gamma, data = co2)
 Deviance Residuals:
      Min
             10
                      Median
                                   3Q
                                           Мах
  -0.90639 -0.22457 -0.01455 0.19478 0.54514
 Coefficients:
                 Estimate Std. Error t value Pr(>|t|)
                6.020e-02 4.361e-03 13.803 < 2e-16 ***
 (Intercept)
             -2.532e-05 6.898e-06 -3.671 0.000434 ***
 conc
                -2.148e-02 5.156e-03 -4.166 7.78e-05 ***
 TypeQuebec
 conc:TypeQuebec 7.311e-06 8.094e-06 0.903 0.369081
 Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
 (Dispersion parameter for Gamma family taken to be 0.0939836)
     Null deviance: 15.8858 on 83 degrees of freedom
 Residual deviance: 8.4588 on 80 degrees of freedom
 AIC: 596.1
 Number of Fisher Scoring iterations: 5
 > new.data = expand.grid(Type=c("Quebec", "Mississippi"),
                           conc=seq(100,1000,100)
                           )
```

> predict.glm(mod5, new.data, se.fit=T, type="response") \$fit 3 1 5 6 7 1 2 8 27.08678 17.34099 28.47595 18.13741 30.01533 19.01050 31.73064 19.97190 9 10 11 12 13 14 15 16 33.65390 21.03572 35.82534 22.21925 38.29633 23.54390 41.13343 25.03650 17 18 19 20 44.42452 26.73116 48.28806 28.67189 \$se.fit 2 3 4 5 6 7 8 1 1.7561008 1.1382195 1.6721912 1.0702451 1.5951945 1.0088967 1.5520705 0.9732649 9 10 11 12 13 14 15 16 1.5921838 0.9959336 1.7853024 1.1180932 2.2010333 1.3739928 2.8953715 1.7851440 18 19 20 17 3.9288835 2.3729936 5.3967264 3.1730722\$residual.scale [1] 0.3065674

Compare the different models and find the best model by looking at their AIC.

13. Plots

Simple plot. *plot()* is a generic base graphic function for the plotting of *R* objects. *x* and *y* objects must be given to the function.

```
> x = 1:5
> y = 1:5
> plot(x,y)
```



Illustration 19: Simple plot

plot(x=c(1,2,3,4), y=c(1,2,3,4))

> args(plot) function (x, y, ...) NULL

Data Analytics

13.1 Beautify the plot

Add type to adjust the view of the plot. What type of plot should be drawn. Possible types are:

- **p** points
- I lines
- **b** both
- c the lines part alone of b
- o both over|plotted
- h histogram like (or high|density) vertical lines
- s stair steps
- n no plotting

> y = 1:5



Illustration 20: Plot with lines

Add some colour.

> plot(x,y, type = "b", col = "green")



Illustration 21: Plot with colour

It also works as a general plot function for many object types. See Illustration 22 for a simple linear regression.

> x = c(1,2,2,3,2,3,4,3,4,5,3)
> y = c(4,8,6,3,5,7,9,2,1,7,4)
> model.1 = lm(y~x)
> class(model.1)
[1] "lm"
> plot(model.1)



Illustration 22: Plot - model1

13.1.1 Plot layers

Add some other values.

> plot(x,y, type = "b", col = "green",xlab = "bottom", ylab = "left")

> title(main = "My plot", xlab = "length", ylab = "height")



Illustration 23: Plot layers

Notice how each or these lines layered on to of each-other. It is necessary to turn off some things in the first line.

> plot(x,y, type = "b", col = "green",xlab = "", ylab = "", xaxt="n", xaxt="n")

> title(main = "My plot", xlab = "length", ylab = "height")







13.1.2 Box type

Use the argument *box type* [bty] to adjust the box.

- o: The default value draws a complete rectangle around the plot.
- **n:** Draws nothing around the plot.
- I, 7, c, u, or]: Draws a shape around the plot area that resembles the uppercase letter of the option. So, the option bty="I" draws a line to the left and bottom of the plot.

```
> plot(rnorm(100))
```

- > plot(rnorm(100),bty="o")
- > plot(rnorm(100), bty="l")
- > plot(rnorm(100), bty="7")
 > plot(rnorm(100), bty="7")
- > plot(rnorm(100), bty="c")
 > plot(rnorm(100), bty="l")
- > plot(rnorm(100),bty="]")
 > plot(rnorm(100),bty="n")
- > piot(rnorm(100), bty="n")



Illustration 25: Box type

13.1.3 Splitting arguments

```
> plot(x,y, xlab="height", ylab="length", cex.axis= 1.2,
          cex.lab=1.5, typ="p", col="red", bty="l", pch=16,
          tck=0.03
         )
> plot(x,y,
                                     # call the plot
       xlab="height", ylab="length", # label the axes
       xlim=c(0,10),
                                      # set specific limits to x-axis
       cex.axis= 1.2, cex.lab=1.5,
                                      # set character size for axis & labels
       typ="p", col="red",
                                      # set plot type & colour
       bty="1",
                                      # set box type around plot
       pch=16,
                                      # set point character type
       tck=0.03
                                      # set axis tick marks (+ve is inside plot)
      )
            S
            4
       ength
           ო
            2
                              2
                                           3
                                                                     5
                 1
                                                        4
                                        height
```

Illustration 26: Plot - splitting arguments

13.1.4 Graphical parameters: par

par(): can be used to set or query graphical parameters. Parameters can be set by specifying them as arguments to *par* in *tag* = *value* form, or by passing them as a list of tagged values. *par()* on its own returns the current settings for default graphical parameters. These defaults can be modified in *par()*. Some graphical parameters must be set in *par()* like background colour.

```
> par(lty=2,  # Set the line type
    pch=17,  # Define the plotting symbol
    cex.axis=3  # Specify the size of the tick labels
    )
```

Icon type: PCH

Sets the icon type.

pch = 0	square	pch = 13	circle cross
pch = 1	circle	pch = 14	square and triangle down
pch = 2	triangle point up	pch = 15	filled square
pch = 3	plus	pch = 16	filled circle
pch = 4	Cross	pch = 17	filled triangle point up
pch = 5	diamond	pch = 18	filled diamond
pch = 6	triangle point down	pch = 19	solid circle up
pch = 7	square cross	pch = 20	bullet (smaller circle)
pch = 8	star	pch = 21	filled circle blue
pch = 9	diamond plus	pch = 22	filled square blue
pch = 10	circle plus	pch = 23	filled diamond blue
pch = 11	triangles up and down	pch = 24	filled triangle point up blue
pch = 12	square plus	pch = 25	filled triangle point down blue

13.1.5 Colours

R has 657 colours to choose from. The colours() function gives a list of the available colour names.

colours(): Creates a list of all available colours.

```
colours = colours()
> head(colours)
[1] "white" "aliceblue" "antiquewhite" "antiquewhite1"
[5] "antiquewhite2" "antiquewhite3"
```

rainbow(n): Creates a vector of *n* contiguous colors.

```
> rainbow = rainbow(5)
> rainbow
[1] "#FF0000FF" "#CCFF00FF" "#00FF66FF" "#0066FFFF" "#CC00FFFF"
```

grey(level): Creates a vector of colours from a vector of gray levels. These are given as a vector of desired gray levels between zero and one; zero indicates *black* and one indicates *white*.

```
> grey = grey(c(0.1,0.3,0.5,0.7,0.9))
> grey
[1] "#1A1A1A" "#4D4D4D" "#808080" "#B3B3B3" "#E6E6E6"
```

13.1.6 text

text(): Insert text at any position of a current open plot.

```
> text(x, y, # x,y plot co-ordinates for the text
labels, # Text to be inserted
pos, # Position (1,2,3,4=below,left,above,right)
offset, # Distance of pos offset from x,y
col, cex, font ... # Other options
)
```

Example:

```
> plot(1,1)
> text(1,1.05,
    "This is a dot in the middle",
    col="red",cex=0.8
    )
```

```
> text(1,1,
    "This is the test to be printed",
    pos=1,offset=1,col="red",cex=0.8
)
```



Illustration 27: Plot text

Another example.



0

Illustration 28: Plot text 2

13.1.7 Points

points(): Insert points at any position of a current open plot.

points(x, y,	
pch,	<pre># Point character type</pre>
cex,	<pre># Character expansion - i.e size</pre>
col,	<pre># Colour of the point</pre>
bg	# Background colour or point
)	

Example:

```
> plot(c(0,2),c(0,25), type='n')
```

```
> points(x=rep(1,25), y=1:25, pch=1:25)
```



Illustration 29: Plot - points
13.1.8 Symbols

symbols(): Insert a shape at any position of a currently open plot.

```
symbols(x, y, # x,y plot co-ordinates
```

circles,	#	Draw circle with of diameter (e.g. circle=0.8)
squares,	#	Draw square of side length (e.g. square=0.5)
rectangles,	#	Draw rectangle (side lengths specified by matrix)
stars,	#	Draw star (points etc specified by 3 column matrix)
thermometers,	#	Craw with matrix of 3 or 4 columns
boxplots,	#	Draw with matrix of 5 columns
add=T,	#	Adds the symbol to the current plot
bg, fg, lwd	#	General settings - see help(symbols)
)		

```
> plot(c(0,4),c(0,6), type='n')
```

```
> symbols(x=1, y=3, circles=3,
            bg='red', fg='green',
            lwd=3, add=T
)
```



Illustration 30: Plot - symbols

13.1.9 Segments

segments(): Insert line segment at any position of a currently open plot.



Illustration 31: Plot - lines

13.1.10 Polygon

polygon(): Insert any shape at any position of a currently open plot.

> polygon(x,		# Vector of x co-ordinates for each point to be joined to m	ake
the shape			
У,		<pre># Vector of corresponding y co-ordinates for the shape (matc</pre>	:hed
to x)			
den	sity, #	Density of shading lines that fill the object	
ang	le, #	Slope of shading lines that fill the object	
col	, #	Colour of shading lines (if density=NA then col is fill colou	ur)
bor	der, #	Colour of the border	
lwd	#	Line width	
)			

- > plot(c(0,8),c(0,3), type='n')
- > polygon(c(1,1:7,7), c(0,1,2,1,2,1,2,1,0), col='blue', lwd=3)
- > polygon(c(1,7,7), c(0,0,1), col='yellow', angle=90, density=7)



Illustration 32: Plot - polygons

13.1.11 Arrows

arrows(): Draw an arrow between pairs of coordinates in a current open plot.

- > plot(c(0,8),c(0,3), type='n')
- > arrows(x0=1,y0=1,x1=6,y1=2,col='dark green',lty=2, lwd=3)



Illustration 33: Plot - arrows

13.1.12 Lines

- **lines():** A generic function taking coordinates given in various ways and joining the corresponding points with line segments.
- abline(): This function adds one or more straight lines through the current plot. It can specify intercept and slope, horizontal or vertical and can take intercept & slope from an *Im* object.
- **curve()** Draws a curve corresponding to a function over the interval *from, to. curve()* can plot also an expression in the variable *xname*, default *x*. Can add the line of an equation to a plot. Note: must set *add=TRUE*.

Example:

- > plot(c(0,8),c(0,8), type='n')
- > lines(x=c(1,5,8), y=c(2,3,1), col='blue', lwd=3)
- > x=c(1,2,3,4,5); y=c(2,2,1,1,0)
- > abline(lm(y~x))
- > curve(x+0.1*x^2, add=T)



Illustration 34: Plot - lines, curves

Example: abline()

<pre>> plot(1:21,-10:10)</pre>	
<pre>> abline(h=0, lty=2)</pre>	<pre># Put horizontal line at y=0</pre>
<pre>> abline(v=5, lty=1)</pre>	<pre># Put vertical line at x=5</pre>



Illustration 35: Plot - abline

13.1.13 Identity

Identify and label a point on a scatter-plot. Use the cursor over a point and it will identify it.

- > x=rnorm(100,0,2)
- > y=rnorm(100,3,3)
- > plot(x,y)
- > identify(x, y, plot=TRUE)



Illustration 36: Plot - identity

13.1.14 Adding equations / Greek letters to graphs

expression(): Creates or tests for objects of mode expression.

examples:

- > plot(c(0,10),c(0,10), type='n')
- > text(x=5, y=8, expression(lambda == 1.3))
- > text(x=5, y=6, expression(bar(X)[female]==0.55))
- > text(x=5, y=4, expression(y[3] ~ x^2 ~ m^-2))



Illustration 37: Plot - expression

13.2 Boxplots

A *box plot* or *boxplot* is a method of representing statistical data on a plot. It consists of a rectangle drawn to represent the second and third quartiles, usually with a vertical line inside to indicate the median value. The lower and upper quartiles are shown as horizontal lines either side of the rectangle.



Illustration 38: Boxplot

Figure 37: Plot - boxplot

Considering the following example. As can be seen from the summary(v) the minimum and maximum points are marked at 5 and 425. The median of 152 is flanked by the lower quartile at 69.5 and the upper quartile at 272.5.

> v = c(101,111,112,123,141,152,193,141, 51,19,43,74,45,26,83,42,65,32,5, 322,354,385,377,381,314,425,416, 214,233,234,226,237,248,269,276)

```
> summary(v)
Min. 1st Qu. Median Mean 3rd Qu. Max.
5.0 69.5 152.0 184.9 272.5 425.0
```



Illustration 39: Boxplot 2

```
> boxplot(v)
```



Illustration 40: Boxplot 3

13.3 Saving

13.3.1 Data

Data can be saved in a number of formats, in this case it is saved in *.csv* formatted file. In the example below the plot is saved to: *28-Sep-2018 00.38-graph name.csv*.

13.3.2 Plots

Plots can also be saved from graphical format to file. In the example below the plot is saved as a *.png* to: *28-Sep-2018_00.35-graph_name.png*. The graphic can also be saved in formats like *.pdf*.

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13.4 Exercise 1: Making a mess

```
> plot(x=1:100,y=rnorm(100))
```



Illustration 41: Plot exercise - start

Taking Illustration 41 apply graphical features liberally to make it as ugly as possible ©.

Adjust the default background colour.

```
> par(bg = "green")
                       # Set background to green
> plot(x=1:100,y=rnorm(100),
       pch = 13,
                               # Change icons to triangles
       cex = 2,
                               # Enlarge icon to 200%
       1wd = 3,
                               # Change line width
       col="orange",
                               # Icon colour
       bg="yellow"
                               # Icon background colour
       col.main = "orange",
                               # Change title colour
       col.axis = "red",
                               # Change axis colour
       col.lab = "white",
                               # Change label colour
       font.axis = 3,
                               # Change axis font
       font.lab = 4,
                               # Change label font
       family = "sans",
                               # Change font
      )
```

Locate a point on the plot with the *locator()* function. Run the function and click on the plot, it will return the location of the cursor. For the text line the shell will hold until the point is clicked.

```
> locator(1)
$x
[1] -16.85286
$y
[1] 3.984065
```

```
> text(locator(1), "A bit of text is here")
> lines(locator(2),col="white",lty=1,lwd=3) # Draw a line
> lines(locator(2), col="black", lty=1, lwd=3) # Draw a line
> locator(2)
$x
[1] 52.10779 51.52666
$y
[1]
    2.06091 -2.54633
> points(x=rep(1,25),y=1:25, type= 'n')
# Draw a line
> segments(52.10779, 2.06091, 51.52666, -2.54633,col="red",lty=2,lwd=4)
# Draw a line
> lines(c(4.455207, 96.273600), c(-0.33893, -0.30497) ,col="yellow",lty=2,lwd=4)
> title("Is it Saint Patricks Day") # Add a title
> polygon(c(1,1:7,7),
                                     # Add a polygon
          c(0,1,2,1,2,1,2,1,0),
          col='blue', lwd=3
         )
```



Illustration 42: Plot - mess

13.5 Exercise 2: Create a boxplot

As has been demonstrated in section 13.2 Boxplots R comes with a *boxplot()* function as in Illustration 43. Assuming there is no such function create a boxplot for the output of the function *rnorm*(100,5,1.5) using the other tools available.

```
> a = rnorm(100,5,1.5)
> boxplot(a)
```





```
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```

```
Answer:
 > a = rnorm(100,5,1.5)
 > mean_a = mean(a)
 > sd_a = sd(a)
 > \min_a = \min(a)
 > max_a = max(a)
 > plot(c(0, 0, min_a, min_a),
                                  # Plot area
        c(0,max_a,max_a,0),
        ylim = c(0,10), xaxt = 'n', yaxt = 'n',
         type = 'n', xlab = '', ylab = ''
        )
                            # Dotted line line
  > segments(1, min_a,
          1, max_a,
           col="black",lty=2,lwd=1
         )
  > segments(0.9,max_a,
                               # Top line
           1.1, max_a,
           col="black",lwd=1
           )
  > segments(0.9,min_a,
                           # Bottom line
          1.1,min_a,
          col="black",lwd=1
          )
  > polygon(c(0.8, 1.2, 1.2, 0.8), # Draw polygon
            c(mean_a - sd_a, mean_a - sd_a, mean_a + sd_a, mean_a + sd_a),
            border = "black",
            col = "white",
            lwd=1
           )
  > segments(0.8,mean_a, 1.2, mean_a,
                                         # Draw middle line
          col="black",lwd=4
          )
```

```
> axis(2, at=c(1,2,3,4,5,6,7,8), labels=c(1,2,3,4,5,6,7,8))
```



Illustration 44: Boxplot answer

13.6 Multiple graphs

13.6.1 par() function again

With the *par()* function, the options *mfrow=c(nrows, ncols)* can be included to create a matrix of *nrows x ncols* plots that are filled in by row. *mfcol=c(nrows, ncols)* fills in the matrix by columns. To plot the model *model.1* in the example will shuffle through four plots.

- Residuals vs Fitted
- Normal Q-Q
- Scale-Location
- Residuals vs Leverage
- > x = c(1,2,2,3,2,3,4,3,4,5,3)

```
> y = c(4,8,6,3,5,7,9,2,1,7,4)
```

```
> model.1 = lm(y ~ x)
```

```
> plot(model.1)
```

Hit <Return> to see next plot: Hit <Return> to see next plot:

However by adjusting the graphical parameters it is possible to create a top-level graphical plot area that will include the four plots on the one main plot area. As there are four plots then a 2×2 main plot frame is required.

```
> x = c(1,2,2,3,2,3,4,3,4,5,3)
```

```
> y = c(4,8,6,3,5,7,9,2,1,7,4)
```

```
> model.1 = lm(y ~ x)
```

```
> par(mfrow=c(2,2))  # Adding this line creates the top-level plot area
```

```
> plot(model.1)
```





13.7 Exercise 3a: Setting graph parameters

Using these data, create a publication quality graph like this.

HINT: to get the axis tick marks on the inside use the *tck* argument in your plot call (e.g. *tck=0.03*).

```
> x = 1:20
> y = seq(from=3, to=7, length.out=20)+rnorm(20,0,2)
```

Answer:



Illustration 46: Exercise - setting graph parameters

13.8 Exercise 3b: Setting graph parameters

Add a second plot using these additional data.

```
> x = 1:20
> proportion.females = runif(20,0.3,0.7)
> y = proportion.females
> plot(x,y, type='1',
       xlab = 'Years since establishment',
       ylab = 'Proportion of females in the population',
       xaxt = 'n', yaxt = 'n',
       bty = '1',
       lty = 2,
       xlim = c(0, 20),
       ylim = c(0, 1),
       font.lab=2
      )
> axis(1, seq(0,20,5),las=2, tck = 0.02)
> axis(2, seq(0,1,0.2),tck = 0.02)
> text(x=1, y=0.98, "b", cex=1.5)
> text(x=10, y=0.15, expression(bar(X)[female]==0.55), cex=1.3)
```



Illustration 47: Exercise - setting graph parameters 2

Create a multiple frame to hold both graphs.

```
mtext(): Write Text into the Margins of a Plot.
```

```
# Data to be plotted
> x = 1:20
> y = seq(from=3, to=7, length.out=20)+rnorm(20,0,2)
> z = runif(20,0.3,0.7)
# First graphic area (left side)
> par(fig=c(0,0.5,0,1))
# Plot the points on first graph
> plot(x,y, type='n',
       xlab = ''.
       ylab = 'Population size (x 1000)',
       xaxt = 'n', yaxt = 'n',
       bty = '1'
       xlim = c(0, 20),
       ylim = c(0, 10),
       font.lab=2
      )
# Redefine the points
> points(x,y, pch=21, bg="black", cex=2)
# Add the first graph axis
> axis(1, seq(0,20,5),tck = 0.02)
> axis(2, seq(0,10,2), tck = 0.02)
# Add the "a" and the function term to 1st graph
> text(x=1, y=9.8, "a", cex=1.5)
> text(x=10, y=9, expression(lambda == 1.3), cex=1.5)
# Second graphic area (right side)
> par(fig=c(0.5,1,0,1), new=T)
# Plot the dashed line
> plot(x,z, type='l',
       xlab = '',
       ylab = 'Proportion of females in the population',
       xaxt = 'n', yaxt = 'n',
       bty = '1',
       lty = 2,
       xlim = c(0, 20),
       ylim = c(0, 1),
       font.lab=2
      )
# Add in the axis graphs
> axis(1, seq(0,20,5),tck = 0.02)
> axis(2, seq(0,1,0.2),tck = 0.02)
# Add the "b" and the function term to 2nd graph
> text(x=1, y=0.98, "b", cex=1.5)
> text(x=10, y=0.15, expression(bar(X)[female]==0.55), cex=1.3)
# Add the common text under both graphs
> mtext("Years since establishment", side = 1, line = -2, outer = TRUE, cex=1.3)
```



Illustration 48: Exercise - setting graph parameters 3

13.9 Exercise 4a: Prediction plots

Using the model *Im(begging~sex+food+sex*food, data = owl)*

Extract predictions (with their standard errors) for both males and females, for a range of food values from 20-30 using the *predict()* function.

```
# Add libraries
> library(scales)
# Import the data
> owl = read.csv('owl_data.csv')
> owl.lm = lm(begging ~ sex + food + sex * food, data = owl)
> male_predict = data.frame(sex=rep('Male',11), food=10:20)
> female_predict = data.frame(sex=rep('Female',11), food=10:20)
> male_out = predict.lm(owl.lm, male_predict, se.fit=T)
> female_out = predict.lm(owl.lm,female_predict, se.fit=T)
# Plot 1
# Generate the first plot
> plot(x=seq(10,20),y=male_out$fit,type='n',
       xlab='Time since last meal',
       ylab='Begging rate',
       bty='l', ylim=c(8,25), cex.lab=1.2
      )
# Add lines to plot
> lines(x=seq(10,20),male_out$fit,
        type='b', pch=16, col='blue'
       )
> lines(x=seq(10,20),female_out$fit,
        type='b', pch=16, col='red'
       )
# Plot 2: Add Standard Errors (SE) lines on the plot
# Add SE lines for males
> lines(x=seq(10,20),male_out$fit+male_out$se.fit,
        type='l', col='blue', lty=2
       )
> lines(x=seq(10,20),male_out$fit-male_out$se.fit,
        type='1', col='blue', lty=2
       )
# Add SE lines for females
> lines(x=seq(10,20),female_out$fit+female_out$se.fit,
        type='l', col='red', lty=2
       )
> lines(x=seq(10,20),female_out$fit-female_out$se.fit,
        type='l', col='red', lty=2
       )
# Plot 3: now add SE polygons
# Add SEs for males
> polygon(x=c(10:20, 20:10),
          y=c(male_out$fit+male_out$se.fit,
          rev(male_out$fit-male_out$se.fit)),
          col=alpha("blue", 0.3), border="blue"
```



Illustration 49: Exercise - prediction plots

13.10 Exercise 4b: Prediction plots

Modify the previous model to a Poisson distribution.

glm(begging~sex+food+sex*food, data=owl, family=poisson)

Repeat exercise 4a, but using the predictions from the model based on a poisson distribution.

HINT:check if your predictions are at the log scale and need to be exponentiated. check the predict function argument *type*. This needs to be set to *response*.

```
# Add libraries
> library(scales)
# Import the data
> owl = read.csv('owl_data.csv')
> owl.lm = glm(begging ~ sex+food+sex*food, data=owl, family=poisson)
> male_predict = data.frame(sex=rep('Male',11), food=10:20)
> female_predict = data.frame(sex=rep('Female',11), food=10:20)
> female_out = predict.glm(owl.lm, male_predict, se.fit=T, type="response")
> male_out = predict.glm(owl.lm,female_predict, se.fit=T, type="response")
# Plot 1
# Generate the first plot
> plot(x=seq(10,20),y=male_out$fit,type='n',
       xlab='Time since last meal',
       ylab='Begging rate',
       bty='1', ylim=c(0,60), cex.lab=1.2
      )
# Add lines to plot
> lines(x=seq(10,20),male_out$fit,
        type='b', pch=16, col='blue'
       )
> lines(x=seq(10,20),female_out$fit,
        type='b', pch=16, col='red'
       ۱
# Plot 2: Add Standard Errors (SE) lines on the plot
# Add SE lines for males
> lines(x=seq(10,20),male_out$fit+male_out$se.fit,
        type='l', col='blue', lty=2
       )
> lines(x=seq(10,20),male_out$fit-male_out$se.fit,
        type='l', col='blue', lty=2
       )
# Add SE lines for females
> lines(x=seq(10,20),female_out$fit+female_out$se.fit,
        type='l', col='red', lty=2
       )
> lines(x=seq(10,20),female_out$fit-female_out$se.fit,
        type='l', col='red', lty=2
       )
```

```
# Plot 3: now add SE polygons
# Add SEs for males
> polygon(x=c(10:20, 20:10),
          y=c(male_out$fit+male_out$se.fit,
          rev(male_out$fit-male_out$se.fit)),
          col=alpha("blue", 0.3), border="blue"
         )
# Add SEs for females
> polygon(x=c(10:20, 20:10),
          y=c(female_out$fit+female_out$se.fit,
          rev(female_out$fit-female_out$se.fit)),
          col=alpha("red", 0.3), border="red"
         )
# Add text to the plot
> text(16,40,'Female', cex=1.5, col='red')
> text(15,9,'Male', cex=1.5, col='blue')
```



Illustration 50: Exercise - prediction plots 2

13.11 Exercise 5: Visualising plot data

Import the *bird_egg.csv* file.

Plot the relationship between eggs (y) and age (x) for clutch 1 (blue points) & clutch 2 (red points) and save it as a pdf file.

HINT: *jitter()* could help visualise the points here eggs is the response variable (y) and age the explanatory (x).

EXTRA: extract predictions from a model where age and clutch (and their interaction) explain the number of eggs. Plot these predictions with their standard errors over the scatter-plot above.

```
> install.packages('scales')
# Add libraries
> library(scales)
# Import the data
> bird = read.csv('bird_egg.csv')
> names(bird)
#[1] 'individual'
                    'year'
                            'clutch'
                                         'age'
                                                  'eggs'
                                                           'dist_food'
#[7] 'fail_fledge'
# Plot the points and
> x1 = bird[bird[,3]==1,4] # Plot the 'age' for the 1st clutch
> y1 = bird[bird[,3]==1,5] # Plot the 'eggs' for the 1st clutch
> x2 = bird[bird[,3]==2,4] # Plot the 'age' for the 2nd clutch
> y2 = bird[bird[,3]==2,5] # Plot the 'eggs' for the 2nd clutch
# Plot the eggs as a DV for age as the IV for 1st clutch
> plot(x1,y1,col='blue',
         xlab='Age', ylab='Eggs'
      )
# Plot the eggs as a DV for age as the IV for 2nd clutch
> points(x2,y2, col='red')
# Add a small amount of noise to the vector.
> points(jitter(x1), jitter(y1), col='blue')
> points(jitter(x2), jitter(y2), col='red')
# Get data for clutch 1 & 2 (poisson)
> bird1 = bird[bird[,3] < 3,]</pre>
> mod1 = glm(eggs~age * as.factor(clutch),
             data=bird1, family='poisson'
> summary(mod1)
# Extract dataframe for 'age' and 'clutch'
> new_data1 = data.frame(age=1:10, clutch=rep(1,10))
> new_data2 = data.frame(age=1:10, clutch=rep(2,10))
# Get predictions
> pred_clutch1 = predict.glm(mod1, new_data1, se.fit=T, type='response')
> pred_clutch2 = predict.glm(mod1, new_data2, se.fit=T, type='response')
# Add clutch1 prediction
# Add lines
> lines(x = 1:10, y = pred_clutch1$fit,
        lty = 1, lwd = 2, col = 'blue'
```

```
)
# Add blue filled in pologon area
> polygon(x = c(1:10, 10:1),
          y = c(pred_clutch1$fit+pred_clutch1$se.fit,
          rev(pred_clutch1$fit-pred_clutch1$se.fit)),
          col = alpha('blue',0.3), border='blue'
         )
# Add clutch2 prediction
# Add lines
> lines(x = 1:10, y = pred_clutch2$fit,
        lty = 1, lwd = 2, col = 'red'
        )
# Add red filled in pologon area
> polygon(x = c(1:10, 10:1),
          y = c(pred_clutch2$fit+pred_clutch2$se.fit,
          rev(pred_clutch2$fit-pred_clutch2$se.fit)),
          col = alpha('red',0.3), border='red'
         )
}
# Add labels
> text(x = 6, y = 5.5, 'Clutch 1',
       cex = 1.5, col = 'blue'
      )
> text(x = 6, y = 2, 'Clutch 2',
       cex = 1.5, col = 'red'
      )
```



Illustration 51: Exercise - visualising plot data

13.12 Exercise 6: Pretty plot

From the data $xy_data_quadplotex.csv$ produce a graph. The model is $y \sim x + x 2$.

HINT: Get the fitted values for the model prediction.

Look at the data
<pre>> read.csv('xy_data_quadplotex.csv')</pre>
х у
1 1 3.9257408
2 2 1.9763139
3 3 3.1513985
4 4 2 6076583
6 6 5.7526961
7 7 6.3944896
8 8 5.4130619
9 9 9.4346628
10 10 9.7635863
11 11 8.9396002
12 12 7.2992594
13 13 9.1645094
14 14 8.1535297
15 15 7 0302138
10 10 9.4554672
17 17 0.2583997
18 18 6.5865369
19 19 5.7238197
20 20 6.0263360
21 21 3.0479905
22 22 3.3684445
23 23 4.5477625
24 24 3.1443438
25 25 -0.3731705
26 26 0.9693107
27 27 -5 7287515
28 28 -3 /355710
20 20 -3.4333713
29 29 -2.9714092
30 30 -7.3180748
Import the data
<pre>> xy = read.csv('xy_data_quadplotex.csv')</pre>
Create the initial plot with values
<pre>> plot(x,y)</pre>
Create linear model
$> mod.1 = lm(xv$v ~ xv$x + I(xv$x^2))$
> v fit = fitted(mod 1)
> y.iit = iitteu(mou.i)
Dray the fit line on plot
Draw the fit fine on piot
<pre>> lines(xy\$x,y.tit, lwd=2)</pre>
<pre>> get.y = matrix(c(xy\$x,y.fit), ncol=2)</pre>
Show area under curve for range 1 - 7
> x1.range <- 1:7
> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2]
> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2]
<pre>> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2] # Show area under curve for range 7 - 20</pre>
<pre>> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2] # Show area under curve for range 7 - 20 > x2.range <- 7:20</pre>
<pre>> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2] # Show area under curve for range 7 - 20 > x2.range <- 7:20 > y2.range <- get.y[get.y[1]>=7 & get.y[1]<=20 2]</pre>
<pre>> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2] # Show area under curve for range 7 - 20 > x2.range <- 7:20 > y2.range <- get.y[get.y[,1]>=7 & get.y[,1]<=20,2]</pre>
<pre>> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2] # Show area under curve for range 7 - 20 > x2.range <- 7:20 > y2.range <- get.y[get.y[,1]>=7 & get.y[,1]<=20,2] # Show area under curve for range 20 - 20</pre>
<pre>> y1.range <- get.y[get.y[,1]>=1 & get.y[,1]<=7,2] # Show area under curve for range 7 - 20 > x2.range <- 7:20 > y2.range <- get.y[get.y[,1]>=7 & get.y[,1]<=20,2] # Show area under curve for range 20 - 30 > x2 range <- 20:20</pre>

```
# Add labeling to each shaded area
> text(3.5,-3, "Treatment\nA", cex=1)
> text(12.5,-3, "Treatment\nB", cex=1)
> text(24,-3, "Treatment\nC", cex=1)
```



Illustration 52: Exercise - pretty plot

13.13 Exercise 7: Icon and colour table

```
# Add the plot
> plot(x = c(0,1.2),y = c(0,25),
      col = "white", xlab = "", ylab = "",
      xaxt = "n",yaxt = "n"
      )
# Add points for icons
> points(x = rep(0.1,25),
        y = 1:25, pch = 1:25,
        cex = 1.2
        )
# Add text for icons
> text(x = rep(0.1,25),
      y = 1:25, paste("pch", 1:25),
      pos = 4, offset = 1
      )
# Add points for colour
> points(x = rep(0.4,8),
        y = 10:17, col = 1:8,
        pch = 16, cex = 3
        )
# Add text for colour
> text(x = rep(0.4,8),y = 10:17,
       paste("colour",1:8),pos = 4,
       offset = 1
      )
# Add lines
> segments(x0 = rep(0.7,20),y0 = 11:16,
           x1 = rep(0.9,20),lty = 1:6,
           lwd = 1.3,col = "black"
          )
# Add text to lines
> text(x = rep(0.9,20),y = 11:16,
       paste("lty",1:6),pos = 4,
       offset = 1
       )
# Add title text
> title_msg = "Exercise 7: reproduce & save this plot for future reference"
> mtext(title_msg,
        side = 3, outer = TRUE, line = -2.2:-2, font = 2
       )
```



Exercise 7: reproduce & save this plot for future reference



14. Generalised Linear Mixed Models (GLMM)

Taking the earlier prediction:

```
> owl = read.csv('owl_data.csv')
```

```
> owl.lm = lm(begging ~ sex + food + sex * food, data = owl)
```

The assumption was that the residuals were random and had no relationship with eachother. However it could be observed for example that there is a higher possibility of the residuals below the model line being say male in blue and above the line being female. Thus there is non-independence of the residuals and this needs to be accounted for. One way is to do separate ANCOVA for set, male and female.



Illustration 54: Non-independence of residuals

However this could get impractical quite quickly. Imagine if there were samples taken from a number of sources and for practical reasons there were more samples taken from certain sources than others. Now there is non-independence between the samples taken from each source and because of the different ratios of samples from each it must be accounted for in the linear models.

- **nlme:** Non-linear Mixed-Effects Models (NLME). This generic function fits a non-linear mixed-effects model in the formulation described in Lindstrom and Bates (1990) but allowing for nested random effects. The within-group errors are allowed to be correlated and/or have unequal variances.
- **nlme:** Fit Linear Mixed-Effects Models (LMM). Fit a LMM to data, via the Restricted (or Residual, or Reduced) Maximum Likelihood (REML).

Import the *owl_data.csv* file. There are four models. The first two are standard Linear Models. The second two models fit the same model as the *Im()* function, however they must have at least one random effect, in this case the *(1/nest)* argument. If you want to fit another type of distribution you use *glmer()* and set the family argument. It is demonstrated below for the family *poisson* however in this case it would fail to converge as the dataset is not suitable for that distribution.





Illustration 56: Plot - Fitting Linear Models

- > par(mfrow=c(2,2))
- > plot(mod.glm)



Illustration 57: Plot - Fitting Generalised Linear Models

```
> plot(mod.lme)
```



Illustration 58: Plot - Linear Mixed-Effects Models
```
> plot(mod.lmer)
```



Illustration 59: Plot - Fit Linear Mixed-Effects Models

15. Qualitative Data Analysis with R

15.1 Introduction

As has already been shown the *R* programming language is very powerful for quantitative analysis, but what or Qualitative analysis? *R* has a <u>R</u> <u>Qualitative Data</u> <u>Analysis (RQDA)</u> for qualitative text and PDF document analysis.

It is particularly useful for inductive thematic analysis however for deductive analysis it is necessary to upload Categories and Codes one by one. <u>RQDA Code Builder</u> resolves this.

This document demonstrates how to use *RQDA()* and the *RQDA Code Builder* on a GNU/Linux platform. *R* and *RQDA()* can be used on other platforms like Microsoft Windows and as the RQDA Code Builder is Python3 based it can easily be adapted for other platform implementations.

It is necessary to have *python3* installed on the platform. Use the *Software Manager* for your GNU/Linux flavour or install using *apt* from the shell terminal.

```
$ sudo apt install python3
$ sudo apt-get install python-yaml
```

Confirm the install and the version of python3.

\$ python3 --version
Python 3.5.2

15.2 Qualitative Content Analysis

Qualitative Content Analysis follows a procedure (Flick, 2014):

- 1. Deciding the research question
- 2. Selecting material
- 3. Building a coding frame
- 4. Segmentation
- 5. Trial coding
- 6. Evaluating and modifying the coding frame
- 7. Main analysis
- 8. Presenting and interpreting the findings.

15.3 Coding

Assuming that steps 1 and 2 are completed and the next step is the building of a coding frame. There are two approaches, inductive and deductive.

The inductive approach has codes extracted directly from the source data. As the researcher reads through each source file (interviews, papers, etc..), he or she highlights key lines and creates a code for it. These codes are added and modified as the researcher reads through all the source material. The codes are then bundled into codes of common category. RDQA() is very suitable for this approach.

The deductive approach involves the researcher developing codes and categories in advance, in a scheme. These codes are then applied to the source data. As RDQA()

expects codes to be added one by one through the graphical interface this is difficult. Application of the *rqda_code_builder.py* program described here helps to fix this.

15.4 Starting RQDA()

Create a directory as a parent for the project and open a command shell in it. Within the parent directory create a *Sources* directory. Place the source files in the *Sources* directory. In this example you can see two source files but typically this would be many files associated with interviews, observation logs, etc..

\$ mkdir Sources



Illustration 60: Sources directory

```
$ ls Sources
Colours_of_Health_and_Sickness_Sociocult.txt
Psychological_Properties_Of_Colours.txt
```

Run the 'R' program.

```
$ R --quiet
>
```

15.4.1 Add the RQDA library

Add the *RDQA() library*, this is the program that allows the researcher to analyse the data.

```
> library(RQDA)
Loading required package: RSQLite
Loading required package: gWidgetsRGtk2
Loading required package: RGtk2
Loading required package: gWidgets
Loading required package: cairoDevice
Loading required package: DBI
Use 'RQDA()' to start the programme.
```

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RQDA: Qualitative Data Analysis	
New Project	Project
Open Project	Files
Close Project	
Project Memo	Codes
Backup Project	Code
Save Project As	Categories
Clean Project	Cases
Close All Codings	Attributes
Path of current project: No project is open.	File Categories
Author: <ronggui.huang@gmail.com> Help: click to join rgda-help mailing list</ronggui.huang@gmail.com>	Journals
License: BSD Version: 0.3-0 Year: 2017	Settings
About	

The graphical tool starts.

Illustration 61: RQDA() GUI

15.4.2 Create a Project

In the Graphical User Interface (GUI):

- Click New Project.
- Enter a name in the desired path: *Colour_project.rqda*. Click *OK*.

A new project file appears in the directory.



Illustration 62: Project SQLite database

You may also notice in the *R* shell that the following command is executed.

> [1] "~/Colour_project.rqda"

Name the coder

Select the Settings tab and define the Name of Coder in the first box.

RQD/	A: Qualitative Data Analysis	- • ×
C	lick to set font	Project
-Settings Name of Coder	File Encoding	Files
JohnnyResearcher	unknown	
Color for Coding	Color for Case	Codes
blue	gold 🗸	Code
Current coding table	Byte Order Mark	Categories
coding	FALSE 🗸	Cases
Show File Property	Type of Retrieval	Attributes
FALSE	unconditional 🗸	File
		Categories
		journais
		Settings
	Default Of	ĸ

Illustration 63: RQDA() interface

15.4.3 Import source files to the project

The next step is to import source data. This can be achieved either through the GUI one by one, or in bulk using the *R* function *write.FileList()* in the *R* shell.

Using the GUI

To use the GUI, select:

- The *Files* tab followed by the *Import* button.
- Browse to each file in turn and select.

RQDA: Qualitative Data Analysis	. • ×
Import New Delete Open Memo rename Attribute	Project
Files	Files
	Codes
	Code Categories
	Cases
	Attributes
	File Categories
	Journals
	Settings

Illustration 64: RQDA() files

Using the R shell

An alternative mechanism is to use the *R* shell. This command using the *addFilesFromDir()* function selects the files in the *Sources* directory that match the pattern. In this case all files that end in the pattern *.txt*.

Execute the command:

```
> addFilesFromDir('Sources', pattern = "*.txt$")
```

If you now check the GUI by clicking the *Files* tab, you will notice that the files from the *Sources* directory have been imported. Alternatively use the *getFiles()* function in the *R* shell to confirm.

```
> getFiles()
[1] "Colours_of_Health_and_Sickness_Sociocult.txt"
[2] "Psychological_Properties_Of_Colours.txt"
attr(,"class")
[1] "RQDA.vector" "fileName"
```

15.5 Coding

15.5.1 Inductive approach

RQDA() is very suitable for the *inductive approach* however it takes significant time.

	RQDA: Qualitative Data Analysis	- • ×
	Import New Delete Open Memo rename Attribute	Project
	Selected.File.id.is.2	Filos
	Colours_of_Health_and_Sickness_Sociocult.txt	- Files
	Psychological_Properties_Of_Colours.txt	Codes
Psycholog	gical_Properties_Of_Colours.txt _ ×	Code Categories
Black Black is associated with evil, and mystery. Black is a mysterious co unknown (black holes). (blacklist, black humor, and authority; it is consi and prestigious color (bl black is the symbol of g Black gives the feeling of background diminishes make you look thinner. photography, you can u make the other colors st bright colors. Combined powerful colors – black g	power, elegance, formality, death, Nor associated with fear and the It usually has a negative connotation 'black death'). Black denotes strength idered to be a very formal, elegant, lack tie, black Mercedes). In heraldry, rief. of perspective and depth, but a black readability. A black suit or dress can When designing for a gallery of art or use a black or gray background to tand out. Black contrasts well with with red or orange – other very gives a very aggressive color scheme.	Cases Attributes File Categories Journals Settings

Illustration 65: RQDA() files 2

Select each document in turn from the *Files* tab, a popup appears with the text from the source file selected.

Black Black is associated with power, elegance, formality, death, evil, and mystery. Black is a mysterious color associated with fear and the unknown (black holes). It usually has a negative connotation (blacklist, black humor, 'black death'). Black denotes strength and authority; it is considered to be a very formal, elegant, and prestigious color (black tie, black Mercedes). In heraldry, black is the symbol of grief. Black gives the feeling of perspective and depth, but a black background diminishes readability. A black suit or dress can make you look thinner. When designing for a gallery of art or photography, you can use a black or gray background to make the other colors stand out. Black contrasts well with bright colors. Combined with red or orange – other very powerful colors – black gives a very aggressive color scheme.

Illustration 66: RQDA() Codes

On the main GUI click the *Codes* tab and as a line is read that requires coding select *Add* and create the code. For example, to add a code *Black*, click *Add*. Enter the new code in the box provided and click *OK*. With text highlighted, select the appropriate code, i.e. *Black* and click *Mark*.

RQDA: Qualitative Data Analysis	- • ×	Psychological_Properties_Of_Colours.txt -
Add Delete rename Memo Anno Coding Unmark Mark Selected.code.id.is.1_2.codings Black	Project Files Codes Code Categories Cases Attributes File	Black BlackBlack is associated with power, elegance, formality, death, evil, and mystery. BlackBlack is a mysterious color associated with fear and the unknown (black holes). It usually has a negative connotation (blacklist, black humor, 'black death'). Black denotes strength and authority; it is considered to be a very formal, elegant, and prestigious color (black tie, black Mercedes). In heraldry, black is the symbol of grief. BlackBlack gives the feeling of perspective and depth, but a black background diminishes readability. A black suit or dress
	Categories Journals Settings	can make you look thinner. When designing for a gallery of art or photography, you can use a black or gray background to make the other colors stand out. Black contrasts well with bright colors. Combined with red or orange – other very powerful colors – black gives a very aggressive color scheme.

Illustration 67: RQDA() Codes 2

As can be seen each line is tagged.

15.5.2 Deductive approach

Unfortunately there does not appear to be a mechanism to import codes into the *RQDA()* database in bulk. For the deductive approach a researcher may have tens or even hundreds of categories and codes, it could be necessary to bulk upload. Extract the files from the *RQDA-Code-Builder.tgz* archive file which will give all the files including the database from this example as well as the *rqda_code_builder.py*. Move this file to the parent directory of the *Sources* directory.



Illustration 68: RQDA() Code Builder

The RQDA Code Builder (*rqda_code_builder.py*) program resolves this.

YAML file

YAML Ain't Markup Language (YAML) is a human-readable data serialisation language that is commonly used for configuration files, but can be used in many applications where data is being stored or transmitted. It is an ideal format for mapping of categories and codes.

The example project demonstrates how to *deduct* the following code schema as a YAML file in the same directory:

```
$ cat RQDA_codes.yam1
 RQDA_codes.yaml
 Colour:
      'Red
   -
      'Green'
      'Yellow'
      'Grey'
      'Black'
      'White'
      'Black'
      'Blue'
      'Pink'
      'Brown'
      'Purple'
 Psychological Properties:
      'Physical'
      'Intellectual'
      'Emotional'
      'Balance'
      'Spiritual'
 Floral Metaphors:
      'Daisy'
      'Juicy'
      'Apple'
      'Berrv'
      'Flower'
      'Peach'
 Human Characteristics:
     'Divinity'
   -
      'Eternity'
      'Infinity'
   _
```

Executing the RQDA Code Builder

Before executing the RQDA Code Builder it is important to shut down the *RQDA()* application by clicking on the *X* in the top right corner and selecting *OK* to the *Really EXIT*? question.

	Confirm	
2	Really EXIT?	
	You can use RQDA() to start this program again.	
	Cancel OK	

Illustration 69: RQDA() Exit

The file that the *RQDA()* program uses to store data is an SQLite database. It is the file that was created at the beginning when the project was opened (*Colour_project.rqda*). The RQDA Code Builder reads the YAML formatted schema and uploads it to the database. It also creates a Structured Query Language (SQL) log of each SQL command it executes and more importantly develops a set of *R* commands that match text blocks to the codes. It has the following switches:

```
    -c|--coder [Name] - Define coder, must match that from RQDA() settings.
    -d|--database [DB] - Define path to SQLite3 database file.
    -y|--yaml [YAML] - Define path to YAML code file.
```

Execute the command, check it is version 1.4 or greater and execute with the relevant switches as demonstrated.

```
$ cat rqda_code_builder.py | grep '# Version' | awk {'print $4'}
1.4
$ ./rqda_code_builder.py -c JohnnyResearcher -d Colour_project.rqda -y RQDA_codes.yaml
RQDA Code Builder
Connecting to the SQLite3 database Colour_project.rqda.
Connected to the SQLite3 database Colour_project.rqda. Uploading..
Upload completed
.....
A full list of SDL commands executed can be seen in the 'RQDA_SQL.log' file.
You can restart the RQDA() library with the following command in the R shell:
> RQDA()
Restart RQDA() as instructed.
```

> RQDA()

Two new files are created, *RQDA_SQL.log* which is a log of the SQL commands executed on the database as well as *RQDA_R_search_cmds.R* which is a list of commands that will be executed in the *R* shell to apply the deductive codes to the source files.



Illustration 70: RQDA() Search commands and Log files

Applying the RQDA 'R' search commands

To apply the RQDA search commands execute the following command in the *R* shell. This bulk executes the commands in the *RQDA_R_search_cmds.R* file on all the source files.

> source('RQDA_R_search_cmds.R')

15.6 Reviewing Coding

Before diving into the coding within the various source files, review the coding statistics. It can be seen from this extract that 385 code blocks were applied to the source texts.

```
> getCodingTable()
    rowid cid fid
                       codename
                                                                        filename
                                      Psychological_Properties_Of_Colours.txt
1
        4
            1
                2
                       Physical
2
        5
            1
                 2
                       Physical
                                      Psychological_Properties_Of_Colours.txt
                                      Psychological_Properties_Of_Colours.txt
                       Physical
3
        6
                 2
            1
4
        7
            1
                 2
                       Physical
                                      Psychological_Properties_Of_Colours.txt
5
        8
                 2
                       Physical
                                      Psychological_Properties_Of_Colours.txt
            1
                 2
                       Physical
                                      Psychological_Properties_Of_Colours.txt
6
        9
            1
                      . . .
                                 . . .
. . .
      . . .
            . .
                 .
                       Eternity Colours_of_Health_and_Sickness_Sociocult.txt
382
      385
           24
                 1
383
      386
           24
                 1
                       Eternity Colours_of_Health_and_Sickness_Sociocult.txt
384
      387
           25
                 1
                       Infinity Colours_of_Health_and_Sickness_Sociocult.txt
385
      388
           25
                1
                       Infinity Colours_of_Health_and_Sickness_Sociocult.txt
    index1 index2 CodingLength
                             135
1
       399
              534
2
      4109
             4197
                             88
                             103
3
      4739
             4842
4
       848
              977
                             129
5
      1454
              1587
                             133
6
      4257
             4394
                             137
      . . .
. . .
               . . .
                             . . .
382
      3189
             3301
                             112
383
     15902
            16020
                             118
384
      3189
             3301
                             112
385 15902 16020
                             118
```

15.7 Reviewing the coded blocks

Selecting the *Codes* tab from the *RQDA()* GUI and select any particular code. In this case *Brown* was selected. Click on the *Coding* button and a popup appears with each instance of sentences within the source files where the word *Brown* or *brown* appeared, such sentences were tagged with the *Brown* tag. The popup also shows for each block the source file from where the sentence appeared.



Illustration 71: Reviewing the coded blocks

This performs an initial *deductive coding*. There may be quirks however, what if one interviewee kept referring to *Beige* but the researcher wanted to code it as *Brown*? or the researcher has a code *Colour* and some of the transcripts were transcribed in American English. In this case sentences with *Color* should be coded with *Colour*.

Carry out additional coding of sentences like this.

First find the *CID* of the *Code* for *Brown*. Select the *Codes* tab, click on the *Brown* code and its *CID* can be seen at the top of the pane as shown by the red circle in Illustration 72.



Illustration 72: Find the CID of a code

Execute the following two lines in the *R* shell and they will be added to the main coding already performed.

```
> codingBySearch("Beige",fid=getFileIds(),cid=21,seperator="[.!?]")
```

```
> codingBySearch("beige",fid=getFileIds(),cid=21,seperator="[.!?]")
```

15.8 Visualising categories

There are some tools built into *RQDA()* for visualisation. For example using the *D3.js JavaScript library* for manipulating data. *D3* helps bring data to life visually using Hypertext Markup Language (HTML), Scalable Vector Graphics (SVG), and Cascading Style Sheets (CSS).

15.8.1 Installing d3Network

On the *R* Shell install D3.js and activate the *d3Network* within *R*.

- > install.packages('d3Network')
- > library(d3Network)

	RQDA: Qualitative Data Analysis	. • ×
Add De	ete rename Memo	Project
Add To Drop	From Unmark Mark	Files
Selected.catego	ry.id.is.3	
Colour	Add New Code to Selected Category	Codes
Floral Metapho Human Charac	Codings of selected category Memo Blot Selected Code Categories	Code Categories
Psychological P	Plot Selected Code Categories with d3	Cases
L 1	Soreby created and	Attributes
Codes.of.This.C	ategory	File
Black		Categories
Black		Journals
Blue		
Brown		Settings
Green		
Grey	-	J

15.8.2 Visualising Categories

Illustration 73: Plot selected code categories with d3

Select the *Categories* tab, highlight a *Category* or many *Categories* using the *ctrl* button and right click. Scroll down to the *Plot selected code categories with d3*. A HTML page will pop-up with diagrams like these:



Illustration 74: d3 plot

15.9 Summary

There are a lot more features to *R* and *RQDA()* that can aid qualitative research. The additional *RQDA Code Builder* program (*rqda_code_builder.py*) allows the researcher to deductively pre-build a code schema and apply it automatically.

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